Check for updates

OPEN ACCESS

EDITED BY Rossana Cecchi, University of Parma, Italy

REVIEWED BY

Francesco Paolo Busardò, Marche Polytechnic University, Italy Semra Akgönüllü, Hacettepe University, Turkey Francesco Saverio Romolo, University of Bergamo, Italy

*CORRESPONDENCE Matteo Marti matteo.marti@unife.it

SPECIALTY SECTION

This article was submitted to Psychopharmacology, a section of the journal Frontiers in Psychiatry

RECEIVED 26 May 2022 ACCEPTED 04 October 2022 PUBLISHED 21 October 2022

CITATION

Barbieri M, Tirri M, Bilel S, Arfè R, Corli G, Marchetti B, Caruso L, Soukupova M, Cristofori V, Serpelloni G and Marti M (2022) Synthetic cannabinoid JWH-073 alters both acute behavior and *in vivo/vitro* electrophysiological responses in mice. *Front. Psychiatry* 13:953909. doi: 10.3389/fpsyt.2022.953909

COPYRIGHT

© 2022 Barbieri, Tirri, Bilel, Arfè, Corli, Marchetti, Caruso, Soukupova, Cristofori, Serpelloni and Marti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use. distribution or reproduction

in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Synthetic cannabinoid JWH-073 alters both acute behavior and *in vivo/vitro* electrophysiological responses in mice

Mario Barbieri¹, Micaela Tirri², Sabrine Bilel², Raffaella Arfè², Giorgia Corli², Beatrice Marchetti², Lorenzo Caruso³, Marie Soukupova¹, Virginia Cristofori⁴, Giovanni Serpelloni⁵ and Matteo Marti^{2,6*}

¹Department of Neuroscience and Rehabilitation, University of Ferrara, Ferrara, Italy, ²Department of Translational Medicine, Section of Legal Medicine and Laboratory for Technologies of Advanced Therapies (LTTA) Centre, University of Ferrara, Ferrara, Italy, ³Department of Environment and Prevention Sciences, University of Ferrara, Ferrara, Italy, ⁴Department of Chemistry and Pharmaceutical Sciences, University of Ferrara, Ferrara, Italy, ⁵Neuroscience Clinical Center and Transcranial Magnetic Stimulation (TMS) Unit, Verona, Italy, ⁶Department for Anti-Drug Policies, Collaborative Center of the National Early Warning System, Presidency of the Council of Ministers, Rome, Italy

JWH-073 is a synthetic cannabinoid (SCB) that is illegally marketed within an "herbal blend", causing psychoactive effects more intense than those produced by Cannabis. Users report that JWH-073 causes less harmful effects than other SCBs, misrepresenting it as a "safe JWH-018 alternative", which in turn prompts its recreational use. The present study is aimed to investigate the in vivo pharmacological activity on physiological and neurobehavioral parameters in male CD-1 mice after acute 1 mg/kg JWH-073 administration. To this aim we investigate its effect on sensorimotor (visual, acoustic, and tactile), motor (spontaneous motor activity and catalepsy), and memory functions (novel object recognition; NOR) in mice coupling behavioral and EEG data. Moreover, to clarify how memory function is affected by JWH-073, we performed in vitro electrophysiological studies in hippocampal preparations using a Long-Term Potentiation (LTP) stimulation paradigm. We demonstrated that acute administration of JWH-073 transiently decreased motor activity for up to 25 min and visual sensorimotor responses for up to 105 min, with the highest effects at 25 min (\sim 48 and \sim 38%, respectively), while the memory function was altered up to 24h (~33%) in treated-mice as compared to the vehicle. EEG in the somatosensory cortex showed a maximal decrease of α (~23%) and γ (~26%) bands at 15 min, β (~26%) band at 25 min, a maximal increase of θ (~14%) band at 25 min and δ (~35%) band at 2 h, and a significant decrease of θ (~18%), α (~26%), and β (~10%) bands during 24 h. On the other hand, EEG in the hippocampus showed a significant decrease of all bands from 10 min to 2 h, with the maximal effect at 30 min for θ (~34%) and γ (~26%) bands and 2 h for α (~36%), β (~29%), and δ (~15%) bands. Notably, the δ band significant increase both at 5 min (~12%) and 24 h (~19%). Moreover, in vitro results support cognitive function impairment (\sim 60% of decrease) by

interfering with hippocampal synaptic transmission and LTP generation. Our results suggest that JWH-073 deeply alters brain electrical responsiveness with minor behavioral symptoms. Thus, it poses a subtle threat to consumers who mistakenly consider it safer than other SCBs.

KEYWORDS

JWH-073, JWH-018, synthetic cannabinoids, NOR, hippocampus, cortex, electrophysiology, NPS

Introduction

JWH-073 [1-butyl-3-(1-naphthoyl) indole] is a synthetic cannabinoid (SCB), a member of the large family of JWH naphthoylindoles compounds. JWH-073 is a computational melding of the chemical structural features of Δ^9 tetrahydrocannabinol (Δ^9 -THC) with the prototypic aminoalkylindole WIN 55,212-2 developed in the early 1990s. JWH-073 is structurally similar to JWH-018 (1), and like Δ^9 -THC, it has a high binding affinity toward CB₁ (Ki ranging from 8.9 \pm 1.8 and 12.9 \pm 3.4 nM) and CB_2 receptors (38 \pm 24 nM) (2–5). Different from Δ^9 -THC, its monohydroxylated metabolites retain significant affinity and activity at CB1 receptors (4, 6-8). JWH-073 has been introduced on the illegal market after JWH-018 was recognized as an addictive substance, and therefore, it was banned in 2008 (9). JWH-073 produces effects qualitatively similar to JWH-018 (5), while its potency is roughly 50% less. This difference is related to the binding sites of CB1 receptors, which is the chemical structure of JWH-018 is more efficient than JWH-073 (10). JWH-073 has a prolonged effect compared to JWH-018, because of its extended half-life and less receptor internalization (11). JWH-073 pharmacological features, together with its milder effect let users pensive these compounds as safer and more attractive. Thus, it leads to less careful and higher abuse. JWH-073 is often used as an ingredient of herbal blend preparations commonly named "Spice", alone or mixed with other SCBs, like JWH-018 or JWH-250 (5, 12). Beyond these mixtures induce "desired" psychoactive effects, other significant psychiatric and physical adverse effects: agitation/anxiety, restlessness, acute psychosis, hallucinations, photophobia, hyperreflexia, unconsciousness, panic, confusion, drowsiness, and alterations in cognitive abilities (13-19). These can easily involve psychological issues such as recurrent psychotic episodes in vulnerable subjects (14, 20). Furthermore, more intense effects of these mixtures have manifested as serious sympathomimetic-like symptoms such as psychomotor agitation, diaphoresis, palpitations, tachycardia, tachyarrhythmia, hyperreflexia, and generalized convulsions (14, 15, 21). Preclinical studies report that JWH-073 reproduces the typical "tetrad" (hypothermia, analgesia,

hypolocomotion, akinesia) effects previously ascertained for Δ^9 -THC (3, 4, 20. 21), as well as neurological effects, are worse than this latter (22). Moreover, JWH-073 facilitates dopamine release in the shell of the nucleus Accumbens and impairs sensorimotor responses in mice (5). Moreover, it has been shown to affect the drug discrimination paradigm and place preference, in rodents (23-25) and monkeys (26). Along with numerous behavioral effects, SCBs induce EEG hyperexcitability when administered at high concentrations (12). These kinds of alterations apparently include also some typical markers of epilepsy, both in vitro (22) and in vivo (27). Moreover, SCBs are known to be deleterious to memory functions, affecting neuronal memory mechanisms (28, 29). The introduction into the illegal market of new and powerful SCBs creates the need to study more and more carefully SCBs toxic effects (30-32), and in particular those effects related to functional changes in the central nervous system activity, which can represent the basis of acute or long-term neurological and psychiatric alterations. Indeed, together with a new screening method useful to identify SCBs (33), also preclinical studies need to keep up with the continued expansion of the illegal market. The present study is aimed at investigating the in vivo pharmacological activity of acute JWH-073 administration in male CD-1 mice at dosage (1 mg/kg i.p.). The dose chosen is underactive or ineffective on physiological and neurobehavioural parameters, as previously demonstrated in rodents (5, 24). Therefore, we investigate the effect of JWH-073 on sensorimotor (visual, acoustic, and tactile), motor (spontaneous motor activity and catalepsy), and memory functions (novel object recognition; NOR) in mice coupling behavioral and electroencephalogram (EEG) data. To clarify the effects of SCBs on brain electrical activity we recorded EEG from areas potentially involved in epilepsy and sensorimotor impairment, like the hippocampus and somatosensory cortex, analyzing quantitatively EEG at the sub-toxic doses of JWH-073. To investigate if JWH-073 at the tested dose may affect the hippocampal-based synaptic plasticity mechanism such as Long Term synaptic Potentiation (LTP), we used an electrophysiology test on the hippocampus in vitro (29, 34, 35).

Materials and methods

Animals

Eighty-seven-male ICR $(CD-1^{(R)})$ mice, 5–7 months old, weighing 35-30 gr (Centralized Preclinical Research Laboratory, University of Ferrara, Italy), were group-housed (five mice per cage; floor area/animal: 80 cm²; minimum enclosure height: 12 cm) on a 12:12-h light-dark cycle (light on at 6:30 AM), the temperature of 20-22°C, the humidity of 45-55% and were provided ad libitum access to food (Diet 4RF25 GLP; Mucedola, Settimo Milanese, Milan, Italy) and water. Experimental protocols were performed in accordance with the European Communities Council Directive of September 2010 (2010/63/EU) and were approved by the Italian Ministry of Health (license n. 335/2016-PR and license n. 956/2020-PR) and by the Animal Welfare Body of the University of Ferrara. According to the ARRIVE guidelines, adequate measures were taken to reduce the number of employed animals and their pain and discomfort.

Drug preparation and dose selection

JWH-073 was purchased from LGC Standards (LGC Standards S.r.L., Sesto San Giovanni, Milan, Italy) while the CB₁ receptor antagonist AM-251 was purchased from Tocris (Tocris, Bristol, United Kingdom). Drugs for *in vivo* tests were initially dissolved in absolute ethanol (final concentration = 5%) and Tween 80 (2%) and brought to the final volume with saline (0.9% NaCl). The solution made with ethanol, Tween 80, and saline was also used as a vehicle. AM-251 (1 mg/kg) was administered 20 min before the JWH-073 injection. Drugs were administered intraperitoneally (i.p.), at a volume of 4 μ l/gr. For *in vitro* electrophysiology the substances were dissolved in absolute ethanol (maximum concentration = 0.1% v/v). The JWH-073 dosage of 1 mg/kg was chosen based on previous studies [(5); see Supplementary materials].

Behavioral studies

Evaluation of sensorimotor and motor responses

The effects of 1 mg/kg JWH-073 were investigated using a consolidated battery of behavioral tests routinely used in our laboratory for preclinical investigation of NPSs in rodents (5, 36–39), in a consecutive manner, according to specific time scheme (for technical details, see Supplementary material).

Behavioral tests together with EEG recordings were conducted in the same ambient conditions of housed animals. The behavior of mice was previously recorded with a B/W camera (Ugo Basile, Italy) and analyzed off-line in blind. Moreover, in order to analyze and correlate in more detail EEG with the degree of motor activity (potential sudden and temporary motor blocks), the horizontal locomotion of the mouse was video recorded during EEG for off-line analysis (Figure 1). Locomotor parameters were analyzed 30 min before (basal values) and after treatment.

Novel object recognition test

In order to evaluate the effect of JWH-073 on the memory functions of the mouse, the Novel Object Recognition (NOR) test was carried out at two-time points (2 and 24 h after the administration) based on the alterations induced by the tested compound on sensorimotor and motor responses (Table 1). As previously reported (29), the test was conducted in three phases: habituation, familiarization, and choice (for technical details of the methods used, see Supplementary material).

Electrophysiological studies *in vitro* in hippocampal slices

Tissue preparation

The hippocampal transverse slice model was used to evaluate the acute effects of JWH-073 on synaptic excitatory transmission and plasticity. Tissue preparation protocols are described in detail by Barbieri et al. (29). In brief, after mouse decapitation, the brain was quickly extracted, and the hippocampus was isolated and placed in ice-cold artificial cerebrospinal fluid (aCSF), of the following composition (mM): NaCl, 126; KCl, 2; KH₂PO₄, 1.25; NaHCO₃, 26; MgSO₄, 2.0; CaCl₂, 2.5; dglucose 10. The aCSF was saturated with 95% O₂/5% CO₂. Transverse hippocampal slices (425 μ m thick) were cut with a tissue chopper, then placed for almost 90 min in a Haas-style incubation chamber for recovery until recording.

Electrophysiological recording

A single slice was placed into a recording chamber (3 ml total volume) and continuously superfused (3.0 ml/min) with warmed (33°C) aCSF. Details of recording procedures are described in 27. In brief, once obtained a stable synaptic response from stimulation\recording from CA1 Shaffer collaterals area for at least 20 min, a stimulus/response curve (SRC) was generated as previously described (29) to extrapolate the stimulation intensity (the one achieving the 40% of maximal EPSP) constantly adopted throughout the experiment. Substances (JWH-073, AM251, or vehicle) were added to the reservoir, and the bath was applied *via* the perfusion line. At steady state effect on fEPSP of JWH-073 1 μ M, the 40% stimulation amplitude was recalculated, repeating the SRC and adopted for the following LTP procedure.

To evaluate modifications of synaptic plasticity we induced LTP following methods detailed in previously studies (5, 40). At the end of the experimental protocols, pathways were



simultaneously activated with TB10 stimulation to evoke the maximally achievable potentiation as a control for slice viability. Amplitude values were used as a reference for the maximal potentiation achievable. The same data was also used to calculate TB5 stimulation-induced LTP as a fraction of maximally inducible potentiation in each slice to equalize differences in LTP between preparations.

Electrophysiological studies in vivo

Surgery and electrode implantation procedures

Under initial i.p. anesthesia (ketamine/xylazine, 75/8.25 mg/kg), mice were secured to a digital stereotaxic device (Stoelting, Wood Dale, IL, USA). The mouse skull was shaved,

rubbed with the anesthetic cream, and skin incised exposing beneath the bone. Was then drilled a 0.9 mm Ø hole to implant an electrode headset at a stereotaxic position relative to bregma. Electrodes were alternatively positioned in two areas: (1) left frontoparietal cortex - somatosensory area (AP -0.8 mm; ML -1.4 mm; DV -0.5 mm) and (2) left ventral hippocampus (AP -2.0 mm; ML -1.4 mm; DV -1.5 mm). Two further holes were drilled on the same side and loaded just upon the brain with stainless-steel screws used to stabilize the headset to the bone. Finally, dental acrylic was overlaid, embedding screws and covering the inserted headset. For headset building and installation details see the supp documents. Post-surgery pain discomfort and infection were prevented i.p. injecting mice with Tramadol (50 mg/kg) and Imipenem (25 mg/kg, i.p.), for 2 days. On the 7th day of recovery, the headset of the mouse was

Test	treatment	Control (-10 min)	10 min	25 min	105 min	120 min	24 h
Spontaneous motor activity (s)	Vehicle	165.5 ± 5.5	152.5 ± 6.5	55.2 ± 3.0	29.3 ± 5.1	15.3 ± 5.1	135.6 ± 4.5
	JWH-073	168.7 ± 6.2	87.3 ± 4.6**	$28.5 \pm 3.2^*$	25.3 ± 5.6	13.6 ± 4.2	145.5 ± 7.5
Visual object (arbitrary units)	Vehicle	9 ± 0	9 ± 0	9 ± 0	8.5 ± 0.94	120 min 15.3 ± 5.1 13.6 ± 4.2 8.8 ± 0.74 8.5 ± 0.76 11.96 ± 0.2 11.72 ± 0.7 12 ± 0 0 ± 0 0 ± 0 0.29 ± 0.023 $0.16 \pm 0.025^{**}$ 45.5 ± 2.4 42.0 ± 4.2	9 ± 0
	JWH-073	9 ± 0	$5.8 \pm 0.15^{**}$	5.6 ± 0.25**	$6.5 \pm 0.32^*$	8.5 ± 0.76	9 ± 0
Acoustic (arbitrary units)	Vehicle	12 ± 0	12 ± 0	11.56 ± 0.3	12 ± 0	11.96 ± 0.2	12 ± 0
	JWH-073	12 ± 0	11.63 ± 0.24	11.55 ± 0.54	11.32 ± 0.31	11.72 ± 0.7	12 ± 0
Overall tactile (arbitrary units)	Vehicle	12 ± 0	12 ± 0	12 ± 0	11.54 ± 0.32	12 ± 0	12 ± 0
	JWH-073	12 ± 0	11.25 ± 0.64	11.47 ± 0.84	11.12 ± 0.21	12 ± 0	12 ± 0
Time on bar (s)	Vehicle	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0	0 ± 0
	JWH-073	0 ± 0	0 ± 0	0 ± 0	$i4$ 11.32 ± 0.31 11.72 ± 0.7 11.54 ± 0.32 12 ± 0 34 11.12 ± 0.21 12 ± 0 0 ± 0 0 ± 0 0 ± 0 0 ± 0 nd 0.29 ± 0.023 nd $0.16 \pm 0.025^{**}$	0 ± 0	
NORRecognition Index	Vehicle	nd	nd	nd	nd	0.29 ± 0.023	0.30 ± 0.024
	JWH-073	nd	nd	nd	nd	$0.16 \pm 0.025^{**}$	$0.20 \pm 0.021^{*}$
NORTOE (s)	Vehicle	nd	nd	nd	nd	45.5 ± 2.4	38.1 ± 3.2
	JWH-073	nd	nd	nd	nd	42.0 ± 4.2	36.0 ± 2.3

TABLE 1 Effects of JWH-073 (1 mg/kg i.p.) or vehicle on spontaneous motor activity, sensorimotor (visual, acoustic and tactile) responses and object memory recognition (NOR) in mice.

Data are expressed as absolute values in seconds (spontaneous motor activity, time on bar and time of object exploration; TOE), arbitrary units (visual, acoustic and tactile responses) and recognition index (novel object recognition test, NOR). Data represent the mean \pm SEM of 5 different animals for each treatment. Statistical analysis was performed with two-way ANOVA followed by Bonferroni's *post-hoc* test for multiple comparisons, while the NOR test was performed with *t*-Student test for unpaired data. *p < 0.05, **p < 0.01, vs. vehicle. Bold values in Table 1 referred to statistically significant variations among all the results.

connected to a swivel system with a 3 wires-cable (Plastics One), and electrode resistance was measured, discarding the implant with electrical resistance, $<1 \text{ M}\Omega$. For 5 days before drug/vehicle administration, mice were daily handled, transferred to the recording arena, and connected to a recording device for 1 h to archive EEG later used as control. The signal was delivered to a preamplifier/data acquisition system (EEG100C, MP150, Biopac), allowing the recording of EEG and video synchronously on a PC using AcqKnowledge 4.5 (Biopac) software. The EEG signal was digitized (2 kHz), amplified (from 5 to 50 k fold), and filtered (5 kHz lp, 0.05 Hz hp). EEG baseline was considered stable when relative Fast Fourier Transformation (FFT; 10s window). showed variations within 10% for a period of almost 10 min. EEG and video were then recorded for 30 min soon after substance injection, then after 2 and 24 h. Drug treatment consisted of intraperitoneal injection of JWH-073 (1 mg/kg; hippocampus n = 6; cortex n = 5) or vehicle (120 µl at 1 mg/kg, n = 5) as sham treatment. Once experimental procedures were terminated, anatomical studies were conducted on all animals to verify the correctness of the implant position.

Data and statistical analysis

Behavioral studies

Data are expressed as absolute values in seconds (spontaneous motor activity, time on bar, and TOE time), arbitrary units (visual object, acoustic and overall tactile responses), and RI in the NOR test. All data are shown as the mean \pm SEM of 5 different animals (n = 5) for each treatment. Statistical analysis of the effects in behavioral tests over time was performed by two-way ANOVA followed by Bonferroni's *post-hoc* test for multiple comparisons. Analysis of the effect induced by treatment in the NOR test was performed with the unpaired *t*-Student test. The significant level was set at p < 0.05. Statistical analysis was performed using the program Prism software (GraphPad Prism, USA).

In vitro data analysis

Data from *in vitro* recordings has been analyzed as previously described (5). In brief, fEPSP slope was measured to calculate drug effects on synaptic excitatory transmission and TB5 stimulation-induced synaptic potentiation. Steady-state values of net potentiation produced by TB5 stimulation were obtained by averaging the values of the 11 consecutive responses recorded over the 5 min period between 40 and 45 min after TB5 stimulation. The maximally achievable potentiation was calculated by averaging the values of 5 responses over the 2 min period between 13 and 15 min after TB10 stimulation. The unpaired *t*-Student test was used to compare the vehicle-treated with untreated control groups and p < 0.05 was considered statistically significant.

In vivo EEG data analysis

EEG data analysis was performed off-line with AcqKnowledge 4.5 software (BIOPAC, USA). EEG showing

components like interictal and/or spikes, spike and wave complexes, or poly-spikes, were defined as "events" when amplitude was at least 2-fold the baseline defined as 20 s trace immediately before and long 2 s or more (41). Spectrograms and FFT (used to quantify EEG power spectrum) were calculated with IgorPro 8.0 software. For spectrograms has been used the Time-Frequency toolbox on data previously resampled to 500 Hz and digitally HP filtered at 0.05 Hz. Quantitative EEG was calculated on a 5 s window. Power for each characteristic EEG band [delta (δ) = 0.3-4 Hz; theta (θ)= 4-8 Hz; alpha (α)= 8-13 Hz; beta (β) = 13-30 Hz; gamma (γ) = 30-48 Hz)] was computed in 10 s bins from FFT for each animal and averaged into 1 min bins when necessary. When calculated, spectral edge frequency (SEF) was 95%.

Two-way ANOVA with the Dunnet test was used to compare the control EEG amplitude of groups before treatment with values after treatment. P < 0.05 was considered statistically significant.

Results

Behavioral studies

Evaluation of sensorimotor and motor responses

Sensorimotor (visual, acoustic, and overall tactile) and motor (time on bar, spontaneous activity) results are reported in Table 1. Sensorimotor parameters were unmodified in vehicletreated mice and the effect was similar to that observed in naïve untreated animals (data not shown). Systemic administration of JWH-073 (1 mg/kg i.p.) reduced the visual object response in mice of ${\sim}36\%$ (at 10 min), ${\sim}38\%$ (at 25 min), and ${\sim}24\%$ (at 105 min) in comparison with vehicle-treated mice [effect of treatment ($F_{1,48} = 36.07, p < 0.0001$), time ($F_{5,48} = 7.436$, p < 0.0001) and time × treatment interaction (F_{5,48} = 6.933, p < 0.0001]. JWH-073 at 1 mg/kg did not affect at any time point the acoustic and tactile sensorimotor responses in mice. The vehicle did not change the bar test response and spontaneous motor activity in mice, and the effects were similar to that observed in naïve untreated animals (data not shown). Systemic administration of JWH-073 (1 mg/kg i.p.) did not cause catalepsy/akinesia in the bar test for up to 24 h. Conversely, JWH-073 reduced spontaneous motor activity in mice at about \sim 43% (at 10 min) and \sim 48% (at 25 min) respect vehicle-treated mice [effect of treatment ($F_{1,48} = 21.97, p < 0.0001$), time ($F_{5,48}$ = 309.9, p < 0.0001), and time × treatment interaction (F_{5,48} = 14.28, *p* < 0.0001)].

Novel object recognition test

To investigate whether the novel synthetic cannabinoid agonist JWH-073 affects memory retention in mice, the NOR test was performed, comparing results with vehicle (Table 1). During the familiarization phase, no difference was seen in the time spent by vehicle untreated-mice to investigate the two objects, and there were no significant differences in the NOR test, similarly to naïve untreated mice (data not shown). In contrast, treatment with the synthetic cannabinoid-induced a significant impairment of recognition memory. In fact, significant changes in RI values were observed at 2 h (t = 3.729, df = 8, p = 0.0058) and at 24 h (t = 2.820, df = 8, p = 0.0225) from the adm inistration of JWH- 073. Clearly, no differences in TOE time between the untreated control animals and vehicle-treated mice (data not shown) were observed. The TOE time in the choice phase was not impaired both at 2 and 24 h after the JWH-073 injection.

Spontaneous horizontal locomotor activity during EEG recordings

The amount of spontaneous horizontal locomotor activity of mice before (30 min) and after (30 min) treatment with vehicle or JWH-073 is shown in Figure 1. After 5 min upon JWH-073 administration, the locomotion is dramatically reduced, resulting in a splitting of the two curves (vehicle vs. JWH-073) immediately after treatment. After \sim 23 min, the curve relative to JWH-073 treated mice displays a return to values relative to vehicle-treated animals' locomotion. The comparison of the cumulative distance traveled by vehicle- and JWH-073-treated groups are summarized in the inset emphasizing the significant difference in the cannabinoid effect to inhibit locomotion. It is worth noting that mice treated with JWH-073 were free of behavioral patterns of pathological significance following Racine stages (42).

In vitro electrophysiological studies

Effects of JWH-073 on synaptic transmission in CA1 hippocampal area

Changes in the average slope amplitude of evoked CA1 synaptic population response are represented in Figure 2A. JWH-073 in all tests analyzed, induced a depressive effect on fEPSP, declining from 2 to 5 min after first contact with JWH-073 1 μ M, with steady state maximal effect after about 60 min. The effect of the vehicle is reported for comparison. Figure 2B are compared steady-state values of fEPSP (vehicle: 100 ± 3.6; JWH-073: 46.7 ± 1.8), with a statistically significant difference (p < 0.01). Application of AM-251 (Figure 2B) confirms the selectivity of JWH-073 on CB1 receptors.

Effects of JWH-073 on paired-pulse stimulation and fiber volley in CA1 hippocampal area

A paired-pulse stimulation protocol was used to test for pre- or postsynaptic effects of JWH-073 (Figure 2C). At JWH-073 steady-state effect, the ratio between fEPSP slope of conditioning (S1) and test pulse (S2) (Figure 2C, upper, trace



2) was modified with respect to vehicle ratio (Figure 2C, upper, trace 1) accounting for a direct presynaptic effect. The fiber volley amplitude comparing JWH-073 to vehicle effect, showed as it was unmodified (data not shown).

Effect of JWH-073 on synaptic plasticity in the CA1 hippocampal area

Figure 3A shows superimposed normalized average experimental points of LTP test experiments, in the presence of the vehicle or JWH-073. Figure 3B summarizes values at a steady fEPSP slope for the two conditions considered, showing a significant statistical difference (p < 0.01). When compared to the vehicle (122.3 \pm 15.3% Δ increase vs. fEPSP slope baseline), JWH-073 reduced the development of LTP (48.1 \pm 3.5%), impairing the formation of any consistent stable synaptic potentiation. In slices treated with JWH-073, the TB10 saturating test (see Methods) was unable to induce any further potentiation, confirming the strong LTP-inhibition from JWH-073 treatment (data not shown).

In vivo electrophysiological studies

Effect of JWH-073 on *in vivo* EEG in cortical area

As illustrated by the representative experiment shown in Figure 4, JWH-073 1 mg/kg administration evoked a series of

complex and rapid modifications in cortical EEG and precisely identified by the spectrogram.

- (A) JWH-073 modifies clearly EEG (Figure 4) after a few minutes (in this case, 5 min) from administration, with short, high amplitude episodes (asterisk marked) followed by similar throughout the whole recording. EEG spectrogram (Figure 4) unveils how after a few minutes from the drug an evident power reduction of α , β , and γ frequencies bands. EEG power trends to recover with time, with increased incidence of high amplitude events. Interestingly, soon after the drug injection, it's usual to observe some δ bursts, without corresponding EEG evidence.
- (B) To characterize the evolution of the power spectrum in Figure 4, we derived the FFT for windows of the 30 s at defined time points (Figure 5): control (recorded before the drug administration); after administration of JWH-073 at 1, 3, 15, and 30 min. Already at point 1 min, the δ band increases macroscopically, splitting out into two subcomponents [arrows labeled; Figure 5; (43)]. Subsequently, this slow δ component amplitude oscillates along the recording while the faster remained roughly constant throughout, here and in all the animals tested about a broad power depression of mild entity interspersed with short powerful δ bursts.
- (C) For a quantitative EEG analysis, we normalized FFT amplitude measured in a 10 s window for each frequency component, averaging the results in blocks of 5 min



(Figure 6; Table 2) and comparing absolute and relative spectrum variations for different animals. Arose clearly three trends: (1) δ and θ amplitudes from 25 min after JWH-073 until 2 h, which showed a significative gradually increasing trend. (2) β and α faced a significant fall between 5 and 20 min, eventually returning to control. (3) γ significantly depressed from 5 min after JWH- 073 effects which persisted almost for 2 h after treatment, extending effects after 24 h. Indeed θ , α , and β were significantly depressed, while other bands completely recovered.

Abnormal excitability in cortical EEG after JWH-073

Different authors agreed on the epileptogenic activity of SCBs (27): coherently with these observations we selected and analyzed two representative events because potentially early markers of epileptic activity (44). To define if EEG spikes labeled with asterisk in Figure 4, could be markers of epileptogenic activity, we analyzed similar events in a representative recording (Figure 7). The EEG trace on the upper part of panel A refers to a recording coping pharmacological protocol of Figure 4. The effect of JWH-073 on EEG is evident, similarly to those described in Figure 4 and exploited by spectrogram. The spectrum shows initial phases of generalized depressed electrical activity (darker areas), alternated with short powerful events (yellow bands).

The later part of EEG is characterized by a deeply dampened power spectrum. For the analysis of possible background proepileptic electrical components, we selected two representative areas, characterized by evident higher electrical activity on EEG, ideally epileptogenic and marked by "@" (3 min after JWH-073 administration), and "#" (20 min after JWH-073 administration). To analyze the "@" area (Figure 7B), we marked four areas of interest (from 2 to 5) having similar amplitudes, but different intrinsic frequencies. Control is area 1, recorded before JWH-073 and not shown as being outside of figure boundaries. Analysis of these segments marked a common δ increase and segment 5, is characterized by a y collapse. Real changes could be misread, by using a logarithmic Y-axis. Indeed, panel C replays the same FFT using a linear Y-axis which was broken to include two upper peaks. The inset shows the spectrum of panel B area 2, with control FFT hindering all the other FFT (Figure 7C). To sum up, this pattern of bursts, frequent during the experiment, is mainly characterized by hovering δ episodes coped with short γ depression. The second waveform marked with "#" (Figure 7D), and frequent after JWH-073, is characterized by short, high-frequency bursts and an electrical oscillation of gradually increasing amplitude, ending suddenly later with a short dampened electrical activity. Similar sequelae are characteristic of the pre-epileptic ictal events previously quoted. Spectral analysis (panel D, bottom) described a stable peak frequency with a fall of spectral edge frequency (SEF)



 $(\approx -70\%)$ and a synchronous gradual increase in mean power. During the later EEG attenuation, the power magnitude halved with a sudden peak of SEF. Taken together, differently from seizure-like events, the analysis shown in Figures 7B–D differs from interictal activity, characterized by definite frequency increase with the rise of power magnitude (44).

Effect of JWH-073 on *in vivo* EEG in hippocampal area

- (A) Administration of JWH-073 modified hippocampal EEG which differed qualitatively and quantitatively from effects seen for the cortex. Early after JWH-073, EEG amplitude shrank slightly. Spectrum analysis revealed a rapid and persistent fall among all EEG frequency bands (Figure 8).
- (B) To focus on how JWH-073 changed EEG bands with time, we sampled the recording at different time points for windows of 30 s whose FFT is represented in Figure 9. The δ band clearly increased at 1 min of JWH-073, decaying partially at 3 an 15 min returned to basal amplitude, when at

30 min δ and θ raised over control. Conversely, all the other bands showed a quickly generalized dampening, mirroring the spectrogram of Figure 8 where, differently from the cortex, any recovery looks absent.

(C) For a cumulative quantitative EEG analysis we adopted identical criteria used for effect of JWH-073 on in vivo EEG in hippocampal area, point (C) and Figure 6. Results are reported in Figure 10 and Table 3. The δ band shows an immediate, transitory increase, as seen in Figure 9, returning rapidly to control. All the other bands depress progressively until the recording ends. It is relevant (see Discussion) θ band, which progressively attenuates; γ has a similar trend for the first few minutes as the literature already described. Different from the cortex, in the hippocampus at 2 h after JWH-073 administration, all bands are depressed, recovering 24 h later except for γ and $\theta.$ Interestingly, 24 h after JWH-073, δ has an evident amplitude lift compared to other bands determining a net differential effect with δ prevalence over all the others.



Anomalous excitability in hippocampal EEG after JWH-073

JWH-073 treatment induced in the hippocampus a sporadic series of electrical anomalous events. Figure 11A refers to a recording with some representative events, arbitrarily selected (tagged from 1 to 3) for analysis. Events 1 and 2 are characterized by a definite pattern repetitively present in all recordings. Indeed, 1 shows an alternation of short, low amplitude electrical signals, and 2 has higher amplitude spikes. Event 3 is a group of repetitive spikes, resembling interictal epileptic-like events. Figure 11B represented a comparison of 10s FFT between control, events 1 and 2. FFT of 1 unveils a consistent generalized depression, while in 2 the δ component is prevalent. Events 1 and 2 alternance retraces a condition already observed in the cortex (Figure 5), where phases of global depression interspersed with others where δ overcomes the amplitude of control. Complex 3, resembles epileptiform interictal activity: high amplitude regular spiking with a "spike and wave" pattern. Figure 11C reproduces this waveform showing a rhythmic activity (13 s phase). Analyzing one complex at a higher speed (Figure 11C, lower part), it differs from fingerprint of the epileptic-ictal spike (45, 46), with a waveform shape alike pre-ictal event, but much slower. Furthermore, spike amplitudes are 2–3 times that of basal EEG too small to be an epileptic focus. As previously verified, JWH-073 has no evident epileptogenic capabilities even at a 30-fold higher dosage, in rats (22), while hippocampal EEG is modified toward hyperexcitability.

Discussion

This study showed that acute JWH-073 (1 mg/kg; i.p.) administration produces relevant and long-lasting *in vivo* EEG alterations in the somatosensory cortex and hippocampus of mice, which seem to be associated with mild and transitory impairment of visual sensorimotor response and locomotion,



the control EEG amplitude of groups before treatment with values after treatment. p < 0.05; p < 0.01.

EEG band	Control	5 min	10 min	15 min	20 min	25 min	30 min	2 h	24 h
Delta	0 ± 5.64	7.17 ± 2.85	2.45 ± 2.99	-0.65 ± 2.69	3.33 ± 2.81	$24.63\pm3.03^{\$}$	$34.12\pm3.42^{\$}$	$35.57 \pm 8.41^{\$}$	1.40 ± 8.50
Theta	9 ± 1.52	-1.40 ± 1.80	$-12.55 \pm 2.17^{\$}$	-5.95 ± 2.27	3.50 ± 2.61	$14.28\pm2.86^{\$}$	$12.48\pm2.83^{\$}$	$12.36\pm6.32^{\$}$	$-18.16\pm4.22^{\$}$
Alpha	0 ± 3.82	-2.61 ± 1.70	$-18.60\pm1.72^{\$}$	$-23.25 \pm 1.55^{\$}$	$-14.64\pm2.28^{\$}$	-8.60 ± 1.96	-4.35 ± 2.29	-5.03 ± 5.21	$-26.29\pm4.15^{\$}$
Beta	0 ± 8.48	$-13.98\pm1.38^{\$}$	$-26.67 \pm 1.15^{\$}$	$-22.81\pm1.28^{\$}$	$-15.49 \pm 1.55^{\$}$	-8.27 ± 1.55	-1.57 ± 1.67	-2.29 ± 3.87	$-10.48\pm3.85^{\ast}$
Gamma	0 ± 4.83	$-12.07 \pm 1.21^{*}$	$-24.05 \pm 0.88^{\$}$	$-26.53 \pm 0.75^{\$}$	$-20.63 \pm 0.94^{\$}$	$-17.63 \pm 0.93^{\$}$	$-18.73 \pm 1.12^{\$}$	$-19.18\pm2.18^{\$}$	-6.42 ± 2.85

TABLE 2 Quantitative EEG cortex.

Values relative to histograms in figure 6 with relative significance. Values correspond to $\% \pm$ s.e. Two-way ANOVA with Dunnet test used to compare the control EEG amplitude of groups before treatment with values after treatment. *p < 0.05; *p < 0.01.

and a persistent memory functions compromission. These results, together with *in vitro* in hippocampal slices data, underlined the detrimental activity of JWH-073 upon memory mechanisms and brain electrophysiological activity.

Behavioral and locomotor effects

JWH-073 (1 mg\kg) transiently inhibited the visual object response, reduced overall motor activity, and deteriorated the

long-lasting working memory. This suggests that visual and motor impairments, albeit in a transient way, are the effects firstly appearing from mild dose intake of this SCB, rising the relevance for the correlation between impaired psychomotor activity (e.g., driving) and modest SCBs intoxication (47, 48). In fact, impaired visual responses with depressed memory functions alter the subject's ability to react quickly to dangerous situations which require a rapid sensorimotor response, and the recruitment of previously acquired and stored information. Specifically, JWH-073 depressed working



memory in the NOR test (2 and 24h post-administration), similarly to other SCBs (29, 49). Despite this, TOE time exclusive memory-cognitive deficits induced by JWH-073, alike other SCB (49), showing its effects on processes involved in memory formation and retention rather than the impairment of motor activity. The transient reduction by JWH-073 of the distance traveled parameter supported the previous observation. Indeed, Ossato and colleagues, showed that AM-251 prevented the psycho-cognitive and memory effects of JWH-073, demonstrating its CB1 receptors mediated action (5). Moreover, during the period of reduced mobility, mice still preserved minimal sporadic activity such as random head movements, scratches, or a short walk, which are incoherent with catalepsy. This is in line with previous studies showing the limited influence of JWH-073 on locomotion and catalepsy (23, 50).

In vitro hippocampal electrophysiological effects

On the hippocampal *in vitro* slice model, JWH-073 produced effects like JWH-018 (29) and other SCBs (35) such as irreversible fEPSP inhibition, slow time to onset of a steady state, significant inhibition of early and late LTP components. Pre- and post- synaptic activity of JWH-073 suggest that it is active on different cellular pathways being early LTP regulated by CaMKII activation, while late LTP requires PKA, MAPK, and CREB activation with TrkB-gene expression and functional prions CPEB2-3-mediated translation (51). Thus, it implies *in-vivo* impairment of short-and long-term memory, depicting in perspective a persistent memory deficit following repeated drug use, even at sub-toxic doses (52).



In vivo effects on EEG: General considerations

To date, alterations in EEG induced by Δ_9 -THC has been previously demonstrated in human (53, 54), primates (55), and rodents (56). The EEG alteration after SCBs administration has been investigated only in rodents analyzing EEG exclusively to spot epileptic events, from 30 min after administration and over, and a few of them analyzed EEG spectrum (52, 57–62) and never quantitatively. Since JWH-073 impaired both locomotor activity and visual object response in mice during the first 30 min after treatments, we followed EEG evolution at the same time point, a time context for EEG analysis after CB1R activation previously seldom investigated in detail (57, 62).

Our results demonstrated for the first time that apparently harmless transitory impairment of sensorimotor visual response and locomotor activity, occurs with significant EEG alterations, persisting far beyond the recovery of visual and locomotion impairments. The second novelty which emerges from EEG analysis, is the frequency-band selectivity of these alterations, exploiting a complex spectrum modification discussed below in detail.

In vivo effects on EEG: Qualitative and quantitative cortical and hippocampal electroencephalographic analysis

In the sensory cortex, JWH-073 injection produced a severe unbalance between δ , θ , and the other bands within the first 30 min (Figure 6; Table 2). Cortical δ , and θ in a lesser extent, remained unchanged until 20 min after JWH-073 administration, but a gradual highly significant increase follows, persisting for 2 h. Interestingly, the γ band faced a deep and persistent depression, as already described (52, 61). From comparison with other bands' trends, the outcome is a severe unbalance, with a prevalence of δ/θ . Behavioral effects of JWH-073 recovered 2 h later, while the EEG alterations persisted. After 24 h, only δ/γ recovered while θ , α , and β



were still depressed. Taken together these results suggest the harmful effect induced by mild doses of JWH-073 on EEG, with alterations persisting much beyond the recovery of predrug behavior. JWH-073 depresses all hippocampal EEG bands except for δ , a condition lasting up to 2 h after drug contact, with partial recover 24 h later (Figure 10; Table 3). This is the first quantitative and qualitative electrophysiological analysis in the hippocampal area immediately after an SCB administration. Following data interpretation required correlating dynamically, EEG bands unbalancing, the connection between two brain areas, and behavioral parameters.

In vivo effects on EEG: Comparison and hypothesis on correlates of hippocampal/cortical functionality

From the comparison of cortical and hippocampal EEG alterations, stand out three relevant differences\similarities: (1) while in the hippocampus the spread of depression development shows no steady state within the first 30 min of recording, the cortex displays somehow recovery, except for γ . (2) θ

drifts in opposite directions: in the hippocampus, it depresses increasingly with time, whereas in the cortex floats from brief mild depression to later highly significant growth. (3) a transitory significant increase of δ power in both areas immediately after the JWH-073 administration. This latter is potential of primary interest even if transitory and less intense as compared to points (1) and (2) and complexity beyond these δ fluctuations is suggested in Figures 5, 7: as frequently seen among all tests, EEG power amplitude alternates macroscopically in time, reflecting a pulsatile irregular alternate of hyper-excitability and depression. Figure 5 overall underlines this fluctuation of δ , mostly soon after JWH-073. Alterations of δ and θ like those described, are known to have different pathological significance. Physiologically, δ is the index of sleep depth, associated with deep stages 3 and 4 of NREM. The δ has the highest amplitude among brainwaves, with a frequency range from 0.01 to 0.1 Hz (Infra Slow Oscillations - ISO) and 0.5–4 Hz (δ). Cognitive performance and pathological condition both modulate δ and ISO. Generated mainly in the thalamus, spreads to the hippocampus and neocortex. δ increase inhibits GABA transmission (63) and T-type calcium channels (64)



EEG band	Control	5 min	10 min	15 min	20 min	25 min	30 min	2 h	24 h
Delta	0 ± 7.54	$12.85\pm8.19^{\ast}$	-0.45 ± 8.40	-0.36 ± 8.73	-9.79 ± 9.05	2.04 ± 11.12	6.34 ± 13.13	$-15.22 \pm 13.74^{\$}$	$19.23\pm14.02^{\$}$
Theta	0 ± 8.06	2.56 ± 7.07	$-15.67 \pm 7.30^{\$}$	$-14.38 \pm 6.63^{*}$	$-23.62\pm5.78^{\$}$	$-25.30\pm5.97^{\$}$	$-34.25\pm4.76^{\$}$	$33.68\pm5.84^{\$}$	$-14.9\pm6.62^*$
Alpha	0 ± 4.74	$-3.27\pm2.86^{\ast}$	$-14.34\pm4.32^{\ast}$	$-13.90 \pm 3.76^{*}$	$-13.80 \pm 4.38^{*}$	$-15.27 \pm 3.99^{\$}$	$-21.08\pm3.87^{\$}$	$-36.66 \pm 3.93^{\$}$	-7.60 ± 2.94
Beta	0 ± 11.76	7.505.23	0.17 ± 6.85	-4.43 ± 5.68	$-12.76 \pm 5.03^{*}$	$-14.05 \pm 4.46^{*}$	$-24.51\pm2.93^{\$}$	$-29.71 \pm 4.86^{\$}$	-8.49 ± 6.86
Gamma	0 ± 5.50	-10.63 ± 1.78	$-18.40 \pm 2.23^{\$}$	$-24.60 \pm 2.09^{\$}$	$-32.67 \pm 1.90^{\$}$	$-26.50 \pm 2.10^{\$}$	$-26.40 \pm 2.58^{\$}$	$-14.32 \pm 4.06^{*}$	$-26.34\pm4.62^{\$}$

TABLE 3 Quantitative EEG hippocampus.

Values relative to histograms in figure 10 with relative significance. Values correspond to $\% \pm$ s.e. Two-way ANOVA with Dunnet test used to compare the control EEG amplitude of groups before treatment with values after treatment. *p < 0.05; $^{\$}p < 0.01$.

impacting multiple brain functions. A wide array of brain disorders involves δ alterations (65) like schizophrenia, with loss of θ pattern with δ burst oscillation, as recorded in our experiment, followed by thalamocortical dysrhythmia. Driving in turn hippocampus and cortex (64), yielded psychotic-like effects as a consequence of described SCBs intoxication (66), by mean of inhibitory effect on excitatory synaptic transmission (29, 35, 67). After JWH-073, we recorded δ bursts, overcoming other EEG bands similarly to a condition known as " δ coma", a form of non-convulsive status epilepticus with high amplitude,

rhythmic δ activity (68) or arising the "extreme delta brush" disorder (69). The δ band was predominant for hours, conditions corresponding to deep drowsiness, comparable with "drug blackout", interspersed by brief hyper-reactivity or schizoid burst, observed at higher doses of SCBs (70, 71). This artificial " δ coma" is transitory and episodic, but in clinical prognosis, it heralds several months apart for damages ranging from severe memory impairment to generalized poor functional outcome. This cannot be underrated knowing that for a single mild dose, EEG alterations last almost 24 h after, so chronic sub-toxic



abuse is likely long-term consequential deficits. Different δ changes between the hippocampus and cortex are probably in consequence of the inhibitory activity of JWH-073 and/or of different connections with the thalamus. The previously described θ/δ unbalance may produce a peculiar area-dependent alteration, with hypersynchrony in the cortex and a shift of synchrony toward a loss of functionality in the hippocampus (61). Alteration of cortical θ/δ with hypersynchrony may reproduce loss of consciousness during sedation (72). In the hippocampus, the altered θ/δ ratio likely reflects *in vivo* NOR test outcome matching LTP failure for in vitro results. "Hippocampal θ rhythm" is a known physiological pattern crucial for memory and navigation (73). Hippocampal θ rhythm depends on projections on the medial septal area from the hypothalamus and brainstem, working as a relay for hippocampal CA3 neurons and involved in reward systems. Alterations of hippocampal θ and γ are markers of psychotic crises, switching from synchronized to chaotic. Neural synchronization regard functions like attention, memory, and sensory-motor integration and behavior. Acute smoked THC has been demonstrated capable of reproducing this loss of synchronism concurrently with psychotic alterations and loss of functionality, like above described (61), while changes have been noticed only for γ (74) and not caring about θ . Cannabinoid control via the endocannabinoid system of hippocampal functions is tightly correlated with control of network hippocampal oscillations, both for memory formation mechanisms and in the coding of spatial\non-spatial information (75). Rhythm disruption likely reflects the synaptic inhibition of glutamate and GABA (53, 67), with differential modulation after uneven CB1R area density. JWH-073 effect, combined a δ prevailing in the cortex with γ and θ depression in the hippocampus. This heralds a complex scenario of persistent deep cognitive depression, involving at the same time: the drive to exploration and knowledge (cortex), with the short-memory process (hippocampus). Being recognitionperception activity blurred (high δ) and hippocampal event storage failing (depressed γ and θ), the necessity of matching these to recreate a reality perception fails to de-cluttering the process information for a functional decision-making activity (76). Immobility observed could be a loss of drive to explore (messed up reality) and it seems to be associated with sensory impairment (5, 38) than drug interference with motor coordination. Spotting for epileptic traces from intersecting EEG data with simultaneous behavior, i.e., motor activity, converges to a temporary blackout than freezing or catalepsy for the seizure of index 1 or 2 (36).

Translation to human toxicology

Considering the remarkable fast change seen in spectrograms (Figures 4, 8), alterations are dramatic soon after 1 min from the administration. Smoking in humans as a way of administration worsens the negative effects of a sudden and massive EEG alteration. Therefore, chronic smoked mild doses of JWH-073, most likely result in cumulative toxicity and permanent subtle alterations overall for adolescent exposure (77). Indeed, differently from the well-known JWH-018, no fatal intoxication has been attributed to JWH-073 (78). The absence of epileptic events contributes to the perception of JWH-073 for safer abuse. In fact, the observed "spikes" in the cortex and hippocampus, originate from δ exacerbation, labeled as " δ epilepsy", involving almost exclusively δ band while typical of epileptic activity involves interictal faster frequencies domain (79).

Conclusion

Taken together these data depict a profile for JWH-073 as SCB capable at a sub-toxic dose (5) to induce immediate and long-lasting behavioral, mnemonic, and electrophysiological alterations. Consequential impairments like perceptive blackout, schizoid traits, non-convulsive epileptic bursts, and inconstant memory effectiveness may lead to hazardous compromising of demanding psychomotor activity (e.g., driving) (47). Thus, further attention will be required on the diffusion of SCs such as JWH-073 that are mistakenly perceived as "safe drugs" (80).

Data availability statement

The original contributions presented in the study are included in the article/Supplementary material, further inquiries can be directed to the corresponding author.

Ethics statement

Experimental protocols were approved by the Italian Ministry of Health (license n. 335/2016-PR and license n. 956/2020-PR) and by the Animal Welfare Body of the University of Ferrara.

Author contributions

MB and MM contributed to the conception and design of the study. MT, SB, RA, GC, BM, MS, and VC performed *in vivo* experiments. MB performed *in vitro* and *in vivo* electrophysiological studies. MB and MM wrote the manuscript. MT, SB, RA, GC, BM, and GS edited sections of the manuscript. MB, MT, SB, and VC performed statistical analysis. All authors contributed to the manuscript revision, and read and approved the submitted version.

Funding

This research has been funded by the Drug Policies Department, Presidency of the Council of Ministers, Italy (project: Effects of NPS: development of a multicentric research for the information enhancement of the Early Warning System to MM), by local funds from the University of Ferrara (FAR 2020 and FAR 2021 to MM).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fpsyt.2022.953909/full#supplementary-material

References

1. Eissenstat MA, Bell MR, D'Ambra TE, Alexander EJ, Daum SJ, Ackerman JH, et al. Aminoalkylindoles: structure-activity relationships of novel cannabinoid mimetics. *J Med Chem.* (1995) 38:3094e3105. doi: 10.1021/jm00016a013

2. Wiley JL, Compton DR, Dai D, Lainton JA, Phillips M, Huffman JW, et al. Structure-activity relationships of indole- and pyrrole-derived cannabinoids. *J Pharmacol Exp Ther.* (1998) 285:995–1004.

3. Aung MM, Griffin G, Huffman JW, Wu M, Keel C, Yang B, et al. Influence of the N-1 alkyl chain length of cannabimimetic indoles upon CB(1) and CB(2) receptor binding. *Drug Alcohol Depend.* (2000) 60:133–40. doi: 10.1016/S0376-8716(99)00152-0

4. Brents LK, Gallus-Zawada A, Radominska-Pandya A, Vasiljevik T, Prisinzano TE, Fantegrossi WE, et al. Monohydroxylated metabolites of the K2 synthetic cannabinoid JWH-073 retain intermediate to high cannabinoid 1 receptor (CB1R) affinity and exhibit neutral antagonist to partial agonist activity. *Biochem Pharmacol.* (2012) 83:952–61. doi: 10.1016/j.bcp.2012.01.004

5. Ossato A, Canazza I, Trapella C, Vincenzi F, De Luca MA, Rimondo C, et al. Effect of JWH-250, JWH-073 and their interaction on "tetrad", sensorimotor, neurological and neurochemical responses in mice. *Prog Neuropsychopharmacol Biol Psychiatry*. (2016) 67:31–50. doi: 10.1016/j.pnpbp.2016.01.007

6. Grotenhermen F. Pharmacokinetics and pharmacodynamics of cannabinoids. *Clin Pharmacokinet.* (2003) 42:327-60. doi: 10.2165/00003088-200342040-00003

7. Tai S, Fantegrossi WE. Pharmacological and toxicological effects of synthetic, cannabinoids and their metabolites. *Curr Top Behav Neurosci.* (2017) 32:249–62. doi: 10.1007/7854_2016_60

8. Lucas CJ, Galettis P, Schneider J. The pharmacokinetics and the pharmacodynamics of cannabinoids. *Br J Clin Pharmacol.* (2018) 84:2477-82. doi: 10.1111/bcp.13710

9. Lindigkeit R, Boehme A, Eiserloh I, Luebbecke M, Wiggermann M, Ernst L, et al. (2009). Spice: a never ending story? *For Sci Int.* 191:58–63. doi: 10.1016/j.forsciint.2009.06.008

10. Brents LK, Zimmerman SM, Saffell AR, Prather PL, Fantegrossi WE. Differential drug-drug interactions of the synthetic Cannabinoids JWH-018 and JWH- 073: implications for drug abuse liability and pain therapy. *J Pharmacol Exp Ther.* (2013) 346:350–61. doi: 10.1124/jpet.113.206003

11. Atwood BK, Lee D, Straiker A, Widlanski TS, Mackie K. CP47,497-C8 and JWH-073, commonly found in 'Spice' herbal blends, are potent and efficacious CB(1) cannabinoid receptor agonists. *Eur J Pharmacol.* (2011) 659:139-45. doi: 10.1016/j.ejphar.2011.01.066

12. Uchiyama N, Kikura-Hanajiri R, Ogata J, Goda Y. Chemical analysis of synthetic cannabinoids as designer drugs in herbal products. *Forensic Sci Int.* (2010) 198:31–8. doi: 10.1016/j.forsciint.2010.01.004

13. Auwärter V, Dresen S, Weinmann W, Müller M, Pütz M, Ferreirós N. 'Spice' and other herbal blends: harmless incense or cannabinoid designer drugs?. J Mass Spectromet. (2009) 44:832–7. doi: 10.1002/jms.1558

14. Papanti D, Schifano F, Botteon G, Bertossi F, Mannix J, Vidoni D, et al. "Spiceophrenia": a systematic overview of "spice"-related psychopathological issues and a case report. *Hum Psychopharmacol.* (2013) 28:379–89. doi: 10.1002/hup.2312

15. Hermanns-Clausen M, Kneisel S, Szabo B, Auwärter V. Acute toxicity due to the confirmed consumption of synthetic cannabinoids: clinical and laboratory findings. *Addiction*. (2013) 108:534–44. doi: 10.1111/j.1360-0443.2012.04078.x

16. Zawilska JB, Wojcieszak J. Spice/K2 drugs-more than innocent substitutes for marijuana. *Int J Neuropsychopharmacol.* (2014) 17:509-25. doi: 10.1017/S1461145713001247

17. Chung EY, Cha HJ, Min HK, Yun J. Pharmacology and adverse effects of new psychoactive substances: synthetic cannabinoid receptor agonists. *Arch Pharm Res.* (2021) 44:402–13. doi: 10.1007/s12272-021-01326-6

18. Martínez L, La Maida N, Papaseit E, Pérez-Mañá C, Poyatos L, Pellegrini M, et al. (2021). Acute pharmacological effects and oral fluid concentrations of the synthetic cannabinoids JWH-122 and JWH-210 in humans after self-administration: an observational study. *Front Pharmacol.* 12:705643. doi: 10.3389/fphar.2021.705643

19. Schifano F, Napoletano F, Chiappini S, Guirguis A, Corkery JM, Bonaccorso S, et al. New/emerging psychoactive substances and associated psychopathological consequences. *Psychol Med.* (2021) 51:30–42. doi: 10.1017/S0033291719001727

20. Müller H, Sperling W, Köhrmann M, Huttner H, Kornhuber J, Maler J. The synthetic cannabinoid Spice as a trigger for an acute exacerbation of cannabis induced recurrent psychotic episodes. *Schizophr Res.* (2010) 118:309–10. doi: 10.1016/j.schres.2009.12.001

21. Gurney SM, Scott KS, Kacinko SL, Presley BC, Logan BK. Pharmacology, toxicology, and adverse effects of synthetic cannabinoid drugs. *For Sci Rev.* (2014) 26:53–78.

22. Breivogel CS, Wells JR, Jonas A, Mistry AH, Gravley ML, Patel RM, et al. Comparison of the neurotoxic and seizure-inducing effects of synthetic and endogenous cannabinoids with Δ 9-tetrahydrocannabinol. *Cannabis Cannabinoid Res.* (2020) 5:32–41. doi: 10.1089/can.2019.0003

23. Marshell R, Kearney-Ramos T, Brents LK, Hyatt WS, Tai S, Prather PL, et al. *In vivo* effects of synthetic cannabinoids JWH-018 and JWH- 073 and phytocannabinoid Delta-THC in mice: inhalation versus intraperitoneal injection. *Pharmacol Biochem Behav.* (2014) 124C:40–7. doi: 10.1016/j.pbb.2014.05.010

24. Uttl L, Szczurowska E, Hájková K, Horsley RR, Štefková K, HloŽek T, et al. Behavioral and pharmacokinetic profile of indole-derived synthetic cannabinoids JWH-073 and JWH-210 as compared to the phytocannabinoid Δ 9-THC in rats. *Front Neurosci.* (2018) 12:703. doi: 10.3389/fnins.2018.00703

25. Cha HJ, Lee KW, Song MJ, Hyeon YJ, Hwang JY, Jang CG, et al. Dependence potential of the synthetic cannabinoids JWH-073, JWH-081, and JWH-210: *in vivo* and *in vitro* approaches. *Biomol Ther.* (2014) 22:363–9. doi: 10.4062/biomolther.2014.039

26. Ginsburg BC, Schulze DR, Hruba L, McMahon LR. JWH-018 and JWH-073: 8?-tetrahydrocannabinol-like discriminative stimulus effects in monkeys. J Pharmacol Exp Ther. (2012) 340:37–45. doi: 10.1124/jpet.111.187757

27. Gounder K, Dunuwille J, Dunne J, Lee J, Silbert P, Lawn N. The other side of the leaf: Seizures associated with synthetic cannabinoid use. *Epilepsy Behav*. (2020) 104(Pt A):106901. doi: 10.1016/j.yebeh.2020.106901

28. Croft RJ, Mackay AJ, Mills AT, Gruzelier JG. The relative contributions of ecstasy and cannabis to cognitive impairment. *Psychopharmacology.* (2001) 153:373–9. doi: 10.1007/s002130000591

29. Barbieri M, Ossato A, Canazza I, Trapella C, Borelli AC, Beggiato S, et al. Synthetic cannabinoid JWH-018 and its halogenated derivatives JWH-018-Cl and JWH-018-Br impair novel object recognition in mice: behavioral, electrophysiological and neurochemical evidence. *Neuropharmacology.* (2016) 109:254–69. doi: 10.1016/j.neuropharm.2016.06.027

30. Malaca S, Busardò FP, Nittari G, Sirignano A, Ricci G. Fourth generation of synthetic cannabinoid receptor agonists: A review on the latest insights. *Curr Pharm Des.* (2021) 27:1–15. doi: 10.2174/1381612827666211115170521

31. Giorgetti A, Busardò FP, Tittarelli R, Auwärter V, Giorgetti R. Post-mortem toxicology: a systematic review of death cases involving synthetic cannabinoid receptor agonists. *Front Psychiatry.* (2020) 11:464. doi: 10.3389/fpsyt.2020.00464

32. Kumar S, Baggi TR. Analytical methods for herbal products containing synthetic cannabinoids: a review. *For Chem.* (2021) 27:100396. doi: 10.1016/j.forc.2021.100396

33. Akgönüllü S, Battal D, Yalcin MS, Yavuz H, Denizli A. Rapid and sensitive detection of synthetic cannabinoids JWH-018, JWH-073 and their metabolites using molecularly imprinted polymer-coated QCM nanosensor in artificial saliva. *Microchem J.* (2019) 104454. doi: 10.1016/j.microc.2019.104454

34. Collins DR, Pertwee RG, Davies SN. The action of synthetic cannabinoids on the induction of long-term potentiation in the rat hippocampal slice. *Eur J Pharmacol.* (1994) 259:R7–8. doi: 10.1016/0014-2999(94)90666-1

35. Hoffman AF, Lycas MD, Kaczmarzyk JR, Spivak CE, Baumann MH, Lupica CR. Disruption of hippocampal synaptic transmission and long-term potentiation by psychoactive synthetic cannabinoid 'Spice' compounds: comparison with δ 9 - tetrahydrocannabinol. *Addict Biol.* (2017) 22:390–9. doi: 10.1111/adb.12334

36. Ossato A, Vigolo A, Trapella C, Seri C, Rimondo C, Serpelloni G, et al. JWH-018 impairs sensorimotor functions in mice. *Neuroscience*. (2015) 300:174–88. doi: 10.1016/j.neuroscience.2015.05.021

37. Marti M, Neri M, Bilel S, Paolo MD, Russa RL, Ossato A, et al. MDMA alone affects sensorimotor and prepulse inhibition responses in mice and rats: tips in the debate on potential MDMA unsafety in human activity. *Forensic Toxicol.* (2019) 37:132–44. doi: 10.1007/s11419-018-0444-7

38. Bilel S, Tirri M, Arfè R, Ossato A, Trapella C, Serpelloni G, et al. Novel halogenated synthetic cannabinoids impair sensorimotor functions in mice. *Neurotoxicology.* (2020) 76:17–32. doi: 10.1016/j.neuro.2019. 10.002

39. Arfè R, Bilel S, Tirri M, Frisoni P, Serpelloni G, Neri M, et al. Comparison of N-methyl-2-pyrrolidone (NMP) and the "date rape" drug GHB: behavioral toxicology in the mouse model. *Psychopharmacology*. (2021) 238:2275–95. doi: 10.1007/s00213-021-05852-5

40. Morini R, Mlinar B, Baccini G, Corradetti R. Enhanced hippocampal longterm potentiation following repeated MDMA treatment in Dark-Agouti rats. *Eur Neuropsychopharmacol.* (2011) 21:80–91. doi: 10.1016/j.euroneuro.2010.07.007

41. Spencer SS, Kim J, Spencer DD. Ictal spikes: a marker of specific hippocampal cell loss. *Electroencephalogr Clin Neurophysiol.* (1992) 83:104–11. doi: 10.1016/0013-4694(92)90023-B

42. Racine RJ. Modification of seizure activity by electrical stimulation. II. Motor seizure. *Electroencephalogr Clin Neurophysiol.* (1972) 32:281–94. doi: 10.1016/0013-4694(72)90177-0

43. Benoit O, Daurat A, Prado J. Slow (0.7–2 Hz) and fast (2–4 Hz) delta components are differently correlated to theta, alpha and beta frequency bands during NREM sleep. *Clin Neurophysiol.* (2000) 111:2103–6. doi: 10.1016/S1388-2457(00)00470-3

44. White A, Williams PA, Hellier JL, Clark S, Dudek FE, Staley KJ. EEG spike activity precedes epilepsy after kainate-induced status epilepticus. *Epilepsia*. (2010) 51:371–83. doi: 10.1111/j.1528-1167.2009.02339.x

45. Misulis K. Atlas of EEG, Seizure Semiology, and Management. Oxford, UK: Oxford University Press (2013).

46. Grinenko O, Li J, Mosher JC, Wang IZ, Bulacio JC, Gonzalez-Martinez J, et al. A fingerprint of the epileptogenic zone in human epilepsies. *Brain.* (2018) 141:117–31. doi: 10.1093/brain/awx306

47. Musshoff F, Madea B, Kernbach-Wighton G, Bicker W, Kneisel S, Hutter M, et al. Driving under the influence of synthetic cannabinoids ("Spice"): a case series. *Int J Legal Med.* (2014) 128:59–64 doi: 10.1007/s00414-013-0864-1

48. Theunissen EL, Reckweg JT, Hutten NRPW, Kuypers KPC, Toennes SW, Neukamm MA, et al. Intoxication by a synthetic cannabinoid (JWH-018) causes cognitive and psychomotor impairment in recreational cannabis users. *Pharmacol Biochem Behav.* (2021) 202:173118. doi: 10.1016/j.pbb.2021.173118

49. Ito S, Deyama S, Domoto M, Zhang T, Sasase H, Fukao A, et al. Effects of the synthetic cannabinoid 5F-AMB on anxiety and recognition memory in mice. *Psychopharmacology.* (2019) 236:2235–42. doi: 10.1007/s00213-019-05222-2

50. Tai S, Hyatt WS, Gu C, Franks LN, Vasiljevik T, Brents LK, et al. Repeated administration of phytocannabinoid $\delta(9)$ -THC or synthetic cannabinoids JWH-018 and JWH-073 induces tolerance to hypothermia but not locomotor suppression in mice, and reduces CB1 receptor expression and function in a brain region-specific manner. *Pharmacol Res.* (2015) 102:22–32. doi: 10.1016/j.phrs.2015.09.006

51. Baltaci SB, Mogulkoc R, Baltaci AK. Molecular mechanisms of early and late LTP. *Neurochem Res.* (2019) 44:281–96. doi: 10.1007/s11064-018-2695-4

52. Robbe D, Montgomery SM, Thome A, Rueda-Orozco PE, McNaughton BL, Buzsaki G. Cannabinoids reveal importance of spike timing coordination in hippocampal function. *Nat Neurosci.* (2006) 9:1526–33. doi: 10.1038/nn1801

53. Schlicker E, Kathmann M. Modulation of transmitter release via presynaptic cannabinoid receptors. *Trends Pharmacol Sci.* (2001) 22:565–72. doi: 10.1016/S0165-6147(00)01805-8

54. Ilan AB, Gevins A, Coleman M, ElSohly MA, de Wit H. Neurophysiological and subjective profile of marijuana with varying concentrations of cannabinoids. *Behav Pharmacol.* (2005) 16:487–96. doi: 10.1097/00008877-200509000-00023

55. Matsuzaki M, Casella GA, Ratner M. delta 9-Tetrahydrocannabinol: EEG changes, bradycardia and hypothermia in the rhesus monkey. *Brain Res Bull.* (1987) 19:223–9. doi: 10.1016/0361-9230(87)90087-6

56. Buonamici M, Young GA, Khazan N. Effects of acute δ9-THC administration of EEG and EEG power spectra in the rat. *Neuropharmacology.* (1982) 21:825–9. doi: 10.1016/0028-3908(82)90071-5

57. Uchiyama N, Kikura-Hanajiri R, Matsumoto N, Huang ZL, Goda Y, Urade Y. Effects of synthetic cannabinoids on electroencephalogram power spectra in rats, *For Sci Int.* (2012) 215:179–83. doi: 10.1016/j.forsciint.2011.05.005

58. Kucewicz MT, Tricklebank MD, Bogacz R, Jones MW. Dysfunctional prefrontal cortical network activity and interactions following cannabinoid receptor activation. *J Neurosci.* (2011) 31:15560–8. doi: 10.1523/JNEUROSCI.2970-11.2011

59. Raver SM, Haughwout SP, Keller A. Adolescent cannabinoid exposure permanently suppresses cortical oscillations in adult mice. *Neuropsychopharmacology.* (2013) 38:2338–47. doi: 10.1038/npp. 2013.164

60. Hajós M, Hoffmann WE, Kocsis B. Activation of cannabinoid-1 receptors disrupts sensory gating and neuronal oscillation: relevance to

schizophrenia. Biol Psychiatry. (2008) 63:1075-83. doi: 10.1016/j.biopsych.2007. 12.005

61. Skosnik PD, Cortes-Briones JA, Hajós M. It's all in the rhythm: the role of cannabinoids in neural oscillations and psychosis. *Biol Psychiatry*. (2016) 79:568–77. doi: 10.1016/j.biopsych.2015.12.011

62. Skosnik PD, Hajós M, Cortes-Briones JA, Edwards CR, Pittman BP, Hoffmann WE, et al. Cannabinoid receptor- mediated disruption of sensory gating and neural oscillations: a translational study in rats and humans. *Neuropharmacology.* (2018) 135:412–23. doi: 10.1016/j.neuropharm.2018.03.036

63. Hobson JA, Pace-Schott EF. The cognitive neuroscience of sleep: neuronal systems, consciousness and learning. *Nat Rev Neurosci.* (2002) 3:679–93. doi: 10.1038/nrn915

64. Zhang Y, Llinas RR, Lisman JE. Inhibition of NMDARs in the nucleus reticularis of the thalamus produces delta frequency bursting. *Front Neural Circuits.* (2009) 3:20. doi: 10.3389/neuro.04.020.2009

65. Sutter R, Kaplan PW. Electroencephalographic patterns in coma: when things slow down. *Epileptologie*. (2012) 29:201–9.

66. Fantegrossi WE, Wilson CD, Berquist MD. Pro-psychotic effects of synthetic cannabinoids: interactions with central dopamine, serotonin, and glutamate systems. *Drug Metab Rev.* (2018) 50:65–73. doi: 10.1080/03602532.2018.14 28343

67. Sherif MA, Cortes-Briones JA, Ranganathan M, Skosnik PD. Cannabinoidglutamate interactions and neural oscillations: implications for psychosis. *Eur J Neurosci.* (2018) 48:2890–902. doi: 10.1111/ejn.13800

68. Kirkpatrick MP, Clarke CD, Sonmezturk HH, Abou-Khalil B. Rhythmic delta activity represents a form of nonconvulsive status epilepticus in anti-NMDA receptor antibody encephalitis. *Epilepsy Behav.* (2011) 20:392–4. doi: 10.1016/j.yebeh.2010.11.020

69. Schmitt SE, Pargeon K, Frechette ES, Hirsch LJ, Dalmau J, Friedman D. Extreme delta brush: a unique EEG pattern in adults with anti-NMDA receptor encephalitis. *Neurology*. (2012) 79:1094–100. doi:10.1212/WNL.0b013e3182698cd8

70. Vigolo A, Ossato A, Trapella C, Vincenzi F, Rimondo C, Seri C, et al. Novel halogenated derivates of JWH-018: Behavioral and binding studies in mice. *Neuropharmacology*. (2015) 95:68–82. doi: 10.1016/j.neuropharm.2015.02.008

71. Canazza I, Ossato A, Trapella C, Fantinati A, De Luca MA, Margiani G, et al. Effect of the novel synthetic cannabinoids AKB48 and 5F-AKB48 on "tetrad", sensorimotor, neurological and neurochemical responses in mice. *In vitro* and *in vivo* pharmacological studies. *Psychopharmacology.* (2016) 233:3685–709. doi: 10.1007/s00213-016-4402-y

72. Supp GG, Siegel M, Hipp JF, Engel AK. Cortical hypersynchrony predicts breakdown of sensory processing during loss of consciousness. *Curr Biol.* (2011) 21:1988–93. doi: 10.1016/j.cub.2011.10.017

73. Nuñez A, Buño W. The theta rhythm of the hippocampus: from neuronal and circuit mechanisms to behavior. *Front Cell Neurosci.* (2021) 15:649262. doi: 10.3389/fncel.2021.649262

74. Cortes-Briones J, Skosnik PD, Mathalon D, Cahill J, Pittman B, Williams A, et al. 89-THC disrupts gamma (γ)-band neural oscillations in humans. *Neuropsychopharmacology*. (2015) 40:2124–34. doi: 10.1038/npp.2015.53

75. Robledo-Menendez A, Vella M, Grandes P, Soria-Gomez E. Cannabinoid control of hippocampal functions: the where matters. *FEBS J.* (2022) 289:2162–75. doi: 10.1111/febs.15907

76. Tavares LCS, Tort ABL. Hippocampal-prefrontal interactions during spatial decision- making. *Hippocampus*. (2022) 32:38–54. doi: 10.1002/hipo.23394

77. Hernandez CM, Orsini CA, Blaes SL, Bizon JL, Febo M, Bruijnzeel AW, et al. Effects of repeated adolescent exposure to cannabis smoke on cognitive outcomes in adulthood. *J Psychopharmacol.* (2021) 35:848–63. doi: 10.1177/0269881120965931

78. World Health Organization. Expert Committee on Drug Dependence Thirtyeight Meeting. JWH-073 Critical Review Report Agenda item 4.11 Geneva, 14–18 November 2016. Geneva (2016).

79. De Curtis M, Avanzini G. Interictal spikes in focal epileptogenesis. *Prog Neurobiol.* (2001) 63:541–67. doi: 10.1016/S0301-0082(00)00026-5

80. Maida N, Papaseit E, Martínez L, Pérez-Mañá C, Poyatos L, Pellegrini M, et al. Acute pharmacological effects and oral fluid biomarkers of the synthetic cannabinoid UR-144 and THC in recreational users. *Biology.* (2021) 10:257. doi:10.3390/biology10040257