

MDPI

Article

Imaging Severity COVID-19 Assessment in Vaccinated and Unvaccinated Patients: Comparison of the Different Variants in a High Volume Italian Reference Center

Vincenza Granata ¹, Roberta Fusco ^{2,*}, Alberta Villanacci ³, Simona Magliocchetti ⁴, Fabrizio Urraro ⁴, Nardi Tetaj ⁵, Luisa Marchioni ⁵, Fabrizio Albarello ³, Paolo Campioni ³, Massimo Cristofaro ³, Federica Di Stefano ³, Nicoletta Fusco ³, Ada Petrone ³, Vincenzo Schininà ³, Francesca Grassi ^{4,6}, Enrico Girardi ⁷ and Stefania Ianniello ³

- Division of Radiology, Istituto Nazionale Tumori IRCCS Fondazione Pascale—IRCCS di Napoli, 80131 Naples, Italy; v.granata@istitutotumori.na.it
- Medical Oncology Division, Igea SpA, 80013 Napoli, Italy
- Diagnostic Imaging of Infectious Diseases, National Institute for Infectious Diseases Lazzaro Spallanzani IRCCS, 00149 Rome, Italy; alberta.villanacci@inmi.it (A.V.); fabrizio.albarello@inmi.it (F.A.); paolo.campioni@inmi.it (P.C.); massimo.cristofaro@inmi.it (M.C.); federica.distefano@inmi.it (F.D.S.); nicoletta.fusco@inmi.it (N.F.); ada.petrone@inmi.it (A.P.); vincenzo.schinina@inmi.it (V.S.); stefania.ianniello@inmi.it (S.I.)
- Division of Radiology, Università degli Studi della Campania Luigi Vanvitelli, 80128 Naples, Italy; simona.magliocchetti@unicampania.it (S.M.); fabrizio.urraro@unicampania.it (F.U.); francescagrassi1996@gmail.com (F.G.)
- Intensive Care Unit, National Institute for Infectious Diseases Lazzaro Spallanzani IRCCS, 00149 Rome, Italy; nardi.tetaj@inmi.it (N.T.); luisa.marchioni@inmi.it (L.M.)
- Italian Society of Medical and Interventional Radiology (SIRM), SIRM Foundation, Via Della Signora 2, 20122 Milan, Italy
- Department of Epidemiology and Research, National Institute for Infectious Diseases Lazzaro Spallanzani IRCCS, 00149 Rome, Italy; enrico.girardi@inmi.it
- * Correspondence: r.fusco@igeamedical.com

Abstract: Purpose: To analyze the vaccine effect by comparing five groups: unvaccinated patients with Alpha variant, unvaccinated patients with Delta variant, vaccinated patients with Delta variant, unvaccinated patients with Omicron variant, and vaccinated patients with Omicron variant, assessing the "gravity" of COVID-19 pulmonary involvement, based on CT findings in critically ill patients admitted to Intensive Care Unit (ICU). Methods: Patients were selected by ICU database considering the period from December 2021 to 23 March 2022, according to the following inclusion criteria: patients with proven Omicron variant COVID-19 infection with known COVID-19 vaccination with at least two doses and with chest Computed Tomography (CT) study during ICU hospitalization. Wee also evaluated the ICU database considering the period from March 2020 to December 2021, to select unvaccinated consecutive patients with Alpha variant, subjected to CT study, consecutive unvaccinated and vaccinated patients with Delta variant, subjected to CT study, and, consecutive unvaccinated patients with Omicron variant, subjected to CT study. CT images were evaluated qualitatively using a severity score scale of 5 levels (none involvement, mild: ≤25% of involvement, moderate: 26-50% of involvement, severe: 51-75% of involvement, and critical involvement: 76-100%) and quantitatively, using the Philips IntelliSpace Portal clinical application CT COPD computer tool. For each patient the lung volumetry was performed identifying the percentage value of aerated residual lung volume. Non-parametric tests for continuous and categorical variables were performed to assess statistically significant differences among groups. Results: The patient study group was composed of 13 vaccinated patients affected by the Omicron variant (Omicron V). As control groups we identified: 20 unvaccinated patients with Alpha variant (Alpha NV); 20 unvaccinated patients with Delta variant (Delta NV); 18 vaccinated patients with Delta variant (Delta V); and 20 unvaccinated patients affected by the Omicron variant (Omicron NV). No differences between the groups under examination were found (p value > 0.05 at Chi square test) in terms of risk factors (age, cardiovascular diseases, diabetes, immunosuppression, chronic kidney, cardiac,



Citation: Granata, V.; Fusco, R.; Villanacci, A.; Magliocchetti, S.; Urraro, F.; Tetaj, N.; Marchioni, L.; Albarello, F.; Campioni, P.; Cristofaro, M.; et al. Imaging Severity COVID-19 Assessment in Vaccinated and Unvaccinated Patients: Comparison of the Different Variants in a High Volume Italian Reference Center. *J. Pers. Med.* 2022, 12, 955. https:// doi.org/10.3390/jpm12060955

Academic Editor: Franco M. Buonaguro

Received: 28 April 2022 Accepted: 9 June 2022 Published: 10 June 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

I. Pers. Med. 2022, 12, 955 2 of 21

pulmonary, neurologic, and liver disease, etc.). A different median value of aerated residual lung volume was observed in the Delta variant groups: median value of aerated residual lung volume was 46.70% in unvaccinated patients compared to 67.10% in vaccinated patients. In addition, in patients with Delta variant every other extracted volume by automatic tool showed a statistically significant difference between vaccinated and unvaccinated group. Statistically significant differences were observed for each extracted volume by automatic tool between unvaccinated patients affected by Alpha variant and vaccinated patients affected by Delta variant of COVID-19. Good statistically significant correlations among volumes extracted by automatic tool for each lung lobe and overall radiological severity score were obtained (ICC range 0.71-0.86). GGO was the main sign of COVID-19 lesions on CT images found in 87 of the 91 (95.6%) patients. No statistically significant differences were observed in CT findings (ground glass opacities (GGO), consolidation or crazy paving sign) among patient groups. Conclusion: In our study, we showed that in critically ill patients no difference were observed in terms of severity of disease or exitus, between unvaccinated and vaccinated patients. The only statistically significant differences were observed, with regard to the severity of COVID-19 pulmonary parenchymal involvement, between unvaccinated patients affected by Alpha variant and vaccinated patients affected by Delta variant, and between unvaccinated patients with Delta variant and vaccinated patients with Delta variant.

Keywords: COVID-19; vaccination; Computed Tomography

1. Introduction

Over two years after the first described SARS-CoV-2 patient, the COVID-19 pandemic is still ongoing, with many countries undergoing new infection waves [1–12]. Extensive vaccination promotion is underway all over the world, although with extremely variable levels of population coverage [1,13-24]. In addition, the pandemic perseveres with the appearance of new variants that could compromise diagnostic tests and vaccine efficacy. Developing evidence has demonstrated that these variants are able to evade the action of neutralizing antibodies [25-43]. Evidence of declining vaccine immunity over time has also arisen: following the second dose, there is a substantial decline in efficacy against symptomatic infection; from a peak of ~90% in the weeks immediately following to a much lower 50–80% six months after vaccination [44–48]. Consequently, several nations are proposing booster vaccinations. Data from these countries have proven the benefit of a booster dose in reducing symptomatic infection and offering a significant decrease in critical outcomes [49-54]. Moreover, the protection level offered by previous SARS-CoV-2 infection, both in terms of infection and disease severity and, therefore, of outcome, is still unclear [55–61]. In this scenario, the main essential element leading to the evolution of SARS-CoV-2 infection is the interaction with the host's immune system. However, there is a need to understand how the new variants can lead to severe forms of the disease, as well as how the time elapsed since vaccination can impact the outcome. An assessment of disease severity requires tools that can objectify the data to reduce the variability between patients due to qualitative evaluation. As to the "gravity assessment" of COVID-19 infection and evaluation of pulmonary parenchymal involvement, several scores have been proposed [62,63]. The main goal of these tools is to establish a well-defined strategy for evaluation of the airways and lungs of COVID-19 positive patients from Computed Tomography (CT) scans, including detected abnormalities [64–74]. Their identification and the volumetric quantification may allow an easier classification in terms of gravity, extent and progression of the disease. Moreover, this may provide a high-impact tool to enhance awareness of the severity of COVID-19 pneumonia [75–90].

In this retrospective cohort study, we aim to analyze the vaccine effect by comparing five groups: (a) unvaccinated patients with Alpha variant; (b) unvaccinated patients with Delta variant; (c) vaccinated patients with Delta variant; (d) unvaccinated patients with Omicron variant; and (e) vaccinated patients with Omicron variant, assessing the

I. Pers. Med. 2022, 12, 955 3 of 21

"gravity" of COVID-19 pulmonary involvement, based on CT findings in critically ill patients admitted to Intensive Care Unit (ICU).

2. Materials and Methods

2.1. Patient Characteristics

The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Institutional Ethics Committee of IRCCS L. Spallanzani. Data acquisition and analysis were performed in compliance with protocols approved by the Ethical Committee of the National Institute for Infectious Diseases IRCCS Lazzaro Spallanzani, Rome, Italy (ethical approval number 164, 26 June 2020). The Local Ethical Committee board renounced patient informed consent, considering the ongoing epidemic emergency.

Patients were selected from the Intensive Care Unit (ICU) database considering the period from December 2021 to 23 March 2022, having COVID-19 infection variant sequencing, according to the following inclusion criteria: (1) patients with proven Omicron variant COVID-19 infection; (2) patients with known COVID-19 vaccination with at least two doses; (3) patients with chest CT study during ICU hospitalization. The exclusion criteria were: (1) no CT study, (2) patients with no data on COVID-19 vaccination status.

We also evaluated the ICU database considering the period from March 2020 to December 2021, to select unvaccinated consecutive patients with Alpha variant, subjected to CT study; consecutive unvaccinated patients with Delta variant, subjected to CT study; consecutive vaccinated patients with Delta variant, subjected to CT study; consecutive unvaccinated patients with Omicron variant, subjected to CT study.

2.2. CT Technique

Chest CT scan was performed with 128 slices using Incisive Philips CT scanners (Amsterdam, The Netherlands). CT examinations were performed with the patient in the supine position in breath-hold, and inspiration using a standard dose protocol, without contrast intravenous injection. The scanning range was from the apex to the base of the lungs. The tube voltage and the current tube were 120 kV and 100–200 mA (and if applicable, using z-axis tube current modulation), respectively. All data were reconstructed with a 0.6–1.0 mm increment. The matrix was 512 mm \times 512 mm. Images were reconstructed using a sharp reconstruction kernel for parenchyma evaluation and hard reconstruction kernel for other lung evaluation. All data were reconstructed with a 0.6–1.0 mm increment. Multiplanar reconstruction (MPR) was also obtained.

2.3. CT Post Processing

DICOM data were transferred into a PACS workstation and CT images were evaluated using the Philips IntelliSpace Portal clinical application CT COPD (Philips Eindihoven, The Netherlands) computer tool.

Philips IntelliSpace Portal clinical application CT COPD software is a CE-marked medical device designed to quantify pulmonary emphysema in patients with chronic obstructive pulmonary disease. The tool provides segmentation of the lungs and of the airway tree. Moreover, the tool helps visualize and quantify the destructive process of diffuse lung disease (e.g., emphysema), providing a guided workflow for airway analysis, reviewing and measuring airway lumen, and assessing trapped air. Compared to others tools, it allows assessment consolidation. For each patient the lung volumetry was performed identifying the percentage value of aerated residual lung volume, and for each lung lobe: right upper lobe volume, right lower lobe volume, medium lobe volume, left upper lobe volume, left lower lobe volume (Figures 1 and 2).

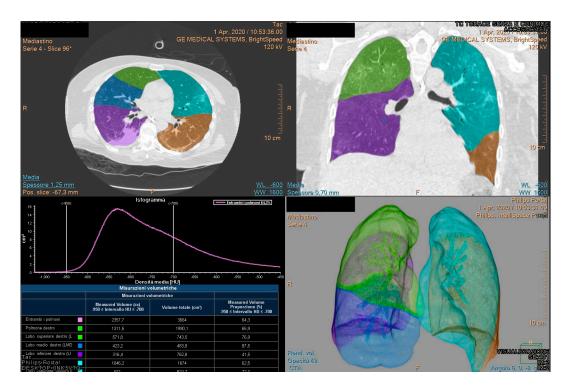


Figure 1. Example 1 of Quantitative assessment of COVID-19 pulmonary parenchymal involvement by automatic tool.

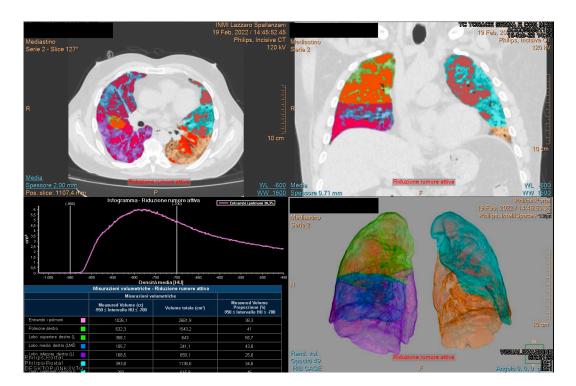


Figure 2. Example 2 of Quantitative assessment of COVID-19 pulmonary parenchymal involvement by automatic tool.

Disease severity was assessed by considering the percentage of aerated residual lung volume: patients with lower aerated residual lung volume were considered more compromised.

J. Pers. Med. 2022, 12, 955 5 of 21

2.4. Radiologists' Analysis

Radiologists attributed for each lung lobe (right upper and lower lobe, medium lobe, left upper and lower lobe) a severity score using a scale of 5 levels (no involvement, mild: \leq 25% of involvement, moderate: 26–50% of involvement, severe: 51–75% of involvement and critic involvement: 76–100%) as reported in Li et al. [91]. Moreover, an overall radiological severity score was obtained summing the scores for each lung lobe and then considering a low severity \leq 5, mild severity 6–10, moderate 11–15, severe 16–20 and critical 21–25. Two radiologists with more than 10 years of thoracic-imaging analysis experience evaluated the severity of images in a double-blind manner. Another, more experienced, radiologist resolved any disagreement between the two radiologists determining a radiological consensus.

In addition, a qualitative assessment including the evaluation of the following CT findings, ground glass opacities (GGOs), consolidation and crazy paving, was defined according to the Fleischner Society glossary [92].

2.5. Statistical Analysis

Continuous data were expressed in terms of median values and range. Chi square test, Mann Whitney test and Kruskal Wallis test were used to verify differences among groups. Intraclass correlation coefficient was used to analyze the correlations and variability among quantitative measurements generated by automatic tool and radiological severity score.

Bonferroni correction was considered for multiple comparisons.

p value < 0.05 was considered significant for all tests.

The statistical analyses were performed using the Statistics and Machine Toolbox of MATLAB R2021b (MathWorks, Natick, MA, USA).

3. Results

According to the inclusion and exclusion criteria, the patient study group was composed of 13 vaccinated patients affected by the Omicron variant (Omicron V). As control groups we identified: 20 unvaccinated patients with Alpha variant (Alpha NV); 20 unvaccinated patients with Delta variant (Delta NV); 18 vaccinated patients with Delta variant (Delta V); and 20 unvaccinated patients affected by the Omicron variant (Omicron NV). Mean age and sex distribution for each group is reported in Table 1.

Table 1. Demographic and CT findings of Patients in the Study.

Characteristic	Alpha Variant n = 20	Unvaccinated Delta Variant n = 20	Unvaccinated Delta Variant n = 18	Unvaccinated Omicron Variant n = 20	Vaccinated Omicron Variant n = 13	p Value
Age (y)						
Mean	62	58	64	69	75	0.07
Range	43–78	37–83	35–87	42–88	55–94	0.07
Sex, no. (%) of p	atients					
Male	14	17	15	13	12	0.43
Female	6	3	3	7	1	0.43
CT Findings						
GGO	19	20	16	19	13	0.89
Crazy Paving	17	20	14	16	11	0.10
Consolidation	15	17	11	16	11	0.70
Exitus	5	5	6	4	5	0.95

Note. p value was evaluated for continuous variable by Mann Whitney test and by Chi square test for categorical variables.

No differences between the groups under examination were found (p value > 0.05 at Chi square test) in terms of risk factors (cardiovascular diseases, diabetes, immunosuppression, chronic kidney, cardiac, pulmonary, neurologic, and liver disease, etc.).

The patient distribution with median value of aerated residual lung volume for each subgroup is reported in Table 2 and Figure 3.

Table 2. Patient distribution and median value of aerated residual lung volume for each subgroup

		Unvaccinated	Vaccinated with 2 Doses	Vaccinated with 3 Doses	p Value
Patients with Alpha Variant		20	0	0	0.001
Patients with Delta variant	Number of patients	20	16	2	0.001
Patients with Omicron	-	20	8	5	
Patients with Alpha Variant	Median value of (range) of	39.95 (19.40–67.50)	-	-	0.05
Patients with Delta variant	Aerated residual lung volume [%]	46.7 (13.60–75.60)	67.10 (17.10–89.80)	52.00 (19.40–84.50)	0.00
Patients with Omicron		48.35 (8.20–83.30)	38.30 (18.90–73.30)	61.9 (31.60–73.60)	

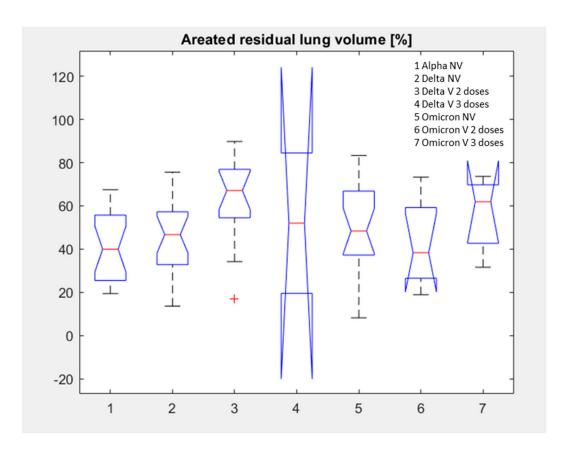


Figure 3. Distribution of aerated residual lung volume for each subgroup.

No statistically significant differences were observed between unvaccinated and vaccinated patients with Omicron variant for aerated residual lung volume, right upper lobe volume, right lower lobe volume, medium lobe volume, left upper lobe volume, or left lower lobe volume in percentage values: p value > 0.05 with Kruskal Wallis test (see boxplots in Figure 4, Table 3).

J. Pers. Med. **2022**, 12, 955 7 of 21

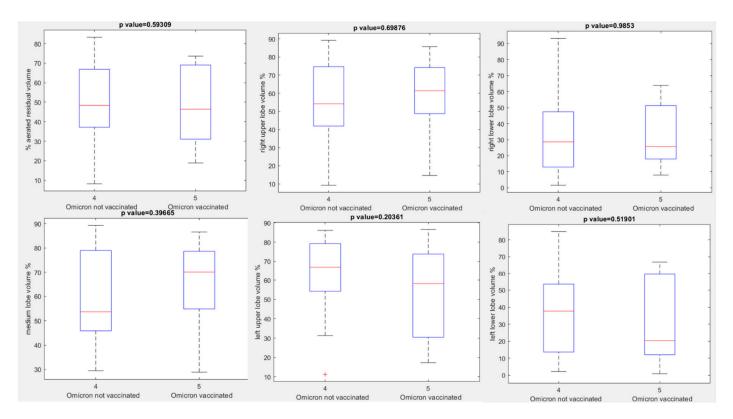


Figure 4. Boxplots of extracted volumes by automatic tool between vaccinated and unvaccinated patients affected by Omicron Variant of COVID-19.

Table 3. Median values of extracted volumes by automatic tool patients affected by Alpha, Delta or Omicron Variant of COVID-19 grouped by vaccination or no vaccination.

	Aerated Residual Volume %	Right Upper Lobe Volume %	Right Lower Lobe Volume %	Medium Lobe Volume %	Left Upper Lobe Volume %	Left Lower Lobe Volume %
Alpha	39.95	47.30	26.00	64.40	55.00	25.05
Unvaccinated	39.95	47.30	26.00	64.40	55.00	25.05
Delta	55.25	56.2	58.35	72.9	32.75	56
Unvaccinated	46.70	39.20	51.30	60.15	23.45	46.65
Vaccinated	67.10	66.50	71.55	83.50	57.00	66.80
Omicron	46.4	46.8	59	68.4	26.9	50
Unvaccinated	48.35	42.2	54.2	53.65	28.65	51.65
Vaccinated	46.4	49.8	61.4	70.1	25.7	45.1
p value at Kruskal Wallis test	0.03	0.06	0.06	0.04	0.004	0.12

A different median value of aerated residual lung volume was observed in the Delta variant groups: median value of aerated residual lung volume was 46.70% in unvaccinated patients compared to 67.10% in vaccinated patients (p value = 0.01 with Kruskal Wallis test). In addition, in patients with Delta variant every other extracted volume by automatic tool showed a statistically significant difference between vaccinated and unvaccinated group (see boxplots in Figure 5, Table 3): p value at Kruskal Wallis test = 0.02, 0.02, 0.02, 0.03, 0.03,

respectively, for percentage values of right upper lobe volume, right lower lobe volume, medium lobe volume, left upper lobe volume and left lower lobe volume.

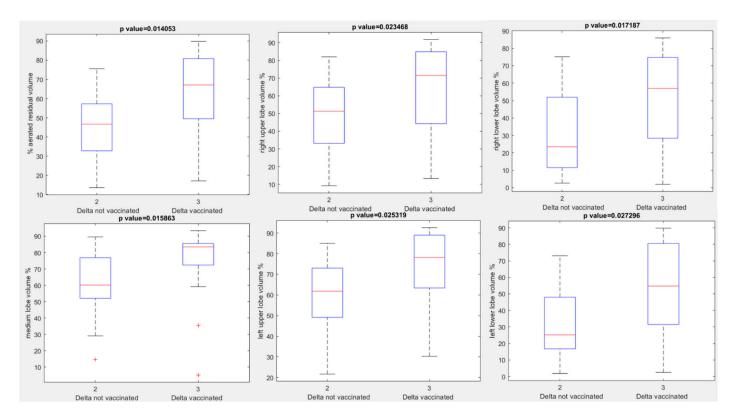


Figure 5. Boxplots of extracted volumes by automatic tool between vaccinated and unvaccinated patients affected by Delta Variant of COVID-19.

No statistically significant differences were observed in terms of aerated residual lung volume among vaccinated or unvaccinated patients with Delta and vaccinated or unvaccinated patients with Omicron variant (p value > 0.05 with Kruskal Wallis test, Figure 6). The only statistically significant differences were observed between vaccinated patients with Delta variant and vaccinated patients with Omicron variant for the right upper lobe volume, medium lobe volume and left lower lobe volume with a p value at Kruskal Wallis test, respectively, of 0.04, 0.03 and 0.01 (Figure 7) and between vaccinated patients with Delta variant and unvaccinated patients with Omicron variant for the right upper lobe volume and medium lobe volume with a p value for Kruskal Wallis test, respectively, of 0.03 and 0.02 (Figure 8).

No difference was observed in terms of each extracted volumes by automatic tool (aerated residual lung volume, right upper lobe volume, right lower lobe volume, medium lobe volume, left upper lobe volume, left lower lobe volume) between unvaccinated patients with the Alpha variant versus vaccinated or unvaccinated patients with the Omicron variant (p value > 0.05 for Kruskal Wallis test).

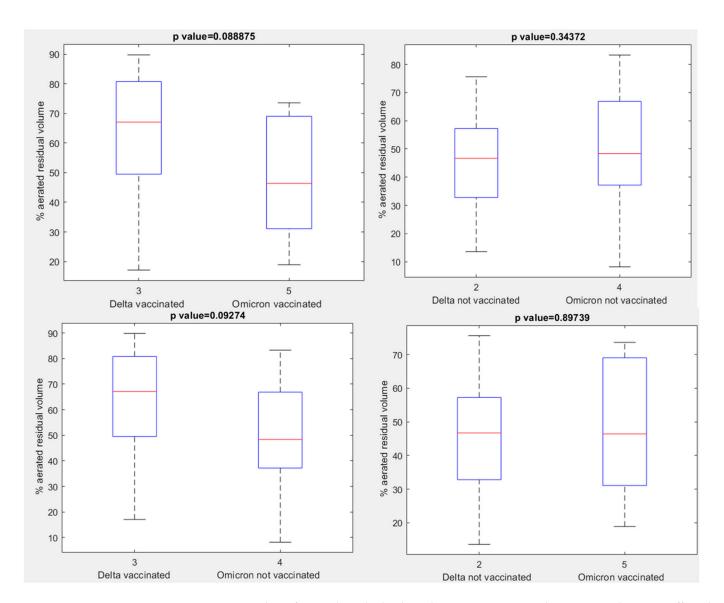


Figure 6. Boxplots of aerated residual volume between unvaccinated or vaccinated patients affected by Delta variant and vaccinated or un-vaccinated patients affected by Omicron variant.

In addition, no difference was observed in terms of each extracted volume by automatic tool between unvaccinated patients with the Alpha variant versus unvaccinated patients with the Delta variant (p value > 0.05 for Kruskal Wallis test). Instead, statistically significant differences were observed for each extracted volume by automatic tool between unvaccinated patients affected by Alpha variant and vaccinated patients affected by Delta variant of COVID-19: p value for Kruskal Wallis test = 0.003, 0.01, 0.01, 0.01, 0.001, 0.01, respectively, for percentage values of aerated residual lung volume, right upper lobe volume, right lower lobe volume, medium lobe volume, left upper lobe volume and left lower lobe volume (see boxplots in Figure 9).

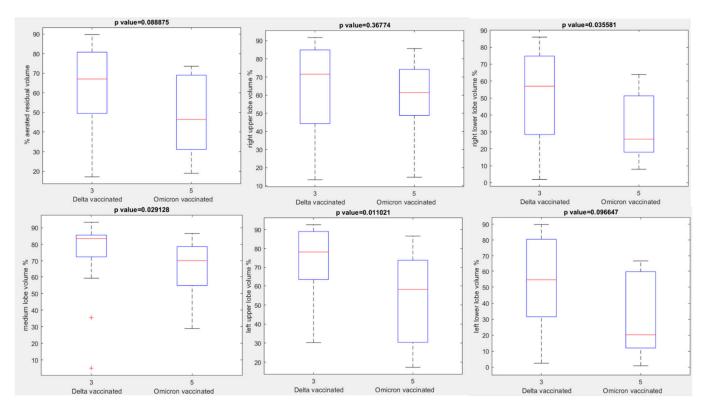


Figure 7. Boxplots of extracted volumes by automatic tool between vaccinated patients affected by Delta Variant of COVID-19 and vaccinated patients affected by Omicron Variant of COVID-19.

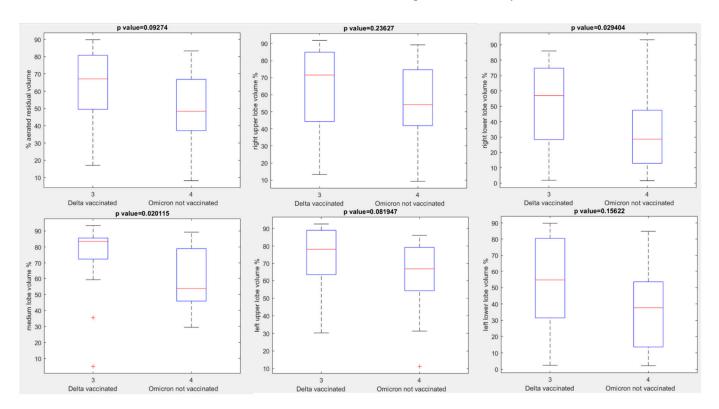


Figure 8. Boxplots of extracted volumes by automatic tool between vaccinated patients affected by Delta Variant of COVID-19 and unvaccinated patients affected by Omicron Variant of COVID-19.

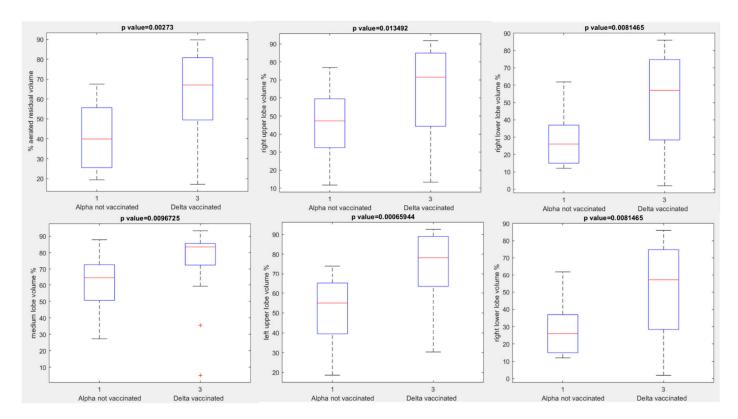


Figure 9. Boxplots of extracted volumes by automatic tool between unvaccinated patients affected by Alpha variant and vaccinated patients affected by Delta variant of COVID-19.

The highest differences were observed in median value of aerated residual lung volume (39.95% versus 67.10%) in unvaccinated patients with Alpha variant compared to vaccinated patients with Delta variant and in left upper lobe volume (55.00% versus 78.15% in unvaccinated patients with Alpha variant compared to vaccinated patients with Delta variant).

Considering all groups together to assess statistically significant differences in terms of median value of extracted volumes by automatic tool, a statistically significant difference was observed in the percentage values of the aerated residual lung volume with a *p*-value of 0.03 for the Kruskal Wallis test (see boxplots in Figure 10 and Table 3) due to the highest value of aerated residual volume in vaccinated patients with Delta variant compered to every other group.

No statistically significant difference was observed in the exitus number among groups (p value = 0.95 at Chi Square test).

Good statistically significant correlations among volumes extracted by automatic tool for each lung lobe and overall radiological severity score were obtained (ICC range 0.71–0.86). Boxplots of the extracted volumes with automatic tool with respect to the overall radiological severity score are reported in Figure 11: aerated residual volume, right upper lobe volume, right lower lobe volume, medium lobe volume, left upper lobe volume, left lower lobe volume in percentage values.

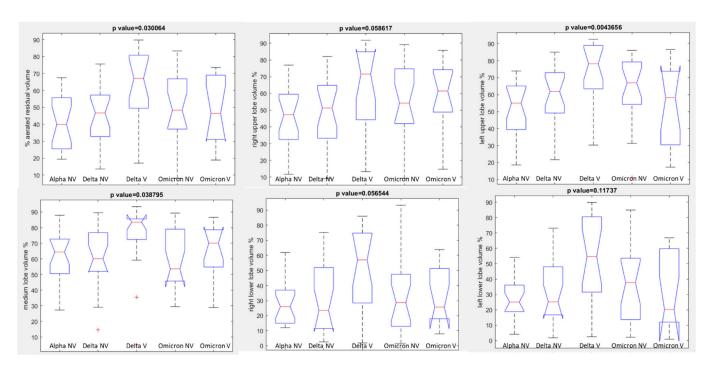


Figure 10. Boxplots of extracted volumes by automatic tool patients affected by Alpha, Delta or Omicron Variant of COVID-19.

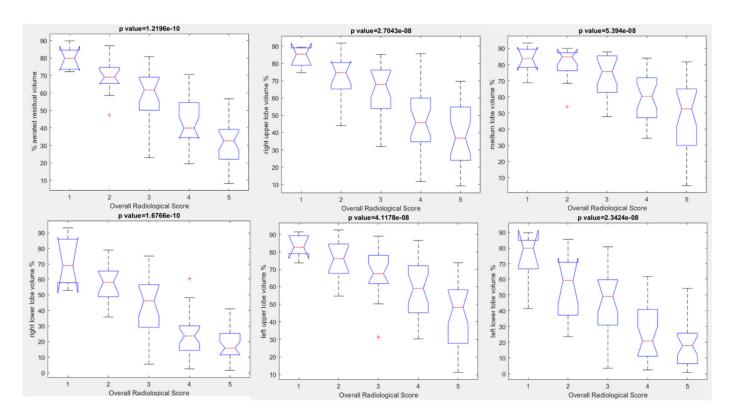


Figure 11. Boxplots of extracted volumes by automatic tool compared to Overall Radiological Severity score.

Table 4 reports the median values of extracted volumes for each patient group (Alpha, Delta and Omicron group) with respect to the overall radiological severity score (from 1 to 5). No statistically significant difference was found in the overall radiological severity

score for each patient group with respect to patients age (p value > 0.05 at Chi square test, Table 5).

Table 4. Median values of extracted volumes by automatic tool for patients affected by Alpha, Delta or Omicron Variant of COVID-19 grouped by overall radiological severity score.

	Overall Radiological SCORE	Aerated Residual Volume %	Right Upper Lobe Volume %	Right Lower Lobe Volume %	Medium Lobe Volume %	Left Upper Lobe Volume %	Left Lower Lobe Volume %
	2	57.50	65.40	48.85	70.50	59.50	26.30
Alpha	3	47.37	72.07	35.27	80.50	66.67	33.03
	4	39.51	40.20	23.73	54.84	49.61	27.15
	5	36.96	39.06	23.71	57.19	44.23	25.16
	1	82.17	86.83	73.63	87.37	84.87	71.67
Delta	2	76.06	74.02	68.02	86.20	83.06	66.08
Dena	3	65.25	68.04	54.91	75.65	72.97	56.29
	4	43.40	47.19	21.51	63.44	61.50	20.47
	5	26.86	29.34	11.54	38.18	45.60	15.18
	1	77.73	80.83	69.03	78.43	81.53	75.93
Omicron	2	67.97	72.62	51.53	78.83	75.50	57.17
	3	47.87	55.15	25.83	65.77	60.47	29.38
	4	46.64	52.84	29.56	57.07	59.79	31.39
	5	34.00	47.12	16.17	51.25	44.08	15.88
p value at Kruskal Wallis test		<0.001	< 0.001	< 0.001	<0.001	< 0.001	<0.001

Table 5. Overall Radiological Severity Score correlated with patients' age for each group.

	Overall Radiological Severity Score	Alpha Variant n = 20	Delta Variant n = 38	Omicronvariant n = 33	p Value at Chi Square Test	
≤65 years	<u><5</u>	0	2	3		
>65 years		0	1	0	0.55	
Total		0	3	3		
≤65 years	<u> </u>	1	2	5		
>65 years		1	3	1	0.32	
Total		2	5	6		
≤65 years	11–15	0	7	4		
>65 years		3	4	2	0.11	
Total		3	11	6		
≤65 years	16–20	4	3	6		
>65 years		4	8	1	0.06	
Total		8	11	7		
≤65 years	21–25	3	3	8		
>65 years	21-23	4	5	3	0.25	
Total		7	8	11		

GGO was the main sign of COVID-19 lesions on CT images. CT showed multiple irregular areas of GGOs in 87 of the 91 (95.6%) patients. Consolidations were found in 70/91 (76.9%) patients and crazy paving sign in 78/91 (86.6%) patients. No statistically significant differences were observed in CT findings (GGO, consolidation or crazy paving sign) among each patient group (p value > 0.05 at Chi square test, Table 1).

4. Discussion

The debate on the efficacy of the vaccine remains, unfortunately, still open, despite the clear evidence of a reduction in the number of patients admitted to ICU [93,94]. A retrospective analysis [94], based from 465 U.S. health care facilities, showed that severe COVID-19 outcomes (i.e., respiratory failure, ICU admission, or death) were rare among adults aged \geq 18 years after primary vaccination. In addition, this study showed that risk for severe COVID-19 outcome after primary vaccination was higher among persons aged \geq 65 years with immunosuppression, diabetes, and chronic kidney, cardiac, pulmonary, neurologic, and liver disease [94]. However, these data were obtained among persons who acquired COVID-19 after primary vaccination during periods of pre-Delta and Delta variant predominance, so that these results should not be applicable to the risk from Omicron variant or future variants [94]. In our study we showed that in critically ill patient no difference was observed in terms of severity of disease due to pulmonary parenchymal involvement, between unvaccinated and vaccinated patients with Omicron variant, between vaccinated or unvaccinated patients with Delta and vaccinated or unvaccinated patients Omicron variant, between unvaccinated patients with the Alpha variant versus vaccinated or unvaccinated patients with the Omicron variant, or between unvaccinated patients with the Alpha variant versus unvaccinated patients with the Delta variant. Instead statistically significant differences were observed between unvaccinated patients affected by Alpha variant and vaccinated patients affected by Delta variant, and between unvaccinated patients with Delta variant versus vaccinated patients with Delta variant. The highest differences were observed between unvaccinated patients with Alpha variant compared to vaccinated patients with Delta variant.

According to our results, no statistically significant difference was observed in the exitus number among groups. This result could be explained by the fact that the patients in the study were all admitted to ICU and for this reason in a serious condition regardless of vaccination. In addition, these results allow us to analyze several issues. Firstly, in critically ill patients the vaccine role is still controversial, and it could be explained by considering the evolution of the disease itself, where pulmonary impairment is also linked to a probable activation of the immune system [95]. Strong evidence indicates that critical illness caused by COVID-19 is qualitatively different from mild or moderate disease, even among hospitalized patients. Although most patients show mild clinical symptoms, about 20% of patients rapidly progress to severe illness characterized by atypical interstitial bilateral pneumonia, acute respiratory distress syndrome and multiorgan dysfunction. Almost 10% of these critically ill patients subsequently die. Insights into the pathogenic mechanisms underlying SARS-CoV-2 infection and COVID-19 progression are emerging and highlight the critical role of the immunological hyper-response in disease exacerbation [96–98].

Secondly, we found no difference between all groups considering pulmonary parenchymal involvement, except in the delta patient group. These data could be explained considering that the prevalence of the delta variant infection, in Italy, corresponds to the period in which the vaccination campaign was more intense, therefore without a decline in vaccine-related immunity, as suggested by emerging evidence [99,100]. A large observational study conducted using nationwide mass vaccination data in Israel showed that a third dose of the BNT162b2 mRNA COVID-19 vaccine is effective in preventing severe COVID-19-related outcomes. Compared with two doses of the vaccine administered at least 5 months before, adding a third dose was estimated to be 93% effective in preventing COVID-19-related admission to hospital, 92% in preventing severe disease, and 81% in preventing COVID-19-related death, as of 7 or more days after the third dose [101]. In our

study group only a few patients had a booster dose; this data could explain to us why not only patients at risk, but also young people in apparent good health were hospitalized in intensive care, and why there were no statistically significant differences between the various risk factors in our sample.

Last but not least, is the tOmicron variant question. Our data do not allow us to establish whether the severity of the disease is linked to a decline in vaccine-related immunity or to the ineffectiveness of the vaccine against the omicron variant. At the present, there are four types of vaccines, i.e., virus vaccines, viral-vector vaccines, DNA/RNA vaccines, and protein-based vaccines [102]. Essentially, the current COVID-19 vaccines in use mainly target the S protein [103]. The 32 amino acid changes, including three small deletions and one small insertion in the spike protein, suggest that these mutations may dramatically enhance the Omicron variant's ability to evade current vaccines [104–106]. Although data has suggested the potential benefit of booster mRNA vaccines for protection against Omicron [107], further studies on a larger sample are necessary.

Our quantitative analysis was obtained by Philips IntelliSpace Portal clinical application CT COPD software, designed to quantify pulmonary emphysema in patients with chronic obstructive pulmonary disease. The tool provides segmentation of the lungs and of the airway tree. Moreover, the tool helps visualize and quantify the destructive process of diffuse lung disease (e.g., emphysema), providing a guided workflow for airway analysis, reviewing and measuring airway lumen, and assessing trapped ait. Compared to others tools, this allows assessment of consolidation. In fact, during our evaluation we used also two others tools, Thoracic VCAR Software (GE Healthcare, Chicago, IL, USA) and a pneumonia module of ANKE ASG-340 CT workstation (HTS Med & Anke, Naples, Italy). However, these tools were unable to identify consolidation in all patients and, to avoid excluding patients, we reported the results obtained with a single tool.

The present study has limitations, first of all the assessed sample size. However, we selected critically ill patients in intensive care who had a CT study for an evaluation of the objective "gravity" of the disease. The possibility of objectively grading the disease made the data robustly comparable, eliminating the variability associated with qualitative assessment [108–120]. Secondly is the small number of patients who had taken a booster dose, which did not allow us to assess whether the additional dose could be protective or not. Third is the selection of the control group, linked to the need to have performed a CT study, which could be responsible for bias in the results. However, we have already explained how an objective quantification of disease severity was considered crucial. Finally, since we did not know the date of the last vaccine dose for all patients, it was not possible to evaluate the severity based on the time of immunity status.

5. Conclusions

The debate on the efficacy of the vaccine remains still open, despite the clear evidence of a reduction in the number of patients admitted to Intensive Care Unit. In our study we showed that in critically ill patients no difference was observed in terms of severity of disease or exitus between unvaccinated and vaccinated patients. The only statistically significant differences were observed, with regard to the severity of COVID-19 pulmonary parenchymal involvement, between unvaccinated patients affected by Alpha variant and vaccinated patients affected by Delta variant, and between unvaccinated patients with Delta variant versus vaccinated patients with Delta variant.

Author Contributions: Each author has participated sufficiently to take public responsibility for the content of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsin-ki and approved by the Institutional Ethics Committee of IRCCS L. Spallanzani. Data acquisition and analysis were performed in compliance with protocols approved by the Ethical

Committee of the National Institute for Infectious Diseases IRCCS Lazzaro Spallanzani, Rome, Italy (ethical approval number 164, 26 June 2020).

Informed Consent Statement: The Local Ethi-cal Committee board renounced patient informed consent, considering the ongoing epidemic emergency.

Data Availability Statement: All data are reported in the manuscript.

Acknowledgments: The authors are grateful to Alessandra Trocino, librarian at the National Cancer Institute of Naples, Italy. Moreover, the authors are grateful to Grazia Della Valle for collaboration.

Conflicts of Interest: The authors have no conflict of interest to be disclosed.

References

- 1. Available online: https://covid19.who.int (accessed on 21 May 2022).
- 2. Stramare, R.; Carretta, G.; Capizzi, A.; Boemo, D.G.; Contessa, C.; Motta, R.; De Conti, G.; Causin, F.; Giraudo, C.; Donato, D. Radiological management of COVID-19: Structure your diagnostic path to guarantee a safe path. *Radiol. Med.* **2020**, 125, 691–694. [CrossRef] [PubMed]
- 3. Granata, V.; Fusco, R.; Izzo, F.; Setola, S.V.; Coppola, M.; Grassi, R.; Reginelli, A.; Cappabianca, S.; Petrillo, A. COVID-19 infection in cancer patients: The management in a diagnostic unit. *Radiol. Oncol.* **2021**, *55*, 121–129. [CrossRef] [PubMed]
- 4. Ashtari, S.; Vahedian-Azimi, A.; Shojaee, S.; Pourhoseingholi, M.A.; Jafari, R.; Bashar, F.R.; Zali, M.R. Características en tomografía computarizada de la neumonía por coronavirus-2019 (COVID-19) en tres grupos de pacientes iraníes: Estudio de un solo centro [Computed tomographic features of coronavirus disease-2019 (COVID-19) pneumonia in three groups of Iranian patients: A single center study]. Radiologia 2021, 63, 314–323. (In Spanish) [CrossRef] [PubMed]
- 5. Gabelloni, M.; Faggioni, L.; Cioni, D.; Mendola, V.; Falaschi, Z.; Coppola, S.; Corradi, F.; Isirdi, A.; Brandi, N.; Coppola, F.; et al. Extracorporeal membrane oxygenation (ECMO) in COVID-19 patients: A pocket guide for radiologists. *Radiol. Med.* **2022**, 13, 369–382. [CrossRef] [PubMed]
- 6. Giovagnoni, A. Facing the COVID-19 emergency: We can, and we do. Radiol. Med. 2020, 125, 337–338. [CrossRef] [PubMed]
- 7. Montesi, G.; Di Biase, S.; Chierchini, S.; Pavanato, G.; Virdis, G.E.; Contato, E.; Mandoliti, G. Radiotherapy during COVID-19 pandemic. How to create a No fly zone: A Northern Italy experience. *Radiol. Med.* **2020**, *125*, 600–603. [CrossRef]
- 8. Ierardi, A.M.; Wood, B.J.; Arrichiello, A.; Bottino, N.; Bracchi, L.; Forzenigo, L.; Andrisani, M.C.; Vespro, V.; Bonelli, C.; Amalou, A.; et al. Preparation of a radiology department in an Italian hospital dedicated to COVID-19 patients. *Radiol. Med.* **2020**, 125, 894–901. [CrossRef]
- 9. Grassi, R.; Cappabianca, S.; Urraro, F.; Feragalli, B.; Montanelli, A.; Patelli, G.; Granata, V.; Giacobbe, G.; Russo, G.M.; Grillo, A.; et al. Chest CT Computerized Aided Quantification of PNEUMONIA Lesions in COVID-19 Infection: A Comparison among Three Commercial Software. *Int. J. Environ. Res. Public Health* **2020**, *17*, 6914. [CrossRef]
- 10. Pediconi, F.; Galati, F.; Bernardi, D.; Belli, P.; Brancato, B.; Calabrese, M.; Camera, L.; Carbonaro, L.A.; Caumo, F.; Clauser, P.; et al. Breast imaging and cancer diagnosis during the COVID-19 pandemic: Recommendations from the Italian College of Breast Radiologists by SIRM. *Radiol. Med.* **2020**, 125, 926–930. [CrossRef]
- 11. Koç, A.; Sezgin, S.; Kayıpmaz, S. Comparing different planimetric methods on volumetric estimations by using cone beam computed tomography. *Radiol. Med.* **2020**, 125, 398–405. [CrossRef]
- 12. Fusco, R.; Simonetti, I.; Ianniello, S.; Villanacci, A.; Grassi, F.; Dell'Aversana, F.; Grassi, R.; Cozzi, D.; Bicci, E.; Palumbo, P.; et al. Pulmonary Lymphangitis Poses a Major Challenge for Radiologists in an Oncological Setting during the COVID-19 Pandemic. *J. Pers. Med.* 2022, 12, 624. [CrossRef] [PubMed]
- 13. de Souza, A.S.; de Freitas Amorim, V.M.; Guardia, G.D.A.; Dos Santos, F.F.; Ulrich, H.; Galante, P.A.F.; de Souza, R.F.; Guzzo, C.R. Severe Acute Respiratory Syndrome Coronavirus 2 Variants of Concern: A Perspective for Emerging More Transmissible and Vaccine-Resistant Strains. *Viruses* 2022, 14, 827. [CrossRef] [PubMed]
- 14. Agostini, A.; Floridi, C.; Borgheresi, A.; Badaloni, M.; Pirani, P.E.; Terilli, F.; Ottaviani, L.; Giovagnoni, A. Proposal of a low-dose, long-pitch, dual-source chest CT protocol on third-generation dual-source CT using a tin filter for spectral shaping at 100 kVp for CoronaVirus Disease 2019 (COVID-19) patients: A feasibility study. *Radiol. Med.* 2020, 125, 365–373. [CrossRef] [PubMed]
- 15. Borghesi, A.; Maroldi, R. COVID-19 outbreak in Italy: Experimental chest X-ray scoring system for quantifying and monitoring disease progression. *Radiol. Med.* **2020**, *125*, 509–513. [CrossRef] [PubMed]
- 16. Neri, E.; Miele, V.; Coppola, F.; Grassi, R. Use of CT and artificial intelligence in suspected or COVID-19 positive patients: Statement of the Italian Society of Medical and Interventional Radiology. *Radiol. Med.* **2020**, *125*, 505–508. [CrossRef]
- 17. Carotti, M.; Salaffi, F.; Sarzi-Puttini, P.; Agostini, A.; Borgheresi, A.; Minorati, D.; Galli, M.; Marotto, D.; Giovagnoni, A. Chest CT features of coronavirus disease 2019 (COVID-19) pneumonia: Key points for radiologists. *Radiol. Med.* **2020**, 125, 636–646. [CrossRef]
- 18. Alyasseri, Z.A.A.; Al-Betar, M.A.; Doush, I.A.; Awadallah, M.A.; Abasi, A.K.; Makhadmeh, S.N.; Alomari, O.A.; Abdulkareem, K.H.; Adam, A.; Damasevicius, R.; et al. Review on COVID-19 diagnosis models based on machine learning and deep learning approaches. *Expert Syst.* **2021**, *28*, e12759. [CrossRef]

19. Borghesi, A.; Zigliani, A.; Masciullo, R.; Golemi, S.; Maculotti, P.; Farina, D.; Maroldi, R. Radiographic severity index in COVID-19 pneumonia: Relationship to age and sex in 783 Italian patients. *Radiol. Med.* **2020**, 125, 461–464. [CrossRef]

- 20. Cozzi, D.; Albanesi, M.; Cavigli, E.; Moroni, C.; Bindi, A.; Luvarà, S.; Lucarini, S.; Busoni, S.; Mazzoni, L.N.; Miele, V. Chest X-ray in new Coronavirus Disease 2019 (COVID-19) infection: Findings and correlation with clinical outcome. *Radiol. Med.* 2020, 125, 730–737. [CrossRef]
- 21. Gatti, M.; Calandri, M.; Barba, M.; Biondo, A.; Geninatti, C.; Gentile, S.; Greco, M.; Morrone, V.; Piatti, C.; Santonocito, A.; et al. Baseline chest X-ray in coronavirus disease 19 (COVID-19) patients: Association with clinical and laboratory data. *Radiol. Med.* 2020, 125, 1271–1279. [CrossRef]
- 22. Granata, V.; Fusco, R.; Vallone, P.; Setola, S.V.; Picone, C.; Grassi, F.; Patrone, R.; Belli, A.; Izzo, F.; Petrillo, A. Not only lymphadenopathy: Case of chest lymphangitis assessed with MRI after COVID 19 vaccine. *Infect. Agent Cancer* 2022, 17, 8. [CrossRef] [PubMed]
- 23. D'Agostino, V.; Caranci, F.; Negro, A.; Piscitelli, V.; Tuccillo, B.; Fasano, F.; Sirabella, G.; Marano, I.; Granata, V.; Grassi, R.; et al. A Rare Case of Cerebral Venous Thrombosis and Disseminated Intravascular Coagulation Temporally Associated to the COVID-19 Vaccine Administration. *J. Pers. Med.* 2021, 11, 285. [CrossRef] [PubMed]
- 24. Granata, V.; Fusco, R.; Setola, S.V.; Galdiero, R.; Picone, C.; Izzo, F.; D'Aniello, R.; Miele, V.; Grassi, R.; Grassi, R.; et al. Lymphadenopathy after BNT162b2 COVID-19 Vaccine: Preliminary Ultrasound Findings. *Biology* **2021**, *10*, 214. [CrossRef] [PubMed]
- 25. Volz, E.; Hill, V.; McCrone, J.T.; Price, A.; Jorgensen, D.; O'Toole, Á.; Southgate, J.; Johnson, R.; Jackson, B.; Nascimento, F.F.; et al. Evaluating the Effects of SARS-CoV-2 Spike Mutation D614G on Transmissibility and Pathogenicity. *Cell* **2021**, 184, 64–75. [CrossRef]
- 26. Zhou, W.; Wang, W. Fast-Spreading SARS-CoV-2 Variants: Challenges to and New Design Strategies of COVID-19 Vaccines. Signal Transduct. Target. Ther. 2021, 6, 226. [CrossRef]
- 27. Zhang, J.; Xiao, T.; Cai, Y.; Lavine, C.L.; Peng, H.; Zhu, H.; Anand, K.; Tong, P.; Gautam, A.; Mayer, M.L.; et al. Membrane Fusion and Immune Evasion by the Spike Protein of SARS-CoV-2 Delta Variant. *Science* **2021**, *374*, 1353–1360. [CrossRef]
- 28. Classification of Omicron (B.1.1.529): SARS-CoV-2 Variant of Concern. Available online: https://www.who.int/news/item/26-1 1-2021-classification-of-omicron-(b.1.1.529)-sars-cov-2-variant-of-concern (accessed on 17 December 2021).
- 29. Ierardi, A.M.; Gaibazzi, N.; Tuttolomondo, D.; Fusco, S.; La Mura, V.; Peyvandi, F.; Aliberti, S.; Blasi, F.; Cozzi, D.; Carrafiello, G.; et al. Deep vein thrombosis in COVID-19 patients in general wards: Prevalence and association with clinical and laboratory variables. *Radiol. Med.* **2021**, *126*, 722–728. [CrossRef]
- 30. Turkahia, Y.; Thornlow, B.; Hinrichs, A.; McBroome, J.; Ayala, N.; Ye, C.; De Maio, N.; Haussler, D.; Lanfear, R.; Corbett-Detig, R. Pandemic-Scale Phylogenomics Reveals Elevated Recombination Rates in the SARS-CoV-2 Spike Region. *bioRxiv* 2021. [CrossRef]
- 31. Covin, S.; Rutherford, G.W. Coinfection, Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2), and Influenza: An Evolving Puzzle. *Clin. Infect. Dis.* **2021**, 72, e993–e994. [CrossRef]
- 32. Giannitto, C.; Sposta, F.M.; Repici, A.; Vatteroni, G.; Casiraghi, E.; Casari, E.; Ferraroli, G.M.; Fugazza, A.; Sandri, M.T.; Chiti, A.; et al. Chest CT in patients with a moderate or high pretest probability of COVID-19 and negative swab. *Radiol. Med.* 2020, 125, 1260–1270. [CrossRef]
- 33. Cuadrado-Payán, E.; Montagud-Marrahi, E.; Torres-Elorza, M.; Bodro, M.; Blasco, M.; Poch, E.; Soriano, A.; Piñeiro, G.J. SARS-CoV-2 and Influenza Virus Co-Infection. *Lancet* **2020**, *395*, e84. [CrossRef]
- 34. Moroni, C.; Cozzi, D.; Albanesi, M.; Cavigli, E.; Bindi, A.; Luvarà, S.; Busoni, S.; Mazzoni, L.N.; Grifoni, S.; Nazerian, P.; et al. Chest X-ray in the emergency department during COVID-19 pandemic descending phase in Italy: Correlation with patients' outcome. *Radiol. Med.* 2021, 126, 661–668. [CrossRef] [PubMed]
- 35. Di Serafino, M.; Notaro, M.; Rea, G.; Iacobellis, F.; Paoli, V.D.; Acampora, C.; Ianniello, S.; Brunese, L.; Romano, L.; Vallone, G. The lung ultrasound: Facts or artifacts? In the era of COVID-19 outbreak. *Radiol. Med.* **2020**, *125*, 738–753. [CrossRef]
- 36. Ravikanth, R. Diagnostic accuracy and false-positive rate of chest CT as compared to RT-PCR in coronavirus disease 2019 (COVID-19) pneumonia: A prospective cohort of 612 cases from India and review of literature. *Indian J. Radiol. Imaging* 2021, 31 (Suppl. 1), S161–S169. [CrossRef] [PubMed]
- 37. Grassi, R.; Belfiore, M.P.; Montanelli, A.; Patelli, G.; Urraro, F.; Giacobbe, G.; Fusco, R.; Granata, V.; Petrillo, A.; Sacco, P.; et al. COVID-19 pneumonia: Computer-aided quantification of healthy lung parenchyma, emphysema, ground glass and consolidation on chest computed tomography (CT). *Radiol. Med.* **2020**, 126, 553–560. [CrossRef] [PubMed]
- 38. Ippolito, D.; Giandola, T.; Maino, C.; Pecorelli, A.; Capodaglio, C.; Ragusi, M.; Porta, M.; Gandola, D.; Masetto, A.; Drago, S.; et al. Acute pulmonary embolism in hospitalized patients with SARS-CoV-2-related pneumonia: Multicentric experience from Italian endemic area. *Radiol. Med.* **2021**, 126, 669–678. [CrossRef]
- 39. Mansbach, R.A.; Chakraborty, S.; Nguyen, K.; Montefiori, D.C.; Korber, B.; Gnanakaran, S. The SARS-CoV-2 Spike Variant D614G Favors an Open Conformational State. *Sci. Adv.* **2021**, *7*, eabf3671. [CrossRef]
- 40. Shaw, B.; Daskareh, M.; Gholamrezanezhad, A. The lingering manifestations of COVID-19 during and after convalescence: Update on long-term pulmonary consequences of coronavirus disease 2019 (COVID-19). *Radiol. Med.* **2021**, 126, 40–46. [CrossRef]
- 41. Rawashdeh, M.A.; Saade, C. Radiation dose reduction considerations and imaging patterns of ground glass opacities in coronavirus: Risk of over exposure in computed tomography. *Radiol. Med.* **2021**, *126*, 380–387. [CrossRef]

42. Teruel, N.; Mailhot, O.; Najmanovich, R.J. Modelling Conformational State Dynamics and Its Role on Infection for SARS-CoV-2 Spike Protein Variants. *PLoS Comput. Biol.* **2021**, 17, e1009286. [CrossRef]

- 43. Benton, D.J.; Wrobel, A.G.; Roustan, C.; Borg, A.; Xu, P.; Martin, S.R.; Rosenthal, P.B.; Skehel, J.J.; Gamblin, S.J. The Effect of the D614G Substitution on the Structure of the Spike Glycoprotein of SARS-CoV-2. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2022586118. [CrossRef] [PubMed]
- 44. Chemaitelly, H.; Tang, P.; Hasan, M.R.; AlMukdad, S.; Yassine, H.M.; Benslimane, F.M.; Al Khatib, H.A.; Coyle, P.; Ayoub, H.H.; Al Kanaani, Z.; et al. Waning of BNT162b2 vaccine protection against SARS-CoV-2 infection in Qatar. *N. Engl. J. Med.* **2021**, 385, e83. [CrossRef] [PubMed]
- 45. Levin, E.G.; Lustig, Y.; Cohen, C.; Fluss, R.; Indenbaum, V.; Amit, S.; Doolman, R.; Asraf, K.; Mendelson, E.; Ziv, A.; et al. Waning immune humoral response to BNT162b2 covid-19 vaccine over 6 months. *N. Engl. J. Med.* **2021**, *385*, e84. [CrossRef] [PubMed]
- 46. Andrews, N.; Tessier, E.; Stowe, J.; Gower, C.; Kirsebom, F.; Simmons, R.; Gallagher, E.; Chand, M.; Brown, K.; Ladhani, S.; et al. Vaccine effectiveness and duration of protection of Comirnaty, Vaxzevria and Spikevax against mild and severe COVID-19 in the UK. *medRxiv* 2021. Available online: https://www.medrxiv.org/content/10.1101/2021.09.15.21263583v2 (accessed on 21 May 2022).
- 47. Goldberg, Y.; Mandel, M.; Bar-On, Y.M.; Bodenheimer, O.; Freedman, L.S.; Haas, E.; Milo, R.; Alroy-Preis, S.; Ash, N.; Huppert, A. Waning immunity of the BNT162b2 vaccine: A nationwide study from Israel. *medRxiv* **2021**. Available online: https://www.medrxiv.org/content/10.1101/2021.08.24.21262423v1 (accessed on 21 May 2022).
- 48. Thomas, S.J.; Moreira, E.D.; Kitchin, N.; Absalon, J.; Gurtman, A.; Lockhart, S.; Perez, J.L.; Marc, G.P.; Polack, F.P.; Zerbini, C.; et al. Six month safety and efficacy of the BNT162b2 mRNA COVID-19 vaccine. *medRxiv* **2021**. Available online: https://www.medrxiv.org/content/10.1101/2021.07.28.21261159v1 (accessed on 21 May 2022).
- 49. COVID-19 Vaccine Booster Doses Administered per 100 People—Our World in Data [Internet]. [Cited 2021 Nov.]. Available online: https://ourworldindata.org/grapher/covid-vaccine-booster-doses-per-capita?country=BRA~{}CHL~{}ISR~{}RUS~{}USA~{}URY~{}OWID_WRL (accessed on 21 May 2022).
- 50. Lombardi, A.F.; Afsahi, A.M.; Gupta, A.; Gholamrezanezhad, A. Severe acute respiratory syndrome (SARS), Middle East respiratory syndrome (MERS), influenza, and COVID-19, beyond the lungs: A review article. *Radiol. Med.* **2021**, *126*, 561–569. [CrossRef]
- 51. Alballa, N.; Al-Turaiki, I. Machine learning approaches in COVID-19 diagnosis, mortality, and severity risk prediction: A review. *Inform. Med. Unlocked* **2021**, 24, 100564. [CrossRef]
- 52. Cozzi, D.; Bindi, A.; Cavigli, E.; Grosso, A.M.; Luvarà, S.; Morelli, N.; Moroni, C.; Piperio, R.; Miele, V.; Bartolucci, M. Exogenous lipoid pneumonia: When radiologist makes the difference. *Radiol. Med.* **2021**, *126*, 22–28. [CrossRef]
- 53. Palmisano, A.; Scotti, G.M.; Ippolito, D.; Morelli, M.J.; Vignale, D.; Gandola, D.; Sironi, S.; De Cobelli, F.; Ferrante, L.; Spessot, M.; et al. Chest CT in the emergency department for suspected COVID-19 pneumonia. *Radiol. Med.* **2021**, *126*, 498–502. [CrossRef]
- 54. Bar-On, Y.M.; Goldberg, Y.; Mandel, M.; Bodenheimer, O.; Freedman, L.; Kalkstein, N.; Mizrahi, B.; Alroy-Preis, S.; Ash, N.; Milo, R.; et al. Protection of BNT162b2 Vaccine Booster against COVID-19 in Israel. *N. Engl. J. Med.* **2021**, *385*, 1393–1400. [CrossRef]
- 55. Waxman, J.G.; Makov-Assif, M.; Reis, B.Y.; Netzer, D.; Balicer, R.D.; Dagan, N.; Barda, N. Comparing COVID-19-related hospitalization rates among individuals with infection-induced and vaccine-induced immunity in Israel. *Nat. Commun.* 2022, 13, 2202. [CrossRef] [PubMed]
- 56. Caruso, D.; Polici, M.; Zerunian, M.; Pucciarelli, F.; Polidori, T.; Guido, G.; Rucci, C.; Bracci, B.; Muscogiuri, E.; De Dominicis, C.; et al. Quantitative Chest CT analysis in discriminating COVID-19 from non-COVID-19 patients. *Radiol. Med.* **2020**, *126*, 243–249. [CrossRef] [PubMed]
- 57. Cardobi, N.; Benetti, G.; Cardano, G.; Arena, C.; Micheletto, C.; Cavedon, C.; Montemezzi, S. CT radiomic models to distinguish COVID-19 pneumonia from other interstitial pneumonias. *Radiol. Med.* **2021**, *126*, 1037–1043. [CrossRef] [PubMed]
- 58. Caruso, D.; Pucciarelli, F.; Zerunian, M.; Ganeshan, B.; De Santis, D.; Polici, M.; Rucci, C.; Polidori, T.; Guido, G.; Bracci, B.; et al. Chest CT texture-based radiomics analysis in differentiating COVID-19 from other interstitial pneumonia. *Radiol. Med.* **2021**, 126, 1415–1424. [CrossRef] [PubMed]
- 59. Masselli, G.; Almberger, M.; Tortora, A.; Capoccia, L.; Dolciami, M.; D'Aprile, M.R.; Valentini, C.; Avventurieri, G.; Bracci, S.; Ricci, P. Role of CT angiography in detecting acute pulmonary embolism associated with COVID-19 pneumonia. *Radiol. Med.* **2021**, 126, 1553–1560. [CrossRef] [PubMed]
- 60. Pecoraro, M.; Cipollari, S.; Marchitelli, L.; Messina, E.; Del Monte, M.; Galea, N.; Ciardi, M.R.; Francone, M.; Catalano, C.; Panebianco, V. Cross-sectional analysis of follow-up chest MRI and chest CT scans in patients previously affected by COVID-19. *Radiol. Med.* **2021**, *126*, 1273–1281. [CrossRef]
- 61. Granata, V.; Ianniello, S.; Fusco, R.; Urraro, F.; Pupo, D.; Magliocchetti, S.; Albarello, F.; Campioni, P.; Cristofaro, M.; Di Stefano, F.; et al. Quantitative Analysis of Residual COVID-19 Lung CT Features: Consistency among Two Commercial Software. *J. Pers. Med.* 2021, 11, 1103. [CrossRef]
- 62. Spinato, G.; Fabbris, C.; Conte, F.; Menegaldo, A.; Franz, L.; Gaudioso, P.; Cinetto, F.; Agostini, C.; Costantini, G.; Boscolo-Rizzo, P. COVID-Q: Validation of the first COVID-19 questionnaire based on patient-rated symptom gravity. *Int. J. Clin. Pract.* **2021**, 75, e14829. [CrossRef]
- 63. Bertolini, M.; Brambilla, A.; Dallasta, S.; Colombo, G. High-quality chest CT segmentation to assess the impact of COVID-19 disease. *Int. J. Comput. Assist. Radiol. Surg.* **2021**, *16*, 1737–1747. [CrossRef]

64. Zieda, A.; Sbardella, S.; Patel, M.; Smith, R. Diagnostic Bias in the COVID-19 Pandemic: A Series of Short Cases. *Eur. J. Case Rep. Intern. Med.* **2021**, *8*, 002575. [CrossRef]

- 65. Giannakis, A.; Móré, D.; Erdmann, S.; Kintzelé, L.; Fischer, R.M.; Vogel, M.N.; Mangold, D.L.; von Stackelberg, O.; Schnitzler, P.; Zimmermann, S.; et al. COVID-19 pneumonia and its lookalikes: How radiologists perform in differentiating atypical pneumonias. *Eur. J. Radiol.* 2021, 144, 110002. [CrossRef] [PubMed]
- 66. Borghesi, A.; Sverzellati, N.; Polverosi, R.; Balbi, M.; Baratella, E.; Busso, M.; Calandriello, L.; Cortese, G.; Farchione, A.; Iezzi, R.; et al. Impact of the COVID-19 pandemic on the selection of chest imaging modalities and reporting systems: A survey of Italian radiologists. *Radiol. Med.* **2021**, *126*, 1258–1272. [CrossRef] [PubMed]
- 67. Cozzi, D.; Bicci, E.; Bindi, A.; Cavigli, E.; Danti, G.; Galluzzo, M.; Granata, V.; Pradella, S.; Trinci, M.; Miele, V. Role of Chest Imaging in Viral Lung Diseases. *Int. J. Environ. Res. Public Health* **2021**, *18*, 6434. [CrossRef] [PubMed]
- 68. Fusco, R.; Grassi, R.; Granata, V.; Setola, S.V.; Grassi, F.; Cozzi, D.; Pecori, B.; Izzo, F.; Petrillo, A. Artificial Intelligence and COVID-19 Using Chest CT Scan and Chest X-ray Images: Machine Learning and Deep Learning Approaches for Diagnosis and Treatment. *J. Pers. Med.* 2021, 11, 993. [CrossRef]
- 69. Reginelli, A.; Grassi, R.; Feragalli, B.; Belfiore, M.P.; Montanelli, A.; Patelli, G.; La Porta, M.; Urraro, F.; Fusco, R.; Granata, V.; et al. Coronavirus Disease 2019 (COVID-19) in Italy: Double Reading of Chest CT Examination. *Biology* **2021**, *10*, 89. [CrossRef]
- 70. Grassi, R.; Fusco, R.; Belfiore, M.P.; Montanelli, A.; Patelli, G.; Urraro, F.; Petrillo, A.; Granata, V.; Sacco, P.; Mazzei, M.A.; et al. Coronavirus disease 2019 (COVID-19) in Italy: Features on chest computed tomography using a structured report system. *Sci. Rep.* 2020, *10*, 17236, Erratum in *Sci. Rep.* 2021, *11*, 4231. [CrossRef]
- 71. Masci, G.M.; Iafrate, F.; Ciccarelli, F.; Pambianchi, G.; Panebianco, V.; Pasculli, P.; Ciardi, M.R.; Mastroianni, C.M.; Ricci, P.; Catalano, C.; et al. Tocilizumab effects in COVID-19 pneumonia: Role of CT texture analysis in quantitative assessment of response to therapy. *Radiol. Med.* **2021**, *126*, 1170–1180. [CrossRef]
- 72. Francolini, G.; Desideri, I.; Stocchi, G.; Ciccone, L.P.; Salvestrini, V.; Garlatti, P.; Aquilano, M.; Greto, D.; Bonomo, P.; Meattini, I.; et al. Impact of COVID-19 on workload burden of a complex radiotherapy facility. *Radiol. Med.* **2021**, *126*, 717–721. [CrossRef]
- 73. Cellini, F.; Di Franco, R.; Manfrida, S.; Borzillo, V.; Maranzano, E.; Pergolizzi, S.; Morganti, A.G.; Fusco, V.; Deodato, F.; Santarelli, M.; et al. Palliative radiotherapy indications during the COVID-19 pandemic and in future complex logistic settings: The NORMALITY model. *Radiol. Med.* **2021**, 126, 1619–1656. [CrossRef]
- 74. De Felice, F.; D'Angelo, E.; Ingargiola, R.; Iacovelli, N.A.; Alterio, D.; Franco, P.; Bonomo, P.; Merlotti, A.; Bacigalupo, A.; Maddalo, M.; et al. A snapshot on radiotherapy for head and neck cancer patients during the COVID-19 pandemic: A survey of the Italian Association of Radiotherapy and Clinical Oncology (AIRO) head and neck working group. *Radiol. Med.* **2020**, 126, 343–347. [CrossRef]
- 75. Ortiz, S.; Rojas, F.; Valenzuela, O.; Herrera, L.J.; Rojas, I. Determination of the Severity and Percentage of COVID-19 Infection through a Hierarchical Deep Learning System. *J. Pers. Med.* **2022**, *12*, 535. [CrossRef] [PubMed]
- Mohiuddin Chowdhury, A.T.M.; Kamal, A.; Abbas, K.U.; Talukder, S.; Karim, M.R.; Ali, M.A.; Nuruzzaman, M.; Li, Y.; He, S. Efficacy and Outcome of Remdesivir and Tocilizumab Combination Against Dexamethasone for the Treatment of Severe COVID-19: A Randomized Controlled Trial. Front. Pharmacol. 2022, 13, 690726. [CrossRef] [PubMed]
- 77. Ibrahim, H.M.; Mohammad, A.A.; Fouda, E.; Abouelfotouh, K.; Habeeb, N.M.; Rezk, A.R.; Magdy, S.; Allam, A.M.; Mahmoud, S.A. Clinical Characteristics and Pulmonary Computerized Imaging Findings of Critically Ill Egyptian Patients with Multisystem Inflammatory Syndrome in Children. *Glob. Pediatr. Health* 2022, 9, 2333794X221085386. [CrossRef] [PubMed]
- 78. Lorent, N.; Vande Weygaerde, Y.; Claeys, E.; Guler Caamano Fajardo, I.; De Vos, N.; De Wever, W.; Salhi, B.; Gyselinck, I.; Bosteels, C.; Lambrecht, B.N.; et al. Prospective longitudinal evaluation of hospitalised COVID-19 survivors 3 and 12 months after discharge. *ERJ Open Res.* **2022**, *8*, 00004–2022. [CrossRef] [PubMed]
- 79. Ohno, Y.; Aoyagi, K.; Arakita, K.; Doi, Y.; Kondo, M.; Banno, S.; Kasahara, K.; Ogawa, T.; Kato, H.; Hase, R.; et al. Newly developed artificial intelligence algorithm for COVID-19 pneumonia: Utility of quantitative CT texture analysis for prediction of favipiravir treatment effect. *Jpn. J. Radiol.* 2022, *9*, 1–14. [CrossRef] [PubMed]
- 80. Yanamandra, U.; Shobhit, S.; Paul, D.; Aggarwal, B.; Kaur, P.; Duhan, G.; Singh, A.; Srinath, R.; Saxena, P.; Menon, A.S. Relationship of Computed Tomography Severity Score with Patient Characteristics and Survival in Hypoxemic COVID-19 Patients. *Cureus* 2022, 14, e22847. [CrossRef]
- 81. Ghafuri, L.; Hamzehzadeh Alamdari, A.; Roustaei, S.; Golshani Beheshti, A.; Nayerpour, A. Predicting Severity of Novel Coronavirus (COVID-19) Pneumonia Based upon Admission Clinical, Laboratory, and Imaging Findings. *Tanaffos* **2021**, *20*, 232–239.
- 82. Karthik, R.; Menaka, R.; Hariharan, M.; Won, D. CT-based severity assessment for COVID-19 using weakly supervised non-local CNN. *Appl. Soft Comput.* **2022**, *121*, 108765. [CrossRef]
- 83. Vargas Centanaro, G.; Calle Rubio, M.; Álvarez-Sala Walther, J.L.; Martinez-Sagasti, F.; Albuja Hidalgo, A.; Herranz Hernández, R.; Rodríguez Hermosa, J.L. Long-term Outcomes and Recovery of Patients who Survived COVID-19: LUNG INJURY COVID-19 Study. Open Forum Infect. Dis. 2022, 9, ofac098. [CrossRef]
- 84. Küçük, M.; Ergan, B.; Yakar, M.N.; Ergün, B.; Akdoğan, Y.; Cantürk, A.; Gezer, N.S.; Kalkan, F.; Yaka, E.; Cömert, B.; et al. The Predictive Values of Respiratory Rate Oxygenation Index and Chest Computed Tomography Severity Score for High-Flow Nasal Oxygen Failure in Critically Ill Patients with Coronavirus Disease-2019. *Balkan Med. J.* 2022, 39, 140–147. [CrossRef]
- 85. Özel, M.; Aslan, A.; Araç, S. Use of the COVID-19 Reporting and Data System (CO-RADS) classification and chest computed tomography involvement score (CT-IS) in COVID-19 pneumonia. *Radiol. Med.* **2021**, *126*, 679–687. [CrossRef] [PubMed]

86. Cereser, L.; Girometti, R.; Da Re, J.; Marchesini, F.; Como, G.; Zuiani, C. Inter-reader agreement of high-resolution computed tomography findings in patients with COVID-19 pneumonia: A multi-reader study. *Radiol. Med.* **2021**, *126*, 577–584. [CrossRef] [PubMed]

- 87. Cappabianca, S.; Fusco, R.; de Lisio, A.; Paura, C.; Clemente, A.; Gagliardi, G.; Lombardi, G.; Giacobbe, G.; Russo, G.M.; Belfiore, M.P.; et al. Correction to: Clinical and laboratory data, radiological structured report findings and quantitative evaluation of lung involvement on baseline chest CT in COVID-19 patients to predict prognosis. *Radiol. Med.* **2021**, *126*, 643, Erratum in *Radiol. Med.* **2021**, *126*, 29–39. [CrossRef] [PubMed]
- 88. Cartocci, G.; Colaiacomo, M.C.; Lanciotti, S.; Andreoli, C.; De Cicco, M.L.; Brachetti, G.; Pugliese, S.; Capoccia, L.; Tortora, A.; Scala, A.; et al. Correction to: Chest CT for early detection and management of coronavirus disease (COVID-19): A report of 314 patients admitted to Emergency Department with suspected pneumonia. *Radiol. Med.* 2021, 126, 642, Erratum in *Radiol. Med.* 2020, 125, 931–942. [CrossRef] [PubMed]
- 89. Bianchi, A.; Mazzoni, L.N.; Busoni, S.; Pinna, N.; Albanesi, M.; Cavigli, E.; Cozzi, D.; Poggesi, A.; Miele, V.; Fainardi, E.; et al. Assessment of cerebrovascular disease with computed tomography in COVID-19 patients: Correlation of a novel specific visual score with increased mortality risk. *Radiol. Med.* 2021, 126, 570–576. [CrossRef] [PubMed]
- 90. Kovács, A.; Palásti, P.; Veréb, D.; Bozsik, B.; Palkó, A.; Kincses, Z.T. The sensitivity and specificity of chest CT in the diagnosis of COVID-19. *Eur. Radiol.* **2021**, *31*, 2819–2824. [CrossRef] [PubMed]
- 91. Li, K.; Wu, J.; Wu, F.; Guo, D.; Chen, L.; Fang, Z.; Li, C. The clinical and chest CT features associated with severe and critical COVID-19 pneumonia. *Investig. Radiol.* **2020**, *55*, 327–331. [CrossRef]
- 92. Hansell, D.M.; Bankier, A.A.; MacMahon, H.; McLoud, T.C.; Muller, N.L.; Remy, J. Fleischner Society: Glossary of terms for thoracic imaging. *Radiology* **2008**, 246, 697–722. [CrossRef]
- 93. Available online: https://who.maps.arcgis.com/apps/dashboards/ead3c6475654481ca51c248d52ab9c61 (accessed on 21 May 2022).
- 94. Yek, C.; Warner, S.; Wiltz, J.L.; Sun, J.; Adjei, S.; Mancera, A.; Silk, B.J.; Gundlapalli, A.V.; Harris, A.M.; Boehmer, T.K.; et al. Risk Factors for Severe COVID-19 Outcomes among Persons Aged ≥ 18 Years Who Completed a Primary COVID-19 Vaccination Series—465 Health Care Facilities, United States, December 2020–October 2021. MMWR Morb. Mortal. Wkly. Rep. 2022, 71, 19–25. [CrossRef]
- 95. Pairo-Castineira, E.; Clohisey, S.; Klaric, L.; Bretherick, A.D.; Rawlik, K.; Pasko, D.; Walker, S.; Parkinson, N.; Fourman, M.H.; Russell, C.D.; et al. Genetic mechanisms of critical illness in COVID-19. *Nature* **2021**, *591*, 92–98. [CrossRef]
- 96. Perico, L.; Benigni, A.; Casiraghi, F.; Ng, L.F.P.; Renia, L.; Remuzzi, G. Immunity, endothelial injury and complement-induced coagulopathy in COVID-19. *Nat. Rev. Nephrol.* **2021**, 17, 46–64. [CrossRef] [PubMed]
- 97. Gupta, S.; Wang, W.; Hayek, S.S.; Chan, L.; Mathews, K.S.; Melamed, M.L.; Brenner, S.K.; Leonberg-Yoo, A.; Schenck, E.J.; Radbel, J.; et al. Association Between Early Treatment with Tocilizumab and Mortality Among Critically Ill Patients with COVID-19. *JAMA Intern. Med.* 2021, 181, 41–51, Erratum in *JAMA Intern. Med.* 2021, 181, 570. [CrossRef] [PubMed]
- 98. Leentjens, J.; van Haaps, T.F.; Wessels, P.F.; Schutgens, R.E.G.; Middeldorp, S. COVID-19-associated coagulopathy and antithrombotic agents-lessons after 1 year. *Lancet Haematol.* **2021**, *8*, e524–e533. [CrossRef]
- 99. Scobie, H.M.; Johnson, A.G.; Suthar, A.B.; Severson, R.; Alden, N.B.; Balter, S.; Bertolino, D.; Blythe, D.; Brady, S.; Cadwell, B.; et al. Monitoring Incidence of COVID-19 Cases, Hospitalizations, and Deaths, by Vaccination Status—13 U.S. Jurisdictions, April 4–July 17, 2021. MMWR Morb. Mortal. Wkly. Rep. 2021, 70, 1284–1290. [CrossRef] [PubMed]
- 100. Mallapaty, S. China's COVID vaccines have been crucial—Now immunity is waning. *Nature* **2021**, *598*, 398–399. [CrossRef] [PubMed]
- 101. Barda, N.; Dagan, N.; Cohen, C.; Hernán, M.A.; Lipsitch, M.; Kohane, I.S.; Reis, B.Y.; Balicer, R.D. Effectiveness of a third dose of the BNT162b2 mRNA COVID-19 vaccine for preventing severe outcomes in Israel: An observational study. *Lancet* **2021**, 398, 2093–2100. [CrossRef]
- 102. Callaway, E. The race for coronavirus vaccines: A graphical guide. Nature 2020, 580, 576–577. [CrossRef]
- 103. Dai, L.; Gao, G.F. Viral targets for vaccines against COVID-19. Nat. Rev. Immunol. 2021, 21, 73–82. [CrossRef]
- 104. Chen, J.; Wang, R.; Gilby, N.B.; Wei, G.W. Omicron (B.1.1.529): Infectivity, vaccine breakthrough, and antibody resistance. *arXiv* **2021**. Update in *J. Chem. Inf. Model.* **2022**, 62, 412–422.
- 105. Ren, S.Y.; Wang, W.B.; Gao, R.D.; Zhou, A.M. Omicron variant (B.1.1.529) of SARS-CoV-2: Mutation, infectivity, transmission, and vaccine resistance. *World J. Clin. Cases.* **2022**, *10*, 1–11. [CrossRef]
- 106. Araf, Y.; Akter, F.; Tang, Y.D.; Fatemi, R.; Parvez, M.S.A.; Zheng, C.; Hossain, M.G. Omicron variant of SARS-CoV-2: Genomics, transmissibility, and responses to current COVID-19 vaccines. *J. Med. Virol.* **2022**, *94*, 1825–1832. [CrossRef] [PubMed]
- 107. Lusvarghi, S.; Pollett, S.D.; Neerukonda, S.N.; Wang, W.; Wang, R.; Vassell, R.; Epsi, N.J.; Fries, A.C.; Agan, B.K.; Lindholm, D.A.; et al. SARS-CoV-2 Omicron neutralization by therapeutic antibodies, convalescent sera, and post-mRNA vaccine booster. bioRxiv 2021. [CrossRef]
- 108. Rampado, O.; Depaoli, A.; Marchisio, F.; Gatti, M.; Racine, D.; Ruggeri, V.; Ruggirello, I.; Darvizeh, F.; Fonio, P.; Ropolo, R. Ef-fects of different levels of CT iterative reconstruction on low-contrast detectability and radiation dose in patients of different sizes: An anthropomorphic phantom study. *Radiol. Med.* **2021**, *126*, 55–62. [CrossRef] [PubMed]
- 109. Schicchi, N.; Fogante, M.; Palumbo, P.; Agliata, G.; Esposto Pirani, P.; Di Cesare, E.; Giovagnoni, A. The sub-millisievert era in CTCA: The technical basis of the new radiation dose approach. *Radiol. Med.* **2020**, *125*, 1024–1039. [CrossRef]

J. Pers. Med. **2022**, 12, 955 21 of 21

110. Palumbo, P.; Cannizzaro, E.; Bruno, F.; Schicchi, N.; Fogante, M.; Agostini, A.; De Donato, M.C.; De Cataldo, C.; Giovagnoni, A.; Barile, A.; et al. Coronary artery disease (CAD) extension-derived risk strati-fication for asymptomatic diabetic patients: Usefulness of low-dose coronary computed tomography angiography (CCTA) in detecting high-risk profile patients. *Radiol. Med.* 2020, 125, 1249–1259. [CrossRef]

- 111. Hussein, M.A.M.; Cafarelli, F.P.; Paparella, M.T.; Rennie, W.J.; Guglielmi, G. Phosphaturic mesenchymal tumors: Radiological aspects and suggested imaging pathway. *Radiol. Med.* **2021**, *126*, 1609–1618. [CrossRef]
- 112. Danti, G.; Flammia, F.; Matteuzzi, B.; Cozzi, D.; Berti, V.; Grazzini, G.; Pradella, S.; Recchia, L.; Brunese, L.; Miele, V. Gastrointestinal neuroendocrine neoplasms (GI-NENs): Hot topics in morphological, functional, and prognostic imaging. *Radiol. Med.* 2021, 126, 1497–1507. [CrossRef]
- 113. Karmazanovsky, G.; Gruzdev, I.; Tikhonova, V.; Kondratyev, E.; Revishvili, A. Computed tomography-based radiomics approach in pancreatic tumors characterization. *Radiol. Med.* **2021**, *126*, 1388–1395. [CrossRef]
- 114. Fusco, R.; Petrillo, M.; Granata, V.; Filice, S.; Sansone, M.; Catalano, O.; Petrillo, A. Magnetic Resonance Imaging Evaluation in Neoadjuvant Therapy of Locally Advanced Rectal Cancer: A Systematic Review. *Radiol. Oncol.* 2017, 51, 252–262. [CrossRef]
- 115. Fusco, R.; Sansone, M.; Granata, V.; Setola, S.V.; Petrillo, A. A systematic review on multiparametric MR imaging in prostate cancer detection. *Infect. Agent Cancer* **2017**, *12*, 57. [CrossRef]
- 116. Granata, V.; Fusco, R.; Avallone, A.; Filice, F.; Tatangelo, F.; Piccirillo, M.; Grassi, R.; Izzo, F.; Petrillo, A. Critical analysis of the major and ancillary imaging features of LI-RADS on 127 proven HCCs evaluated with functional and morphological MRI: Lights and shadows. *Oncotarget* 2017, 8, 51224–51237. [CrossRef] [PubMed]
- 117. Granata, V.; Grassi, R.; Fusco, R.; Belli, A.; Cutolo, C.; Pradella, S.; Grazzini, G.; La Porta, M.; Brunese, M.C.; De Muzio, F.; et al. Diagnostic evaluation and ablation treatments assessment in hepatocellular carcinoma. *Infect. Agent Cancer* **2021**, *16*, 53. [CrossRef]
- 118. Barabino, M.; Gurgitano, M.; Fochesato, C.; Angileri, S.A.; Franceschelli, G.; Santambrogio, R.; Mariani, N.M.; Opocher, E.; Carrafiello, G. LI-RADS to categorize liver nodules in patients at risk of HCC: Tool or a gadget in daily practice? *Radiol. Med.* **2021**, *126*, 5–13. [CrossRef] [PubMed]
- 119. Granata, V.; Fusco, R.; Filice, S.; Catalano, O.; Piccirillo, M.; Palaia, R.; Izzo, F.; Petrillo, A. The current role and future prospectives of functional parameters by diffusion weighted imaging in the assessment of histologic grade of HCC. *Infect. Agent Cancer* **2018**, 3, 23. [CrossRef] [PubMed]
- 120. Orlacchio, A.; Chegai, F.; Roma, S.; Merolla, S.; Bosa, A.; Francioso, S. Degradable starch microspheres transarterial chemoembolization (DSMs-TACE) in patients with unresectable hepatocellular carcinoma (HCC): Long-term results from a single-center 137-patient cohort prospective study. *Radiol. Med.* 2020, 125, 98–106. [CrossRef]