

ORIGINAL ARTICLE

Relaxed selective constraints drove functional modifications in peripheral photoreception of the cavefish *P. andruzzii* and provide insight into the time of cave colonization

L Calderoni¹, O Rota-Stabelli², E Frigato¹, A Panziera^{1,3}, S Kirchner⁴, NS Foulkes⁵, L Kruckenhauser⁴, C Bertolucci¹ and S Fuselli¹

The genetic basis of phenotypic changes in extreme environments is a key but rather unexplored topic in animal evolution. Here we provide an exemplar case of evolution by relaxed selection in the Somalian cavefish *Phreatichthys andruzzii* that has evolved in the complete absence of light for at least 2.8 million years. This has resulted in extreme degenerative phenotypes, including complete eye loss and partial degeneration of the circadian clock. We have investigated the molecular evolution of the nonvisual photoreceptor melanopsin *opn4m2*, whose mutation contributes to the inability of peripheral clocks to respond to light. Our intra- and inter-species analyses suggest that the 'blind' clock in *P. andruzzii* evolved because of the loss of selective constraints on a trait that was no longer adaptive. Based on this change in selective regime, we estimate that the functional constraint on cavefish *opn4m2* was relaxed at ~5.3 Myr. This implies a long subterranean history, about half in complete isolation from the surface. The visual photoreceptor rhodopsin, expressed in the brain and implicated in photophobic behavior, shows similar evolutionary patterns, suggesting that extreme isolation in darkness led to a general weakening of evolutionary constraints on light-responsive mechanisms. Conversely, the same genes are still conserved in *Garra barreimiae*, a cavefish from Oman, that independently and more recently colonized subterranean waters and evolved troglomorphic traits. Our results contribute substantially to the open debate on the genetic bases of regressive evolution.

Heredity (2016) **117**, 383–392; doi:10.1038/hdy.2016.59; published online 3 August 2016

INTRODUCTION

Phreatichthys andruzzii is a cavefish that inhabits the subterranean waters of central Somalia and shows an extreme level of adaptation to the dark environment. The adult fishes completely lack eyes, pigments and scales and their metabolic rate is extremely reduced. In particular, the eye is totally regressed 36 h post fertilization and 1 month later there remains only a rudimentary cyst, with loss of the optic nerves and strongly reduced optic lobes (Berti *et al.*, 2001; Stemmer *et al.*, 2015). This degeneration is rather premature compared, for example, with what is known for the blind Mexican cavefish *Astyanax mexicanus*. In this Mexican species, small eye primordia develop during embryogenesis, grow for a while, and only later arrest in development, degenerate and sink into the orbits (Wilkens and Strecker, 2003; Jeffery, 2009). Other important biological peculiarities of *P. andruzzii* connected to the absence of light involve the circadian clock, a physiological timing mechanism that allows organisms to anticipate and adapt to the day–night cycle. Our previous studies showed significant regression of this mechanism in terms of light sensitivity. When exposed to cycles of light and dark, the cavefish lacks behavioral rhythmicity, and the expression of a set of clock-regulated and light-regulated genes is also arrhythmic. Interestingly, the clock of

P. andruzzii does not respond to light, but is still sensitive to other timing signals, such as food availability that are possibly more important in the dark environment (Cavallari *et al.*, 2011). The regression of the light-related pathway is extended to the mechanism of light perception, in particular to some nonvisual photoreceptors, melanopsin (*opn4m2*) and teleost multiple tissue opsin. In *P. andruzzii*, loss-of-function (LOF) mutations in the genes coding for these peripheral photoreceptors abolish their ability to respond to light. Consequently, the downstream cascade in the light-responsive pathway is not induced, and the peripheral circadian clocks are light insensitive (Cavallari *et al.*, 2011).

The extreme troglomorphy developed by *P. andruzzii* in the absence of light is probably the result of both complete and long-term isolation. Although the water table where these fish live is very superficial (from some meters to tens of centimeters), for the sake of simplicity we will refer to it as 'the cave'. *P. andruzzii* cave populations probably originated from normal-eyed ancestors that lived in surface water when Somalia had a wetter climate. As a consequence of increases in North East African aridity after 2.8 Myr that led to the desertification at the surface (deMenocal, 1995), *P. andruzzii* ancestors that colonized the cave environment remained isolated in the dark of

¹Department of Life Sciences and Biotechnology, University of Ferrara, Ferrara, Italy; ²Research and Innovation Centre, Fondazione Edmund Mach, San Michele all'Adige, Italy;

³Department of Biodiversity and Molecular Ecology, Fondazione Edmund Mach, San Michele all'Adige, Italy; ⁴Central Research Laboratories, Museum of Natural History of Vienna, Vienna, Austria and ⁵Institute of Toxicology and Genetics, Karlsruhe Institute of Technology, Karlsruhe, Germany

Correspondence: Dr S Fuselli, Department of Life Sciences and Biotechnology, University of Ferrara, Via L Borsari 46, Ferrara 44121, Italy.

E-mail: fss@unife.it

Received 26 November 2015; revised 18 April 2016; accepted 25 April 2016; published online 3 August 2016

the phreatic waters. This complete isolation is because of the disappearance of surface populations, unlike some other species that maintain some level of gene flow with surface-dwelling populations (Li and He, 2009; Kruckenhauser *et al.*, 2011). The extent to which gene flow occurs between surface- and cave-dwelling forms is still debated for *A. mexicanus*, the model species in cavefish evolution (Bradic *et al.*, 2012; Gross, 2012; Strecker *et al.*, 2012). Another key factor that determines the extent of regression is time. Mutations that render the gene nonfunctional may require a certain time to enter the population ($1/(2N\mu)$ generations on the average if $N\mu$ is small), and, if they do get fixed, the time will be $4N$ generations. Thus, the integrity of a gene, and its ability to code for a functional protein, may persist for a long period purely because of chance alone, unless selection favors degeneration. In this case the fixation time depends inversely on the selective coefficient and directly on the logarithm of the population size (Kimura and Ohta, 1969). The period of time that the Somalian caves have been inhabited by populations of *P. andruzzii* is unknown, although the desertification of the surface environment, which occurred at ~ 2.8 Myr, gives us an approximate lower limit for the colonization event. Adaptation to constant darkness probably started before the complete isolation of the Somalian cavefish, although the time required to reach this extreme level of troglomorphy is still unknown. Inferences on the pattern of molecular evolution of proteins involved in the development of troglomorphic traits can be exploited to reconstruct the evolutionary history of *P. andruzzii*. For example, environmental change such as the colonization of a new environment often eliminates or weakens a source of selection that was formerly important for the maintenance of a particular trait but then subsequently lost its adaptive value (Lahti *et al.*, 2009). Proteins contributing to determining the trait start to evolve neutrally, accumulating amino acid substitutions and LOF mutations that will eventually impair their function (Motoo, 1983). An alternative hypothesis is that losing the trait may directly or indirectly increase the fitness of an organism in the new environment (Espinasa and Espinasa, 2008). In this case, after a LOF mutation gets fixed, the gene loses its ability to code for a functional protein and its neutral evolution begins. Opposing hypotheses invoking neutral mutation or natural selection have been advanced to explain regressive evolution in subterranean animals, and *Astyanax* cavefish, in particular (Poulson and White, 1969; Wilkens, 1988; Culver and Wilkens, 2000; Jeffery, 2005, 2009; Wilkens, 2010).

If new environmental conditions mean relaxation of natural selection on a specific trait, the time when the trait was freed of constraint can be put in relation with the time since colonization of the new habitat (Wang *et al.*, 2004; Zhao *et al.*, 2010). In the case of *P. andruzzii*, characterizing the molecular evolution of a mutated

photoreceptor gene and estimating when selection became relaxed would provide insight into the time of cave colonization. This will in turn shed light on the mechanisms underlying regressive evolution in this species.

With this aim, we first tested the hypothesis that the lack of response to light of the circadian clock of *P. andruzzii* observed by Cavallari *et al.* (2011) results from a change in selective regime on the light-responsive mechanism, as expected when the phenotypic trait is no longer adaptive. In particular, we investigated the molecular evolution of the nonvisual photoreceptor melanopsin *opn4m2*, whose premature stop codon accounts at least in part for the inability of peripheral clocks to respond to light. To address the question of whether other light-related mechanisms are also undergoing degeneration, we also tested for a change in the rate of evolution on the visual receptor rhodopsin (*rho*) that is expressed in the brain of *P. andruzzii*, is still functional and is probably involved in photophobic behavior (Tarttelin *et al.*, 2012). These gene sequences were also analyzed in another blind cavefish, *Garra barreimiae*, from Oman that much more recently colonized subterranean waters and evolved troglomorphic traits (Kruckenhauser *et al.*, 2011). The two cavefish show an extremely similar osteology (Banister, 1984), and a recent phylogenetic study proposed that *Phreatichthys* should be treated as a junior synonym of *Garra* (Yang *et al.*, 2012). Similar to *P. andruzzii*, the eyes of *G. barreimiae* are submitted to ontogenetic regression. Although the juveniles develop eyes, they are completely reduced during growth to the adult stage, and hence the adults have no externally visible eyes, and the optic lobes are not fully developed (Banister, 1984). The time of divergence between the two cavefish species was estimated using a fossil-calibrated phylogeny of mitochondrial *cytochrome b* (*cytb*) sequences of 148 teleost species. This divergence time and the results of our selection analyses were used to date when selection became relaxed on *opn4m2*. Based on our results we conclude that the change in selective regime that followed the colonization of the new environment predates the desertification of Somalia by ~ 2.5 million years. This indicates that the complete isolation of *P. andruzzii* may have occurred several million years after cave colonization.

MATERIALS AND METHODS

Cavefish samples

The *P. andruzzii* and *G. barreimiae* individuals typed in this study are described in Table 1 (Berti and Messana, 2010; Kruckenhauser *et al.*, 2011).

Molecular techniques

The genomic DNA of two specimens from two different sampling sites of *P. andruzzii* (Bud Bud and Gheriale) and of five specimens from three sampling locations of *G. barreimiae* was extracted using DNAeasy blood and tissue kit

Table 1 Description of *Phreatichthys andruzzii* and *Garra barreimiae* individuals analyzed in this study with sampling localities

Species	Individual IDs	Locality	Latitude/longitude	Phenotypes
<i>P. andruzzii</i> ^a	PA07 Cavefish	Bud Bud (Somalia)	04°11'19"N/46°28'27"E	Troglomorphic
	PA11 Cavefish	Gheriale (Somalia)	04°08'22"N/46°29'05"E	Troglomorphic
<i>G. barreimiae</i> ^b	Gbar215	Wadi Al Falahi (Oman)	23°04'50"N/57°21'14"E	Surface
	Gbar216	Wadi Al Falahi (Oman)	23°04'50"N/57°21'14"E	Surface
	Gbar223 Cavefish	Hoti Pit (Oman)	23°06'10"N/57°22'12"E	Troglomorphic
	Gbar224 Cavefish	Hoti Pit (Oman)	23°06'10"N/57°22'12"E	Troglomorphic
	Gbar284	Wadi Misfat (Oman)	23°08'52"N/57°18'57"E	Surface

^aBerti and Messana (2010).

^bHoti Pit is a cave adjacent to the Al Hoota cave described in Kruckenhauser *et al.* (2011).

(Qiagen, Venlo, The Netherlands) following the manufacturer's protocol. Part of the *opn4m2* gene (1187 bp spanning nucleotides from 40 to 1226, premature stop codon position: nucleotide 886, CDS length: 1503 in zebrafish ENSDARG0000007553) and the entire *rhodopsin* (*rho*) gene were amplified and Sanger sequenced in each of the 7 individuals (Supplementary Methods).

To estimate the divergence time between *P. andruzzii* and *G. barreimiae*, the mitochondrial *cytb* gene was amplified and sequenced in both species with primers Glu32-fwd 5'-GACTTGAAGAACCACCGTTGT-3' and Thr1-rev 5'-GATCTTCGGATTACAAGACCG-3' (Supplementary Methods). A fragment of 903 bp of *P. andruzzii* from the ATG start codon was aligned with the entire coding region of *G. barreimiae cytb* and with previously published *cytb* sequences available in GenBank (see the section 'Analysis of divergence time between *P. andruzzii* and *G. barreimiae*') and used for analysis.

For the three genes, the coding sequences were translated into their respective protein sequences and aligned using ClustalW available in MEGA version 6 (Tamura *et al.*, 2013) with default parameters. Once aligned, the protein sequences were translated back to their respective nucleotide sequences that were used for the following analyses.

Estimation of the functional impact of nonsynonymous substitutions

For both the *opn4m2* and *rho* genes, we identified amino acids at a specific position in only one of the five cyprinid lineages of the tree shown in Figures 1a and b (*Phreatichthys andruzzii*, *Garra barreimiae*, *Danio rerio*, *Cyprinus carpio* and *Rutilus rutilus* or *Carassius auratus*, accession numbers are listed in Supplementary Table 1). We considered those amino acids to be the result

of a nonsynonymous substitution that, at present, characterizes only that specific branch (hereafter 'species-specific'). As the aim of the analysis was to compare the effect of fixed nonsynonymous substitutions that differ among species, in the case of *P. andruzzii* and *G. barreimiae* polymorphic nonsynonymous substitutions were not considered. We predicted the functional impact of each species-specific amino acid change using two different approaches, both based on the degree of conservation of specific residues in evolutionary related proteins.

First, we used the Coding SNP Analysis tool (Thomas *et al.*, 2006) based on the PANTHER database (<http://pantherdb.org/>) that calculates a score called 'substitution position-specific evolutionary conservation' (subPSEC). This score corresponds to the negative logarithm of the probability ratio of the wild-type and mutant amino acids at a particular position. PANTHER subPSEC scores are converted to a probability of deleterious effect on protein function; a subPSEC of -3 is the cutoff point for functional significance and corresponds to a $P_{\text{deleterious}}$ of 0.5 (50% probability that the substitution is deleterious).

Second, we applied the SIFT sequence prediction tool, interrogating three different databases: UniRef90, UniProt-SwissProt and UniProt-TREMBL (<http://sift.jcvi.org/>) (Kumar *et al.*, 2009). Probability values of ≤ 0.05 indicate that the substitution would not be phenotypically tolerated (deleterious).

Overall, the substitutions were considered 'deleterious' if three tests out of four predicted a deleterious effect, 'mild' if two out of four predicted a deleterious effect and, otherwise 'tolerated'. We tested whether the *P. andruzzii* or *G. barreimiae* observed proportion of deleterious and tolerated/mild substitutions differ significantly from those of the other species by means of a χ^2 test of independence with Yates' correction.

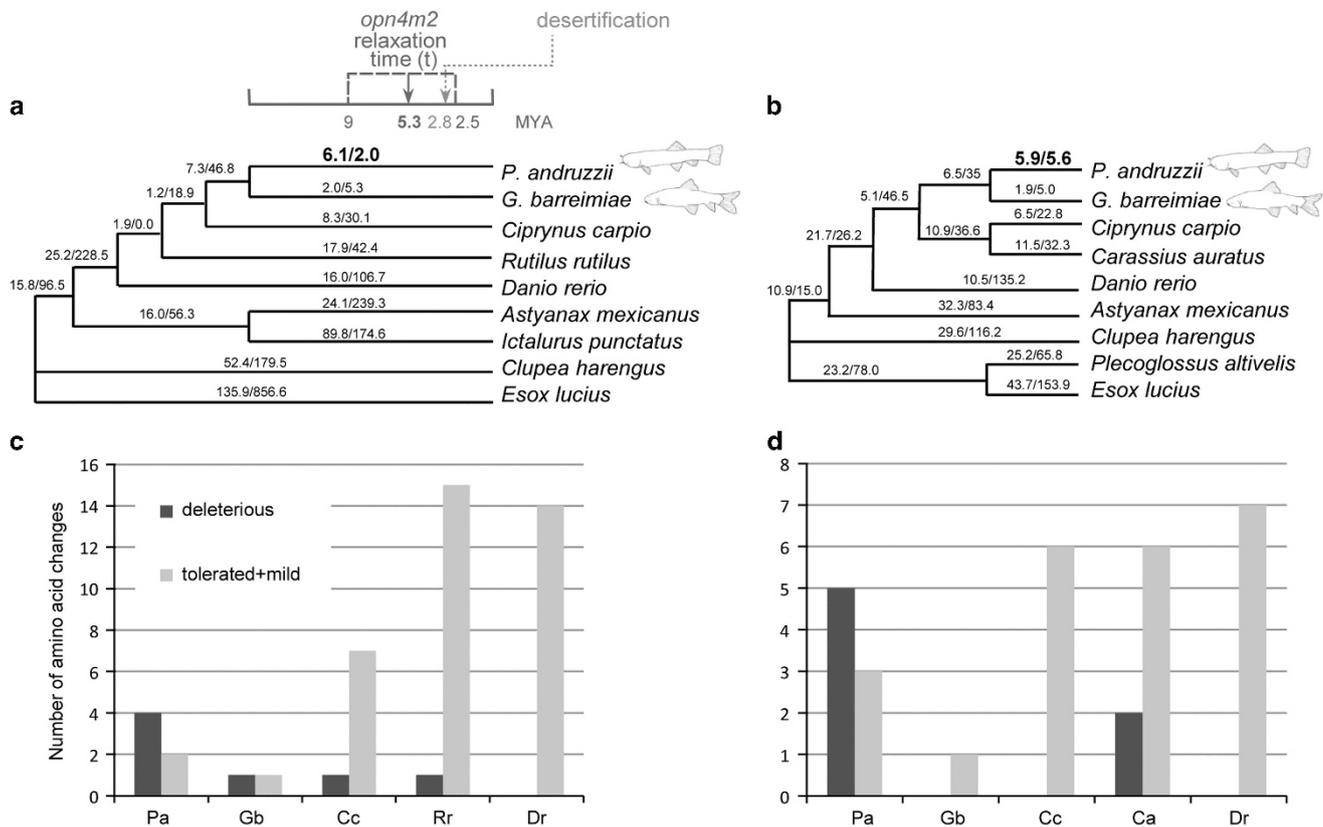


Figure 1 The two species trees used for CODEML analyses of (a) *melanopsin* (*opn4m2*) and (b) *rhodopsin* (*rho*) genes. Branch lengths are not drawn to scale. The two numbers shown along each branch are the maximum likelihood estimates of the number of nonsynonymous and synonymous substitutions (N*dN and S*dS) along that branch (CODEML, free ratio model). Dates indicated with the dark gray arrow and dashed lines are inferred from the molecular genetic evidence of this study (*opn4m2* relaxation time), whereas for the shift in climatic condition (desertification) we referred to deMenocal (1995). Melanopsin (c) and rhodopsin (d) species-specific amino acid changes are more likely to be deleterious in *P. andruzzii* (Pa) compared with changes in other cyprinid species (Ca, *Carassius auratus*; Cc, *Cyprinus carpio*; Dr, *Danio rerio*; Gb, *Garra barreimiae*; Pa, *Phreatichthys andruzzii*; Rr, *Rutilus rutilus*).

Detecting selection (and selection relaxation) acting on particular lineages

To analyze the selective constraints on *opn4m2* and *rho*, we estimated the nonsynonymous/synonymous rate ratio ($\omega = d(N)/d(S)$), under different evolutionary models in a likelihood framework. This ratio is <1 under purifying selection, approaches 1 under neutral rates of evolution and is >1 under positive selection. For both genes, we analyzed the coding sequence of nine teleost fishes (Figures 1a and b), including those of *P. andruzzii* (showing a premature stop codon at nucleotide 886 of the *opn4m2* sequence) and *G. barreimiae* (uninterrupted open reading frame) from this study. In particular, we used the sequence of *P. andruzzii* from Bud Bud (PA07), and of *G. barreimiae* from the Hoti Pit cave (Gbar223 and Gbar224 are identical and homozygotes at both genes, see Supplementary Table 2). Orthologous sequences were retrieved using BLAST protein: we considered melanopsin protein sequences with query cover $>90\%$ and excluded isoforms and low-quality proteins (default algorithm parameters, database: non-redundant protein sequences, organism: Teleostei). As the melanopsin of *P. andruzzii* is truncated, the known orthologous sequence of zebrafish was used as a query. The respective coding sequences were obtained from GenBank and aligned using the MAFFT online version with default settings (Katoh and Standley, 2013). An alignment of rhodopsin coding sequences was obtained with the same procedure, including, when available, the same species of the melanopsin alignment (see Supplementary Table 1 for Ensembl or GenBank IDs). Codons showing indels were not considered in the analyses.

Maximum likelihood trees for the coding sequences of melanopsin and rhodopsin were estimated using the program RAXML 8.1.2 (Stamatakis, 2014) and the GTR+gamma substitution model. The inferred topologies are shown in Figures 1a and b and were used as input (unrooted tree) for the following tests for selection. Three different approaches were used: Branch models (Yang, 1998), Clade models (Bielawski and Yang, 2004; Weadick and Chang, 2012) and Branch-site models (Zhang *et al.*, 2005). Codon frequencies were approximated using the F3x4 calculation. Each model was fit to the data multiple times from different starting ω or k (the transition/transversion rate ratio) values to avoid local optima.

In particular, we considered the next Branch models: the simplest model assumes the same ratio for all branches in the phylogeny ('one-ratio' model); a second and nested model assumes that the branches of interest (in our case *P. andruzzii* or *G. barreimiae*) have a dN/dS ratio (ω_{Pa} or ω_{Gb}) that is different

from the background ratio ω_0 ('two-ratio model', both ω estimated). We also considered a two-ratio model where ω was fixed at 1 in the lineage of *P. andruzzii* or *G. barreimiae*, as we are specifically interested in testing for relaxed molecular evolution in dark conditions. Finally, the 'free-ratio' model that assumes an independent dN/dS ratio for each branch in the phylogeny was used to calculate branch-specific rate of variation. The analyses were performed using the CODEML module of PAML package (Yang, 2007).

Although Branch models assume that ω varies across branches of the phylogeny, but that it is invariant across sites, Clade models and Branch-site models incorporate among-site variation in selective constraint. We tested for the signature of divergent selective constraint at specific sites between the foreground branch (either *P. andruzzii* or *G. barreimiae*) and the background branches (the rest of the phylogeny) using the Clade model C (CmC). This model assumes three site classes, Class 0 and Class 1 corresponding to codons that evolve under purifying selection ($0 < \omega_0 < 1$) and neutral evolution ($\omega_1 = 1$), respectively. These two site classes are assumed to be shared between foreground and background branches, whereas the selection pressure at the third class can differ between the background and the foreground ($\omega_2 \neq \omega_3$). M2a_rel (Weadick and Chang, 2012) assumes three classes of sites, $0 < \omega_0 < 1$, $\omega_1 = 1$, and $\omega_2 > 0$, shared by background and foreground branches, and was used as a null model to test for the presence of divergently selected sites. A Bayes empirical Bayes approach was used to identify specific codons with high posterior probability of being in the divergent class (Yang *et al.*, 2005).

Clade models and Branch-site models similarly assume different classes of sites (three and four, respectively), the first two ($0 < \omega_0 < 1$ and $\omega_1 = 1$) shared between the background and the foreground. However, in Branch-site models the final two classes correspond to sites that are under purifying selection or neutral evolution in the background branches, but positive selection ($\omega_2 > 1$) in the foreground branches. The null model to test for the presence of positively selected sites assumes $\omega_2 = 1$.

The likelihoods of different models were then compared by means of likelihood ratio tests (LRTs) of hypotheses through comparison of nested statistical models. The LRT was calculated as twice the difference in the log-likelihoods values between models. The significance of the LRT statistic was determined by using the χ^2 distribution where the degrees of freedom equal the difference between the number of estimated parameters in the two nested models (Tables 2 and 3).

Table 2 Log-likelihood values (lnL), parameter estimates and likelihood ratio tests for various Branch-based models testing for heterogeneous selection pressures for melanopsin and rhodopsin

Branch model	ω_0	ω_{Pa}	ω_{Gb}	lnL ^a	np ^b	H ₀	H ₁	2ΔlnL ^c	d.f.	P-value
<i>Melanopsin (opn4m2)</i>										
A. One ratio: all branches have the same ω	0.08	ω_0	ω_0	-5800.498	17					
B. One ratio: all branches have the same $\omega = 1$	1	ω_0	ω_0	-6407.852	16	B	<u>A</u>	1214.708	1	3.88E-266
C. Two ratio: (ω_0), ω_{Pa}	0.077	1.189	ω_0	-5794.580	18	A	<u>C</u>	11.836	1	0.0006
D. Two ratio: (ω_0), $\omega_{Pa} = 1$	0.077	1	ω_0	-5794.597	17	<u>D</u>	C	0.033	1	0.856
E. Two ratio: (ω_0), ω_{Gb}	0.079	ω_0	0.167	-5800.181	18	<u>A</u>	E	0.634	1	0.426
F. Free ratio (each branch has one ω)	Variable by branch ^d	1.127	0.137	-5777.164	31	C	<u>F</u>	34.832	13	0.001
<i>Rhodopsin (rho)</i>										
A. One ratio: all branches have the same ω	0.055	ω_0	ω_0	-4379.978	17					
B. One ratio: all branches have the same $\omega = 1$	1	ω_0	ω_0	-4971.150	16	B	<u>A</u>	1182.344	1	4.19E-259
C. Two ratio (ω_0), ω_{Pa}	0.053	0.209	ω_0	-4378.015	18	A	<u>C</u>	3.926	1	0.048
D. Two ratio: (ω_0), $\omega_{Pa} = 1$	0.053	1	ω_0	-4379.183	17	<u>D</u>	C	2.337	1	0.126
E. Two ratio: (ω_0), ω_{Gb}	0.054	ω_0	0.110	-4379.770	18	<u>A</u>	E	0.414	1	0.520
F. Free ratio (each branch has one ω)	Variable by branch ^d	0.199	0.072	-4365.531	31	C	<u>F</u>	24.968	13	0.023

Abbreviations: Gb, *Garra barreimiae*; H₀ and H₁, null and alternative hypothesis; Pa, *Phreatichthys andruzzii*. The best-fit model is underlined.

^aln(likelihood) value.

^bNumber of parameters.

^cTwice the difference of ln(likelihood) between the two models compared.

^d(N*dN) and (S*dS) values are shown in Figures 1a and b.

Analysis of divergence time between *P. andruzzii* and *G. barreimiae*

Divergence time between *P. andruzzii* and *G. barreimiae* was estimated using the Bayesian implementation Phylobayes 3.3f (Lartillot *et al.*, 2009). We used a dating strategy involving multiple calibration points based on 20 fossil constraints obtained from the literature (Near *et al.*, 2012; Wang *et al.*, 2012; Supplementary Table 3) coupled with a root prior for the origin of Neopterygii of 360 Myr and a permissive s.d. of 20 Myr (Near *et al.*, 2012). The data set included the first 903 nucleotide positions of the *cytb* gene for 148 Teleost species (26 of which were cyprinids, including the specimens of *P. andruzzii* and *G. barreimiae* that were sequenced in this work) and an Amiid (*Amia calva*) as outgroup (Supplementary Tables 3 and 4). Details of the analysis are described in the Supplementary Methods.

Dating the relaxation of selection on the *opn4m2* gene in *P. andruzzii*

To estimate when the functional constraint on the *P. andruzzii opn4m2* gene became relaxed, we followed a method described in Zhao *et al.* (2010), based on the rate of nucleotide substitution across the tree of Figure 1a. According to this method, we assumed that the level of selective constraint observed in the other branches of the phylogeny became suddenly and completely relaxed in the *P. andruzzii* branch *t* Myr. Time of relaxation of selection *t* was estimated as:

$$t = \frac{\left[\frac{d_{pa}}{d_0} \left(\frac{D_0}{D_{Gb}} \right) - 1 \right] T}{1/\omega - 1}$$

where d_{pa} is the total number of nonsynonymous substitutions per site for the *P. andruzzii* branch, d_0 is the total number of nonsynonymous substitutions per site for all the branches of the phylogeny except the *P. andruzzii* branch, D_0 is the total number of synonymous and nonsynonymous substitutions for the same branches and D_{Gb} is the total number of synonymous and nonsynonymous substitutions for the *G. barreimiae* branch. d_{pa} , d_0 , D_0 and D_{Gb} were obtained running a free-ratio model in PAML that assumes a different dN/dS ratio for each branch in the tree. The ω is the dN/dS ratio before the functional relaxation, that is, the background ratio in a two-ratio Branch model of PAML where *P. andruzzii* is set as foreground (Branch model C, Table 2). *T* is the estimated divergence time between *P. andruzzii* and *G. barreimiae*. Specifically, we estimated the divergence time between the two cavefish at 15.1 Myr, with a 95% support interval between 7.4 and 26.7 Myr. This age is the average of the point estimates obtained in the three Phylobayes analyses, and the support interval is conservatively computed using the lowest and the highest limits of the three 95% highest posterior density intervals, jointly considered (Supplementary Table 5; Rota-Stabelli *et al.*, 2013). Random sampling from a normal approximation of the posterior distribution of *T* produced by Phylobayes was used to estimate the uncertainty of *t*.

RESULTS**Genetic variation of *opn4m2* and *rho* within and between *P. andruzzii* and *G. barreimiae***

In *P. andruzzii* the deletion at position 815 (codon 272) of *opn4m2* that causes a frameshift leading to a premature stop codon 24 amino acids downstream (Cavallari *et al.*, 2011) was observed in each of the four chromosomes typed in this study. Within *P. andruzzii* we observed three polymorphic sites, all of them synonymous (Supplementary Table 2). The *opn4m2* gene showed four polymorphic positions in surface and cave individuals of *G. barreimiae* (six and four chromosomes, respectively), two of which are nonsynonymous and carried by surface individuals (Supplementary Table 2). Conversely, the two individuals from the cave are identical and homozygous at this locus. Comparing the two species, we observed seven synonymous and eight nonsynonymous fixed differences.

Within *P. andruzzii* the *rho* gene shows two synonymous and four nonsynonymous polymorphic positions, whereas no variation was identified within *G. barreimiae*. Comparing the two species, we observed eight synonymous and eight nonsynonymous fixed differences. When the *rho* gene was first characterized in *P. andruzzii*, an

in-frame insertion of 3 bp was identified in comparison with zebrafish, with the cavefish showing an additional valine at amino acid position 349 (Tarttelin *et al.*, 2012). As the total length of the protein is 355 amino acids, the valine insertion might not have major phenotypic consequences. In addition, this protein region is rather repetitive, and this may have caused recurrent insertion and deletion, as also supported by the lack of three amino acids in *G. barreimiae* compared with *P. andruzzii* (or two in comparison with zebrafish) (Supplementary Table 2).

Predictions of the functional impact of nonsynonymous mutations

We predicted the effects of the species-specific amino acid changes for the five species of cyprinids shown in Figures 1a and b for both the *opn4m2* and *rho* genes. We observed a significant excess of nonsynonymous substitutions predicted to be deleterious in the *opn4m2* gene of *P. andruzzii* compared with the other cyprinid species (χ^2 test with Yates' correction $P < 0.005$) (Figures 1c, d and 2 and Supplementary Table 6). One of the six amino acid changes was located downstream of the premature stop codon (K343T, deleterious). Conversely, the melanopsin of *G. barreimiae* showed only two species-specific amino acid changes, one mild and one deleterious, and the six amino acid changes shared by the two cavefish are almost completely tolerated. These results are expected given that *P. andruzzii* melanopsin is truncated and its photoreceptor activity is abolished (Cavallari *et al.*, 2011), thus purifying selection should no longer preserve the protein structure. However, the significant excess of nonsynonymous substitutions predicted to be deleterious in *P. andruzzii* compared with the other cyprinid species is less expected for the functional rhodopsin (χ^2 test with Yates's correction $P < 0.005$) (Tarttelin *et al.*, 2012). Conversely, *G. barreimiae* showed only one species-specific amino acid substitution, and this change showed a predicted tolerated effect (Figures 1c, d and 2 and Supplementary Table 6).

Estimation of synonymous and nonsynonymous substitution rates and detection of natural selection on cavefish lineages

We used maximum likelihood estimates of parameters for each branch of the trees shown in Figures 1a and b under the CODEML free-ratio model to calculate the number of nonsynonymous and synonymous substitutions per lineage. The *P. andruzzii* branch shows a higher number of estimated nonsynonymous substitutions compared with synonymous ones. This unique pattern may be the result of a different relative rate of evolution (neutral or adaptive) of this cavefish lineage relative to the rest of the tree. To formally test this hypothesis, we compared a series of evolutionary models in the likelihood framework. The results obtained from Branch model analyses are summarized in Table 2. We found that the average ω across the *opn4m2* tree is 0.08 and differs significantly from 1 (model A vs B, LRT, $P < 10^{-60}$), as expected if purifying selection is in general acting on the protein. A two-ratio model with *P. andruzzii* as the foreground branch (estimated $\omega_{pa} = 1.189$) compared with the rest of the tree (estimated $\omega_0 = 0.077$) fits the data better than the one-ratio (model C vs A, LRT, $P < 10^{-3}$). This indicates neutral evolution of the CDS, as the neutral model with $\omega_{pa} = 1$ fitted the data better than the previous (model C vs D, LRT, $P = 0.856$), suggesting an evident relaxation of natural selection on this protein. In the other blind cavefish, *G. barreimiae*, *opn4m2* did not show a significantly different rate of evolution compared with the rest of the tree (ω_{Gb} : 0.167; ω_0 : 0.079; model E vs A, LRT, $P = 0.426$), indicating that the protein is conserved in this lineage. The free-ratio model (F) fits the data significantly better than the two-ratio model (C), where *P. andruzzii* is the foreground and the rest of the species are

the background (model F vs C, LRT, $P=0.001$). This indicates that the dN/dS ratios are indeed different among background lineages (Figure 1a).

Exactly the same pattern was observed for rhodopsin. In *P. andruzzii* the protein seems to have lost the signature for selective constraint, whereas in *G. barreimiae* it is under purifying selection. In this case, however, the level of constraint across the tree ($\omega=0.055$) seems more pronounced than for melanopsin ($\omega=0.08$), and the estimated ω_{Pa} is lower (0.209), although the model with $\omega_{Pa}=1$ is still preferred (model C vs D, LRT, $P=0.126$).

To test for divergent selection across the phylogeny incorporating among-site variation in selective constraint, we applied Clade model analyses on both the *opn4m2* and *rho* data sets (Table 3). *P. andruzzii* branch showed a signature of significant divergent selection at ~60% of the melanopsin sites, with $\omega_2=0.014$ on the background lineages and $\omega_{3(Pa)}=1.37$ on the cavefish lineage (CmC_Pa vs M2a_rel, LRT, $P<10^{-3}$). Of the 392 analyzed codons, 22 had a posterior probability of >0.70 to come from the class of divergent sites (Bayes empirical Bayes analysis under the CmC_Pa model). The 22 sites were rather homogeneously distributed across the protein sequence, and included all the six amino acid changes specific to *P. andruzzii* (Table 4 and Supplementary Table 6). Conversely, *G. barreimiae opn4m2* did not show signature of site-specific divergence from the background (CmC_Gb vs M2a_rel, LRT, $P=0.343$). The results obtained from Clade model analyses of *rho* failed to identify a class of divergent sites, although the *P. andruzzii* trend was close to significance ($\omega_2=0.001$ and $\omega_{3Pa}=0.059$; CmC_Pa vs M2a_rel, LRT, $P=0.06$).

The possibility that the divergence shown by *P. andruzzii* melanopsin is due to positive selection was tested using Branch-site models. Branch-site model A estimated that all the sites come from the third and fourth classes (that is, purifying selection or neutral evolution on the background and positive selection on the foreground), whereas no sites are consistently under purifying selection or neutral evolution on background and foreground branches. Under this model, 7% of sites evolve neutrally ($\omega_1=1$) and 93% are under purifying selection on the background lineages (estimated $\omega_0=0.061$), and the estimated ω for

these sites in *P. andruzzii* is $\omega_{Pa}=1.135$. The constrained null model with ω_{Pa} fixed at 1 fitted the data better than Branch-site model A (LRT, $P=0.45$).

In summary, the results of Branch models and Clade models consistently suggest that selective constraint was relaxed on melanopsin in *P. andruzzii*. The $\omega_{3(Pa)}=1.37$ estimated by CmC may indicate

Table 4 Melanopsin sites identified by Bayes empirical Bayes analysis as coming from the class of divergent sites under the CmC_Pa model (PP cutoff = 70%)

Amino acid site	PP	Protein domain
34	0.73301	Extracellular
37	0.83092	Extracellular
45	0.81544	Extracellular
50	0.712	Extracellular
69 ^a	0.9634	Transmembrane 1
160	0.77478	Transmembrane 3
<u>162^a</u>	0.78929	Cytoplasmic
188	0.70835	Transmembrane 4
192	0.70139	Transmembrane 4
<u>202^a</u>	0.73608	Transmembrane 4
<u>205^a</u>	0.76531	Extracellular
235	0.70811	Transmembrane 5
261	0.72722	Cytoplasmic
265	0.73131	Cytoplasmic
271	0.7328	Cytoplasmic
274	0.79893	Cytoplasmic
<u>285^a</u>	0.76222	Cytoplasmic
<u>343^a</u>	0.81869	Transmembrane 7
366	0.70826	Cytoplasmic
370	0.71896	Cytoplasmic
376	0.8297	Cytoplasmic
397	0.78226	Cytoplasmic

Abbreviation: PP, posterior probability.

^aAmino acid changes specific of *Phreatichthys andruzzii* branch. The amino acid changes predicted to be deleterious by Panther and SIFT analyses are underlined (Supplementary Table 6).

Table 3 Log-likelihood values (lnL), parameter estimates and likelihood ratio tests for Clade model analyses of the melanopsin and rhodopsin data set

Clade model	Site class 0		Site class 1		Site class 2		lnL ^a	np ^b	2ΔlnL ^c	d.f.	P-value
	ω_0	p_0	ω_1	p_1	ω_2, ω_3	p_2					
<i>Melanopsin (opn4m2)</i>											
CmC_Pa	0.180	0.375	1	0.019	$\omega_2=0.014$	0.606	-5706.616	21	21.721	1	3.154E-06
					$\omega_{3(Pa)}=1.370$						
CmC_Gb	0.017	0.614	1	0.019	$\omega_2=0.184$	0.367	-5717.026	21	0.8999	1	0.343
					$\omega_{3(Gb)}=0.484$						
M2a_rel	0.017	0.617	1	0.019	$\omega_2 = \omega_3=0.187$	0.365	-5717.476	20			
<i>Rhodopsin (rho)</i>											
CmC Pa	0.147	0.329	1	0.028	$\omega_2 = 0.0013$	0.643	-4279.794	21	3.420	1	0.064
					$\omega_{3(Pa)} = 0.059$						
CmC Gb	0.154	0.314	1	0.027	$\omega_2 = 0.0031$	0.659	-4281.461	21	0.086	1	0.770
					$\omega_{3(Gb)} = 0.0$						
M2a_rel	0.152	0.317	1	0.028	$\omega_2 = \omega_3 = 0.0028$	0.655	-4281.504	20			

^aln(likelihood) value.

^bNumber of parameters.

^cTwice the difference of ln(likelihood) between the two models compared.

the action of weak positive selection as well, a scenario that was not supported by Branch-site models. Rhodopsin showed a signature of relaxation in *P. andruzzii* under different Branch models, whereas CmC showed a similar trend, but the divergence between the cavefish lineage and the background was not significant. Both proteins were under selective constraint in *G. barreimiae*.

Dating the relaxation of selection on the melanopsin photoreceptor

To estimate when the selective pressure on *P. andruzzii* melanopsin became relaxed, we used two sources of information: the estimated rates of synonymous and nonsynonymous substitutions across the lineages of the tree shown in Figure 1a and the estimated divergence time between *P. andruzzii* and *G. barreimiae* (*T*). Posterior estimates using Phylobayes indicate a mean divergence time between the two cavefish species of 15.1 Myr, although with a large range of uncertainty when considering 95% posterior densities (7.4–26.7 Myr) (Supplementary Table 5 and Supplementary Figures 1 and 2).

The divergence time between the two cavefish, *T*, and its 95% interval were used to calculate when the functional constraint on the *P. andruzzii* *opn4m2* gene became relaxed. The frequency distribution of relaxation time ranges from 2.5 to 9.0 Myr, with mean and median of 5.3 Myr (Figure 1a).

DISCUSSION

Statistically supported evidence for relaxed selection on melanopsin and rhodopsin photoreceptors

Studies on the mechanism of regressive evolution often start from the identification of a regressed phenotypic trait, connect the trait to a specific environmental condition and then test for relaxed selection on candidate genes possibly involved in the phenotype. If relaxation of selection is the best model to explain the observed data, then the existence of a causal relationship between the new environment, the genotype and the regressive phenotype is inferred. For example, Niemiller *et al.* (2013) studied regressive evolution in the amblyopsid blind cavefish from North America investigating whether the candidate gene rhodopsin showed evidence of relaxed selection. The authors concluded that the loss of functional constraints caused parallel inactivation of the protein in different cave lineages, suggesting multiple independent cave colonization events. Similarly, Veilleux *et al.* (2013) studied the evolution of the *opn1sw* gene, coding for visual receptors sensitive to diurnal short wavelengths, in lemurs living in a nocturnal environment. The ‘regressive’ hypothesis was that monochromatic primates may have lost the dichromatic color vision as a result of the relaxation of selective constraints on genes such as *opn1sw*, as the ability to perceive short wavelengths would be useless in nocturnal animals. The study concluded that selection on *opn1sw* was weakened or relaxed in animals living in closed canopy rainforests, although the gene, and thus dichromacy, was still under strong purifying selection in similar species living in a brighter environment.

In our study, we did not have to hypothesize that gene may have lost the selective constraints that, in turn, may have led to the regression of the phenotype. Instead, we started from the results of comparative functional analysis of the circadian clock that proved how specific LOF mutations on photoreceptors caused the impairment of an important physiological and adaptive mechanism (Cavallari *et al.*, 2011). Specifically, the LOF mutation, the gene, the phenotype and the environmental condition that led to the regression were already well defined. Having this information available, we could test whether the experimentally proven relationship between an event of pseudogenization and a regressive phenotype arose by relaxation of natural selection in a new environment. Our results suggest that the melanopsin protein

is under neutral evolution in *P. andruzzii*, most probably as a consequence of the loss of selective constraint on this photoreceptor in the dark environment. Although this conclusion may sound obvious given that the *opn4m2* gene is actually a pseudogene, statistically supported evidence of relaxed selection have rarely been found in similar studies (Leys *et al.*, 2005; Li and He, 2009; Zhao *et al.*, 2010), possibly because statistical tests have little power if the change in selective regime is relatively recent. To generalize our conclusions, we tested for the presence of relaxed selection in a second protein, the widely studied photoreceptor rhodopsin. We expected this protein to be under purifying selection in *P. andruzzii* for the following reasons: (1) in this cavefish the *rho* gene shows an intact open reading frame that encodes a full-length opsin photoreceptor, (2) functional experiments have proven its ability to respond to light and (3) behavioral experiments suggest that it may be involved in negative phototaxis (Tarttelin *et al.*, 2012). Indeed, in analyzing this gene in both cave and surface *G. barreimiae* individuals (10 chromosomes) we did not observe a single nucleotide change within the species (Supplementary Table 2), and comparison with other cyprinid species showed only one species-specific amino acid change that should not affect the function of the protein (Figure 2). Conversely, *P. andruzzii* showed both intra- and inter-specific variation in amino acids, most of which with a predicted disrupting effect (Figure 2), although crucial amino acid residues for the photoreception are conserved, such as a lysine (K296) in the seventh transmembrane domain acting as a Schiff's base linkage to the chromophore and the ERY[W] motif (134–136), necessary to bind the α -subunit of transducine (Tarttelin *et al.*, 2012). We believe that this observation, together with the statistical support obtained by a model for neutral evolution of the *rho* gene on *P. andruzzii* branch (Table 2), reinforce our conclusion that relaxed selection operates on light-responsive mechanisms in this species.

Besides neutrality (Wilkens, 1988, 2010), other theories have been proposed to explain how regressive evolution may happen (Espinasa and Espinasa, 2008). In particular, two main hypotheses have been explored: first, positive selection directly eliminates traits, either because the reduction in function is itself beneficial or because it allows the preservation of energy (Protas *et al.*, 2007; Lang *et al.*, 2012); second, positive selection indirectly causes the loss of a trait as a by-product of the gain of a new adaptive function, through mechanisms such as genetic hitchhiking or pleiotropy (Jeffery, 2009). It may be argued that the neutral scenario supported by our results as the most plausible explanation for the evolution of a ‘blind’ circadian clock does not definitively rule out alternative possibilities. In particular, the Branch models implemented in CODEML were originally designed to detect adaptive evolution by positive selection on a specific lineage (Yang, 1998), although it is very commonly used in the literature for cases of relaxed selection (Li and He, 2009; Niemiller *et al.*, 2013; Zhao *et al.*, 2010; Feng *et al.*, 2014; Marková *et al.*, 2014). As only a few sites may be affected by positive selection in an otherwise conserved background, this approach of averaging rates over sites and over time may have little power (Yang and Nielsen, 2002). If that was also the case for the *opn4m2* gene of *P. andruzzii*, a lack of power of the test may be the reason why $\omega_{pa}=1.189$ resulted not significantly higher than 1 (Table 2). To overcome this limitation, we analyzed our data sets with Clade models that account for heterogeneity across sites. The results show that $\sim 60\%$ of the analyzed melanopsin sites are evolving at a different rate in *P. andruzzii* compared with the rest of the tree (Tables 3 and 4). The estimated $\omega_{pa}=1.37$ would indicate that selective constraint was relaxed on this lineage, although the action of weak positive selection cannot be ruled out. However, the

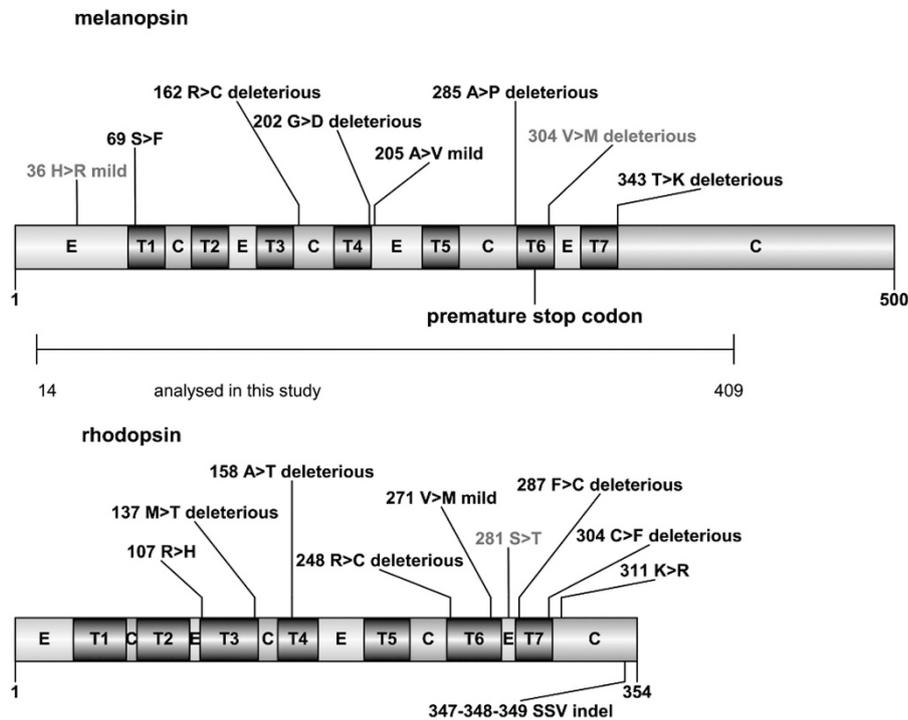


Figure 2 Graphic representation of the two proteins analyzed in this study. Species-specific nonsynonymous fixed substitutions of *P. andruzzii* (black) and *G. barreimiae* (gray) are indicated together with their predicted effect. C, cytoplasmic domain; E, extracellular domain; T, transmembrane domain.

hypothesis that a class of sites is evolving under positive selection in *P. andruzzii* was not supported by a third approach based on Branch-site models. In addition, it is difficult to imagine a direct or indirect adaptive explanation for the inactivation of this melanopsin photoreceptor. In the case of eye regression, the energy-saving hypothesis was not completely dismissed, but still lacks experimental support. In particular, the burden posed by the development and the maintenance of the eye on the organism's survival in the cave is still unclear (Sadoglu, 1967; Protas *et al.* 2007; Espinasa and Espinasa, 2008). Such a direct benefit seems even less convincing in the case of a single nonvisual photoreceptor that is unlikely to contribute to eye maintenance. Furthermore, its downstream signaling pathway is still sensitive to other external signals and, therefore, still partially functional (Cavallari *et al.*, 2011). As for the indirect outcome of positive selection, at present, improvements in other physiological mechanisms due to the loss of melanopsin function are hard to envisage. In addition, the selection coefficient should be high enough to counteract the effect of genetic drift in populations that presumably have rather limited effective population sizes (Brdic *et al.*, 2012). As at this point no credible selective hypothesis can be formulated or tested, thus we are confident to conclude that neutrality explains the regressive evolution of melanopsin in *P. andruzzii*.

Divergence time between the two cavefish *P. andruzzii* and *G. barreimiae*

The divergence time between the two cavefish *P. andruzzii* and *G. barreimiae* was required to infer when the selective pressure on the *opn4m2* gene became relaxed in *P. andruzzii*. Using a Bayesian approach and the *cytb* as a marker, Colli *et al.* (2008) found that divergence times at 95% highest posterior density interval were 8.3–11.5 Myr for the subclade including *P. andruzzii* and *G. barreimiae* and 0.8–2.0 Myr for the epigean and hypogean *G. barreimiae*.

However, Kruckenhauser *et al.* (2011) estimated a split below 1 Myr between the two forms of *G. barreimiae*, and their experiments indicate that the sequences of the *cytb* gene used by Colli *et al.* (2008) may be derived partly from the functional *cytb* gene and partly from nuclear pseudogenes (numts (copies of mitochondrial genes that have been translocated into the nuclear genome)). For this reason, in the present study we resequenced the *cytb* gene in both *G. barreimiae* and *P. andruzzii* cavefish and analyzed this together with *cytb* sequences from an additional 146 species (Supplementary Figures 1 and 2). We provide a posterior mean estimate of 15.1 Myr and 95% highest posterior density interval of 7.4–26.7 Myr for the divergence between the two species, an estimate that turned out to be robust to changes in tree prior (Birth-Death or Uniform), and replacement model (GTR or CAT) (Supplementary Table 5).

Estimating the time of the Somalian cave colonization

If the absence of light typical of the cave environment caused the relaxation of selective pressure on the melanopsin protein, estimating when the molecule was freed from constraint would allow us to infer when the cave was colonized. After concluding that the melanopsin in *P. andruzzii* shows a signature of relaxation of natural selection, we could assume a neutral rate of evolution for this genomic sequence, and we used this information to estimate for how long the cavefish has been in the dark environment. We can confidently assume that *P. andruzzii* populations have been isolated under the Somalian desert at least since the desertification of the surface, but what happened before the colonization and the isolation in the cave is still unknown. Our results based on the rate of evolution of the *opn4m2* gene across a phylogeny of teleost species suggest that the colonization may have happened at ~5.3 Myr, with an interval that ranges from 2.5 to 9.0 Myr (Figure 1a). If, on the other hand, positive selection drove the LOF to fixation, the neutral evolution would have started only after the pseudogenization event, and would not coincide with the

colonization of the cave. However, at present this possibility is not supported by any evidence. In addition, the fact that the LOF was found on both chromosomes of each of the two individuals from different sampling localities suggests that the pseudogenization happened only once. In this case, the inactivation of the gene would have been already present in the ancestral populations that then colonized different parts of the hydrographic system. This is supported by the analysis of the genetic variation of 30 allozyme loci indicating the presence of a genetic structure between sampling localities and a respective low or absent level of gene flow between them (Sbordoni *et al.*, 1996). However, the possibility that the historic locality of certain cave populations may have been much closer together than they are presently should be considered. We cannot exclude that the fixation of the LOF predates the colonization of the subterranean environment, but this alternative scenario seems quite unlikely. Conversely, no line of evidence from this and previous studies on this species support multiple independent colonization events of the dark environment, as observed in other species (Strecker *et al.*, 2012; Bradic *et al.*, 2013).

CONCLUSIONS

In summary, our results suggest that *P. andruzzii* represents an excellent model to study regressive evolution in constant darkness, as the long time spent in the absence of light allowed an extreme level of genetic and phenotypic changes and adaptations. Our results suggest that in the case of this species, the evolutionary framework in which the circadian clock lost its ability to be synchronized by light is very likely neutral regressive evolution due to loss of selective constraint. The evolution of remarkable troglomorphic traits can be due to a long history of divergence of *P. andruzzii* from similar cyprinid species living in internal water of the same continent, a rather old colonization of the phreatic system under the Somalian desert, and the total absence of gene flow with surface populations given the disappearance of the epigeal forms. Although clearly further genetic or genomic analysis will be required to obtain a higher resolution and improve the confidence of our time estimation of this process, this study represents the first key step to reconstructing the evolutionary history of this fascinating species using genetic information.

Data archiving

Sequence data have been submitted to GenBank: KU926693, KU738784, KT953343–KT953348, KU302709–KU302711.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We acknowledge Giorgio Bertorelle and Lino Ometto for important scientific input and discussion, Helmut Sattmann and Elisabeth Haring for help with sample collection and critical discussions, Barbara Däubel for technical assistance, and Francesca Tassi and Silvia Guglielmi for helping with figures. This project was partly funded within the SYNTHESYS framework (project no. AT-TAF-3369).

- Berti R, Messana G (2010). Subterranean fishes of Africa. In: Trajano E, Bichuette ME, Kapoor BG (eds), *Biology of Subterranean Fishes*. Science Publishers: Enfield, NH, pp 357–395.
- Bielawski JP, Yang Z (2004). A maximum likelihood method for detecting functional divergence at individual codon sites, with application to gene family evolution. *J Mol Evol* **59**: 121–132.
- Bradic M, Beerli P, García-de León FJ, Esquivel-Bobadilla S, Borowsky RL (2012). Gene flow and population structure in the Mexican blind cavefish complex (*Astyanax mexicanus*). *BMC Evol Biol* **12**: 9.
- Bradic M, Teotónio H, Borowsky RL (2013). The population genomics of repeated evolution in the blind cavefish *Astyanax mexicanus*. *Mol Biol Evol* **30**: 2383–2400.
- Cavallari N, Frigato E, Vallone D, Fröhlich N, Lopez-Olmeda JF, Foà A *et al.* (2011). A blind circadian clock in cavefish reveals that opsins mediate peripheral clock photoreception. *PLoS Biol* **9**: e1001142.
- Colli L, Paglianti A, Berti R, Gandolfi G, Tagliavini J (2008). Molecular phylogeny of the blind cavefish *Phreatichthys andruzzii* and *Garra barreimiae* within the family Cyprinidae. *Environ Biol Fishes* **84**: 95–107.
- Culver DC, Wilkens H (2000). Critical review of the relevant theories of the evolution of subterranean animals. In: Wilkens H, Culver DC, Humphreys WF (eds). *Ecosystems of the World. Subterranean Ecosystems* Vol. 30, Elsevier: Amsterdam, pp 381–398.
- deMenocal PB (1995). Plio-Pleistocene African climate. *Science* **270**: 53–59.
- Espinasa M, Espinasa L (2008). Losing sight of regressive evolution. *Evol Educ Outreach* **1**: 509–516.
- Feng P, Zheng J, Rossiter SJ, Wang D, Zhao H (2014). Massive losses of taste receptor genes in toothed and baleen whales. *Genome Biol Evol* **6**: 1254–1265.
- Gross JB (2012). The complex origin of *Astyanax* cavefish. *BMC Evol Biol* **12**: 105.
- Jeffery WR (2005). Adaptive evolution of eye degeneration in the Mexican blind cavefish. *J Hered* **96**: 185–196.
- Jeffery WR (2009). Regressive evolution in *Astyanax* cavefish. *Annu Rev Genet* **43**: 25–47.
- Katoh K, Standley DM (2013). MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. *Mol Biol Evol* **30**: 772–780.
- Kimura M, Ohta T (1969). The average number of generations until fixation of a mutant gene in a finite population. *Genetics* **61**: 763–771.
- Motoo K (1983). *The Neutral Theory of Molecular Evolution*. Cambridge University Press: Cambridge.
- Kruckenhauser L, Haring E, Seemann R, Sattmann H (2011). Genetic differentiation between cave and surface-dwelling populations of *Garra barreimiae* (Cyprinidae) in Oman. *BMC Evol Biol* **11**: 172.
- Kumar P, Henikoff S, Ng PC (2009). Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc* **4**: 1073–1081.
- Lahti DC, Johnson N a, Ajie BC, Otto SP, Hendry AP, Blumstein DT *et al.* (2009). Relaxed selection in the wild. *Trends Ecol Evol* **24**: 487–496.
- Lang M, Murat S, Clark AG, Gouppil G, Blais C, Matzkin LM *et al.* (2012). Mutations in the neverland gene turned *Drosophila paccha* into an obligate specialist species. *Science* **337**: 1658–1661.
- Lartillot N, Lepage T, Blanquart S (2009). PhyloBayes 3: a Bayesian software package for phylogenetic reconstruction and molecular dating. *Bioinformatics* **25**: 2286–2288.
- Leys R, Cooper SJB, Strecker U, Wilkens H (2005). Regressive evolution of an eye pigment gene in independently evolved eyeless subterranean diving beetles. *Biol Lett* **1**: 496–499.
- Li Z, He S (2009). Relaxed purifying selection of rhodopsin gene within a Chinese endemic cavefish genus *Sinocyclocheilus* (Pisces: Cypriniformes). *Hydrobiologia* **624**: 139–149.
- Marková S, Searle JB, Kotlík P (2014). Relaxed functional constraints on triplicate α -globin gene in the bank vole suggest a different evolutionary history from other rodents. *Heredity (Edinb)* **113**: 64–73.
- Near TJ, Eytan RI, Dornburg A, Kuhn KL, Moore JA, Davis MP *et al.* (2012). Resolution of ray-finned fish phylogeny and timing of diversification. *Proc Natl Acad Sci USA* **109**: 13698–13703.
- Niemiller ML, Fitzpatrick BM, Shah P, Schmitz L, Near TJ (2013). Evidence for repeated loss of selective constraint in rhodopsin of amblyopsid cavefishes (teleostei: Amblyopsidae). *Evolution (NY)* **67**: 732–748.
- Poulson TL, White WB (1969). The cave environment. *Science* **165**: 971–981.
- Protas M, Conrad M, Gross JB, Tabin C, Borowsky R (2007). Regressive evolution in the Mexican Cave Tetra, *Astyanax mexicanus*. *Curr Biol* **17**: 452–454.
- Rota-Stabelli O, Daley AC, Pisani D (2013). Molecular timetrees reveal a cambrian colonization of land and a new scenario for ecdysozoan evolution. *Curr Biol* **23**: 392–398.
- Sadoglu P (1967). The selective value of eye and pigment loss in mexican cave fish. *Evolution (NY)* **21**: 541–549.
- Sbordoni MC, Matthaes E, Mattoccia M, Berti R, Sbordoni V (1996). Genetic variability and differentiation of hypogean Cyprinid fishes from Somalia. *J Zool Syst Evol Res* **34**: 75–84.
- Stamatakis A (2014). RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**: 1312–1313.
- Stemmer M, Schuhmacher L-N, Foulkes NS, Bertolucci C, Wittbrodt J (2015). Cavefish eye loss in response to an early block in retinal differentiation progression. *Development* **142**: 743–752.
- Strecker U, Hausdorf B, Wilkens H (2012). Parallel speciation in *Astyanax* cave fish (Teleostei) in Northern Mexico. *Mol Phylogenet Evol* **62**: 62–70.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013). MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* **30**: 2725–2729.

Banister KE (1984). A subterranean population of *Garra barreimiae* (Teleostei: Cyprinidae) from Oman, with comments on the concept of regressive evolution. *J Nat Hist* **18**: 927–938.

Berti R, Durand JP, Becchi S, Brizzi R, Keller N, Ruffat G (2001). Eye degeneration in the blind cave-dwelling fish *Phreatichthys andruzzii*. *Can J Zool* **79**: 1278–1285.

- Tarttelin EE, Frigato E, Bellingham J, Di Rosa V, Berti R, Foulkes NS *et al.* (2012). Encephalic photoreception and phototactic response in the troglobiont Somalian blind cavefish *Phreatichthys andruzzii*. *J Exp Biol* **215**: 2898–2903.
- Thomas PD, Kejariwal A, Guo N, Mi H, Campbell MJ, Muruganujan A *et al.* (2006). Applications for protein sequence-function evolution data: mRNA/protein expression analysis and coding SNP scoring tools. *Nucleic Acids Res* **34**: W645–W650.
- Veilleux CC, Louis EE, Bolnick DA (2013). Nocturnal light environments influence color vision and signatures of selection on the OPN1SW opsin gene in nocturnal lemurs. *Mol Biol Evol* **30**: 1420–1437.
- Wang X, Thomas SD, Zhang J (2004). Relaxation of selective constraint and loss of function in the evolution of human bitter taste receptor genes. *Hum Mol Genet* **13**: 2671–2678.
- Wang X, Gan X, Li J, Mayden RL, He S (2012). Cyprinid phylogeny based on Bayesian and maximum likelihood analyses of partitioned data: implications for Cyprinidae systematics. *Sci China Life Sci* **55**: 761–773.
- Weadick CJ, Chang BSW (2012). An improved likelihood ratio test for detecting site-specific functional divergence among clades of protein-coding genes. *Mol Biol Evol* **29**: 1297–1300.
- Wilkens H (1988). Evolution and genetics of Epigeal and Cave *Astyanax fasciatus* (Characidae, Pisces). In: Hecht MK, Wallace B (eds), *Evolutionary Biology*. Springer US: Boston, MA, pp 271–367.
- Wilkens H (2010). Genes, modules and the evolution of cave fish. *Heredity (Edinb)* **105**: 413–422.
- Wilkens H, Strecker U (2003). Convergent evolution of the cavefish *Astyanax* (Characidae, Teleostei): genetic evidence from reduced eye-size and pigmentation. *Biol J Linn Soc* **80**: 545–554.
- Yang Z (1998). Likelihood ratio tests for detecting positive selection and application to primate lysozyme evolution. *Mol Biol Evol* **15**: 568–573.
- Yang Z (2007). PAML 4: phylogenetic analysis by maximum likelihood. *Mol Biol Evol* **24**: 1586–1591.
- Yang L, Arunachalam M, Sado T, Levin BA, Golubtsov AS, Freyhof J *et al.* (2012). Molecular phylogeny of the cyprinid tribe Labeonini (Teleostei: Cypriniformes). *Mol Phylogenet Evol* **65**: 362–379.
- Yang Z, Nielsen R (2002). Codon-substitution models for detecting molecular adaptation at individual sites along specific lineages. *Mol Biol Evol* **19**: 908–917.
- Yang Z, Wong WSW, Nielsen R (2005). Bayes empirical Bayes inference of amino acid sites under positive selection. *Mol Biol Evol* **22**: 1107–1118.
- Zhang J, Nielsen R, Yang Z (2005). Evaluation of an improved branch-site likelihood method for detecting positive selection at the molecular level. *Mol Biol Evol* **22**: 2472–2479.
- Zhao H, Yang J-R, Xu H, Zhang J (2010). Pseudogenization of the umami taste receptor gene *Tas1r1* in the giant panda coincided with its dietary switch to bamboo. *Mol Biol Evol* **27**: 2669–2673.

Supplementary Information accompanies this paper on Heredity website (<http://www.nature.com/hdy>)