## Photosystem II photoinhibition and photoprotection in a lycophyte, Selaginella martensii

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## **Supporting information**

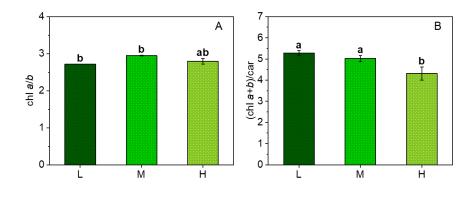
**Fig. S1.** Photosynthetic pigments quantification in *Selaginella martensii* acclimated to deep shade (L), intermediate shade (M) and full sunlight (H).

**Fig. S2.** Native thylakoid composition in *Selaginella martensii* acclimated to deep shade (L), intermediate shade (M) and full sunlight (H).

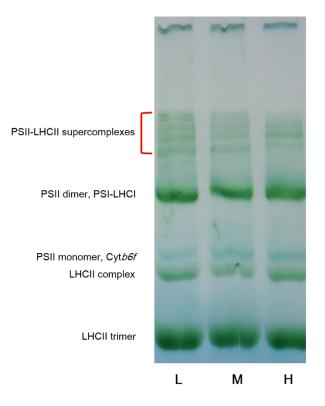
**Fig. S3.** Examples of minimum fluorescence as a function of Non-Photochemical Quenching (NPQ) in *Selaginella martensii*.

Fig. S4. NPQ kinetics curves of S. martensii plants recorded at different light intensities.

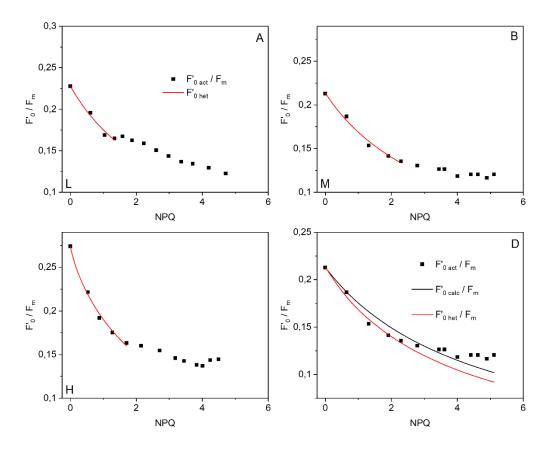
**Fig.S5.** Photochemical quenching measured in the dark in *S. martensii* plants acclimated to deep shade (L), intermediate shade (M) and full sunlight (H) upon independent exposure to increasing irradiances.



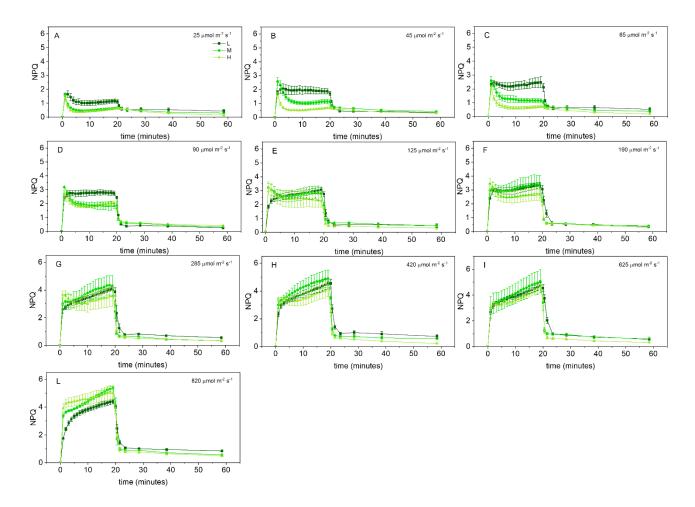
**Fig. S1.** Photosynthetic pigments quantification in *Selaginella martensii* acclimated to deep shade (L), intermediate shade (M) and full sunlight (H). Pigment ratios in isolated thylakoids are reported as the molar ratio between chlorophyll (chl) *a* and *b* content (A) and chlorophyll *a*+*b* and carotenoids (B). Histograms represent average values ±Standard Error for *n*=3. Different letters indicate a significant difference determined with ANOVA followed by Tukey's test at p<0.05.



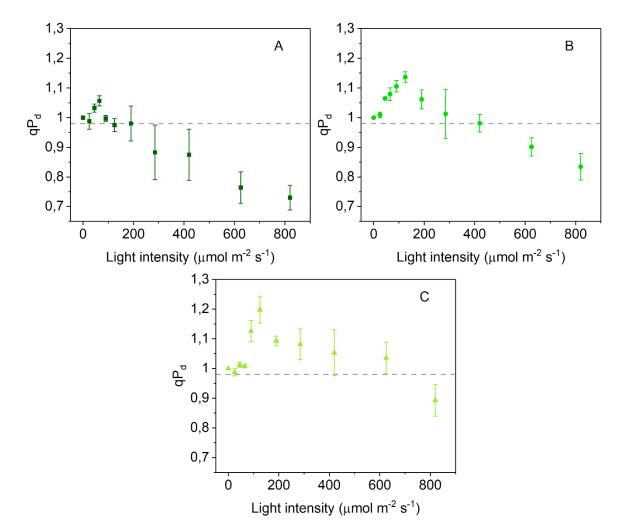
**Fig. S2.** Native thylakoid composition in *Selaginella martensii* acclimated to deep shade (L), intermediate shade (M) and full sunlight (H). Thylakoids corresponding to 8 mg of chlorophyll were solubilized with 1.5%  $\beta$ -dodecylmaltoside to a final chlorophyll concentration of 0.5 mg ml<sup>-1</sup> and subjected to Blue-Native Polyacrylamide gel Electrophoresis (BN-PAGE). Electrophoretic bands were identified according to Järvi *et al.* (2011) and Ferroni *et al.* (2014). The heavy PSII-LHCII supercomplexes are more abundant in L plants than in M or H plants. In contrast, the intensity of the band corresponding to the LHCII trimers is almost invariable in the three types of plants. The thylakoid organization in the plant material used for the fluorometric experiments is therefore uniform to that used in Ferroni *et al.* (2016).



**Fig. S3.** Examples of minimum fluorescence as a function of Non-Photochemical Quenching (NPQ) in *Selaginella martensii* acclimated to deep shade (L), intermediate shade (M) and full sunlight (H). F'<sub>0</sub> is expressed relative to its respective maximum fluorescence level ( $F_m$ ). (A-C) experimental, actual values of minimum fluorescence (F'<sub>0</sub>, black squares) together with the fitting function of F'<sub>0</sub> accounting for a heterogeneous antenna (red line) in the three light regimes. The fitting procedure involves only the values of F'<sub>0 act</sub> drop as a function of NPQ is still approximately hyperbolic only in M and H plants (A-B), while the decrease in L plants is linear (C). (D) Example of F'<sub>0 act</sub> (black squares), F'<sub>0 calc</sub> (black line, homogeneous antenna, Oxborough and Baker 1997) and F'<sub>0 het</sub> (red line, heterogeneous antenna, Ware *et al.* 2015) relative to the sample in (B).



**Fig. S4.** Kinetic curves of NPQ during 19 minutes of continuous actinic light illumination followed by 40 minutes of dark relaxation in *Selaginella martensii* plants acclimated to deep shade (L), intermediate shade (M) and full sunlight (H). Levels of light intensity: 25, 45, 65, 90, 125, 190, 285, 420, 625, 820  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>. Average values ±Standard Error for *n* = 3-6.



**Fig. S5.** Light curves of  $qP_d$  as function of light intensity.  $qP_d$  was calculated according to Ruban and Murchie (2012) in *Selaginella martensii* plants acclimated to deep shade (L), intermediate shade (M) and full sunlight (H).  $qP_d$  values were selected at specific times of the induction curve in order to closely match the number of photons that reached the samples at the end of each light interval of the light curve (the one treated in the paragraph "In sun plants higher photoprotection is accompanied by extensive PSII antenna uncoupling")

## References

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Järvi, S., Suorsa, M., Paakkarinen, V., & Aro, E.M. (2011). Optimized native gel systems for separation of thylakoid protein complexes: novel super-and mega-complexes. *Biochemical Journal*, 439(2), 207-214.

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Ware, M.A., Belgio, E., & Ruban, A.V. (2015). Photoprotective capacity of non-photochemical quenching in plants acclimated to different light intensities. *Photosynthesis Research*, 126(2), 261-274.