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New insights into perfluorinated adsorbents for analytical and bioanalytical applications

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Abstract Perfluorinated (*F*-) adsorbents are generally prepared by bonding perfluoro-functionalized silanes to silica gels. They have been employed for a long time essentially as media for solid-phase extraction of *F*-molecules or *F*-tagged molecules in organic chemistry and heterogeneous catalysis. More recently, this approach has been extended to proteomics and metabolomics. Owing to their unique physicochemical properties, namely fluorophilicity and proteinophilicity, and a better understanding of some fundamental aspects of their behavior, new applications of *F*-adsorbents in the field of environmental science and bio-affinity studies can be envisaged. In this article, we revisit the most important features of *F*-adsorbents by focusing, in particular, on some basic information that has been recently obtained through (nonlinear) chromatographic studies. Finally, we try to envisage new applications and possibilities that *F*-adsorbents will allow in the near future.

Keywords HPLC · Perfluoroalkyl acids · Endocrine disruptors · Trace elements · Surfactants · Fluorous

Introduction

Highly fluorinated compounds (*F*-compounds) are characterized by the presence in their structure of a portion in which a

substantial number of hydrogen atoms (typically 7 to 20) attached to carbon atoms are replaced with fluorine atoms. This gives the molecules specific properties that are different from those of their parent hydrocarbon analogs. In the terminology of fluorous chemistry a portion or domain of a molecule rich in sp^3 carbon–fluorine bonds is termed a fluorous label or tag (more specifically, if at least six fully fluorinated sp^3 carbons are present, the *F*-portion is referred to as a “ponytail”) [1].

F-alkyl chains are bulkier and more rigid than alkyl chains (cross section of around 30 \AA^2 vs. about 20 \AA^2 for alkyl chains) and adopt a helical-like structure in place of the typical planar zigzag structure of alkyl chains. The stiffness of *F*-alkyl chains is claimed to be responsible, on the one hand, for their ordered stacking and, on the other, for their slower equilibration and exchange kinetics compared with those of alkyl chains [2–5].

F-alkyl chains are more hydrophobic than alkyl chains of similar length. According to the Hildebrand and Scott solubility scale, for instance, the solubility parameter (that strongly correlates with polarity) for *F*-alkanes is roughly $5 \text{ cal}^{1/2} \text{ cm}^{3/2}$, whereas it is 7 for *n*-alkanes (and 15 for water) [6]. In addition, *F*-alkyl chains possess a lipophobic character (sometimes referred to as oleophobicity [1]) and are less polarizable than the corresponding hydrocarbons, as indicated by their Kamlet–Taft dipolarity/polarizability parameters and by lower refractive indexes than hydrocarbons [7]. These characteristics, together with the strength of the C–F bond and the enhanced electroattracting character of fluorine (that reinforces the C–C backbone), explains the well-known chemical inertness and thermal stability of perfluorocarbons [1].

Perfluorocarbons have recently found important applications in many fields of research, including synthesis, catalyst technology, adsorption/purification processes, materials chemistry, and biomedical applications. Table 1 lists some of the most popular perfluorinated materials for applications in HPLC and fluorous solid-phase extraction (F-SPE).

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t1.1 **Table 1** Commercially available perfluorinated stationary phases for both HPLC and F-SPE applications

t1.2	Brand name	Provider	Bonding phase	Particle size (µm)	Pore size (Å)	End-capped
t1.3	FluoroFlash	Fluorous Technologies Inc.	C ₈ F ₁₇	5; 40	60	No
t1.4	FluoroSep-RP Octyl	ES	C ₈ F ₁₇	5	60	N/A
t1.5	Fluofix	Wako	C ₆ F ₁₃ -branched	5	120; 300	No, yes
t1.6		Thermo	C ₆ F ₁₃ -branched	5	100; 300	Yes
t1.7	Fluophase RP/WP	Thermo	C ₆ F ₁₃	5	100; 300	Yes
t1.8	Tridecafluoro	Silicycle	C ₆ F ₁₃	40; 63	60	Yes
t1.9	FluoroSep-RP Propyl	ES	C ₃ F ₇	5	300	N/A
t1.10	Fluorochrom	Silicycle	N/A	40; 63	60	No

N/A not available

71 **Perfluorinated (F-) adsorbents**

72 **F-silica gels and fluorophilicity**

73 F-silica gels have the general structure silica–O–
 74 Si(Me)₂(CH₂)_n R_f, where n is either 2 or 3 and R_f is C₆F₁₃ or
 75 C₈F₁₇ [1]. Sometimes, pentafluorophenyl (PFP)-functional-
 76 ized silica gels have been considered F-materials but, strictly
 77 speaking, they are not and will not be considered in this
 78 review.

79 Since their first appearance in the early 1980s, F-silica gels,
 80 thanks to their extreme hydrophobicity and low polarity, were
 81 claimed to be ideal candidates as adsorbents for the reversed-
 82 phase (RP) separation of large biomolecules under
 83 nondenaturing conditions (i.e., with minimum amount of
 84 organic modifier in the mobile phase) [8, 9]; however, the
 85 field has not been explored in depth and F-adsorbents have
 86 never actually been considered a real alternative to traditional
 87 RP phases, such as octyl- (C₈) or octadecyl-functionalized
 88 (C₁₈) silica gels. In contrast, F-silica gels have been applied
 89 as adsorbents for F-SPE and separation of fluorous molecules
 90 from non-fluorous ones and from each other [10, 11]. The
 91 unique ability of F-molecules to recognize other molecules
 92 possessing an F-portion is referred to as fluorous affinity or
 93 fluorophilicity. It arises from selective, strong noncovalent
 94 interactions between the perfluoroalkyl segments of mole-
 95 cules, in a sort of “like dissolves like” interaction.

96 On the basis of the concept of fluorophilicity, a series of
 97 cutting edge applications of F-adsorbents have appeared in
 98 recent years in the omics sciences [12, 13], food chemistry
 99 [14], environmental science [15], amongst others.

100 **Perfluoro-selectivity through liquid chromatography studies**

101 Fluorophilicity is generally quantified by fluorous/organic
 102 liquid/liquid partition coefficients [1]. High-performance liq-
 103 uid chromatography (HPLC) offers an alternative approach to
 104 evaluate fluorous phase affinity, when a perfluoroalkyl sta-
 105 tionary phase is used.

Indeed, the dependence of the logarithm of the chromato-
 graphic retention factor (*k*) upon the number of perfluoro-
 carbon units (*n*_{CF₂}) in the backbone chain of a perfluoroalkyl
 homologous series (at fixed mobile phase composition) per-
 mits the estimation of the change of Gibbs free energy, Δ*G*_{CF₂}
 , for the transfer from the mobile to the stationary phase of a
 perfluoromethylene group as follows [15–18]:

$$\Delta G_{CF_2} = -RT \frac{d \ln k}{dn_{CF_2}} = -RT \ln \alpha_{CF_2} \quad (1)$$

where α_{CF₂}, *R* and *T*, are the selectivity, the universal gas
 constant, and the absolute temperature, respectively. As an
 example of this approach, Fig. 1 shows how ln *k* changes by
 changing the amount of acetonitrile in the mobile phase for
 four perfluoroalkyl acids of environmental concern. In the
 region delineated by green points, ln *k* decreases quasi-
 linearly with increasing organic modifier. At each mobile
 phase composition, the slope of the ln *k* vs. *n*_{CF₂} plot gives
 the natural logarithm of the selectivity (for the sake of clarity,
 the case of 50 % acetonitrile in the mobile phase is illustrated
 in Fig. 1). Table 2 reports Δ*G*_{CF₂} values (calculated by this
 approach through Eq. 1) as a function of the mobile phase
 composition for two different stationary phases, a traditional
 octadecyl and a straight-chain perfluorinated one [17]. Even if
 the transfer of the CF₂ moiety from the mobile to the station-
 ary phase is thermodynamically favorable on both phases at
 all mobile phase compositions (its value always being nega-
 tive), the ability of the F-adsorbent to “recognize”, and thus to
 stabilize, this moiety is markedly larger than that of the C₁₈
 phase. This is demonstrated by absolute values of Δ*G*_{CF₂}, on
 average much larger (+70 %) on the F-adsorbent than on the
 C₁₈ one.

Another example of the improved selectivity of fluorinated
 stationary phases over hydrocarbon ones towards the separa-
 tion of fluorinated solutes is given in Fig. 2, where chromato-
 grams of the separation of benzene and five fluorinated ana-
 logues on the two phases are compared [19].

142 Adsorption of organic compounds from multicomponent
143 mixtures on *F*-adsorbents

144 The adsorption of organic compounds from multicomponent
145 mixtures on *F*-adsorbents can be measured by several tech-
146 niques, including HPLC [20, 21]. When the adsorption iso-
147 therm is measured through chromatography, the information
148 can be given either as the excess or absolute isotherm. The
149 quantity directly measured in an adsorption experiment is the
150 excess (usually indicated by Γ). It is defined as the excess of
151 solute contained in the adsorption system (considered as a
152 whole) compared to what would be present in a hypothetical
153 system where solute concentration is uniform throughout the
154 whole volume of the eluent and equal to the equilibrium
155 concentration in the bulk phase of the real system [22–24].
156 On the other hand, the total adsorbed amount (q) is the amount
157 of solute contained in an adsorbed layer of finite thickness.
158 Clearly the definition of the adsorbed layer is arbitrary and
159 needs the adoption of some convention [25, 26]. Figure 3
160 shows both types of isotherms measured for the adsorption
161 of acetonitrile from water/acetonitrile binary mixtures on a
162 straight-chain perfluorohexylethylsiloxane-bonded stationary
163 phase [27]. Acetonitrile is strongly adsorbed by the *F*-phase
164 with a saturation value of roughly 13 μmol of acetonitrile
165 adsorbed per square meter of solid. The negative excess of
166 acetonitrile at organic-rich mobile phase compositions corre-
167 sponds to a positive excess of water on the surface. This is due
168 to the adsorption by unreacted silanols, which remain on the
169 silica surface after its functionalization, that, under these con-
170 ditions, have not been saturated yet [27].

171 Adsorption isotherm data allow one to estimate several
172 characteristic properties of the system under examination,
173 including solvent fluorophilicity, interfacial tension at the
174 solid/liquid interface, and wetting properties, which all have
175 important implications for technological applications of catal-
176 ysis, material engineering, environmental science, the
177 fluorotelomer industry, pollution research, etc. as will be
178 further discussed in the “Outlook” section.

179 Retention mechanisms in liquid chromatography
180 with *F*-stationary phases

181 A recent study focusing on the chromatographic behavior of
182 silica-based *F*-adsorbents revealed that, under typical RP con-
183 ditions (with aqueous acetonitrile eluents), the major features
184 described for these phases can be understood and rationalized
185 in terms of traditional liquid–solid chromatographic models
186 based (1) on the formation of a mixed stationary phase and
187 (2) partitioning of solutes between the mobile and this station-
188 ary phase [17]. As an example, the so-called U-shape retention
189 behavior of *F*-adsorbents (i.e., the U-shaped dependence of $\ln k$
190 with increasing amount of acetonitrile in the mobile phase, see
191 Fig. 1), which in some cases has been described as a sort of

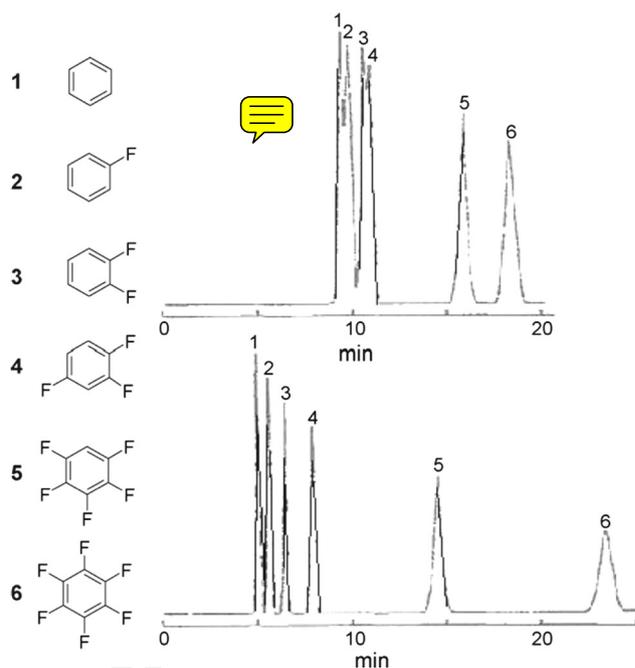


Fig. 1 3D plot showing the dependence of $\ln k$ and both the mobile phase composition (expressed as volume fraction of acetonitrile) and number of perfluorocarbon units in the backbone chain, n_{CF_2} . Sample, mixture of four perfluoroalkyl acids (perfluoropentanoic, $n_{\text{CF}_2} = 4$, perfluorohexanoic, $n_{\text{CF}_2} = 5$, perfluoroheptanoic, $n_{\text{CF}_2} = 6$, perfluorooctanoic $n_{\text{CF}_2} = 7$). Column, perfluorohexylpropylsiloxane-bonded silica (Fluophase-RP from Thermo Scientific); mobile phase, water/acetonitrile mixtures (+0.1% v/v formic acid); temperature, 298 K. Adapted from ref. [15]

192 peculiar characteristic of these materials [28], can be explained
193 by considering a mixed-mode retention mechanism in which
194 both fluorophilic (hydrophobic) and silanophilic (hydrophilic)
195 interactions are simultaneously present (exactly as happens
196 with C_{18} silica-based adsorbents) [17].

197 Although *F*-adsorbents certainly exhibit a different selec-
198 tivity than traditional RP adsorbents (e.g., C_{18} or C_8), the true
199 peculiarity of *F*-adsorbents is when they are used with *F*-
200 compounds. Fluorophilicity can be thus modulated by careful
201 choice of the eluent composition [15, 17]. With aqueous/
202 acetonitrile mixtures, fluorophilicity can be maximized by
203 maximizing the content of water in the mobile phase (to
204 reduce the competitive adsorption of acetonitrile). It has been
205 demonstrated, however, that to allow the complete wetting
206 [29] of the (meso)porous structure of silica-based *F*-adsorb-
207 ents, a minimum amount of organic (approx. 5–10 % in
208 volume) is necessary in the eluent [30].
209

Outlook

210
211 In this section, we briefly present our views on the future of *F*-
212 adsorbents by trying to anticipate new solutions and

t2.1 **Table 2** Free-energy change, ΔG_{CF_2} , for the transfer from the mobile to the stationary phase of a CF_2 unit as a function of mobile phase (MP) composition (expressed as acetonitrile volume fraction) on an F - and an octadecyl stationary phase

t2.2	MP	F -alkyl (J/mol)	C_{18} (J/mol)
t2.3	0.5	-2,239	-1,399
t2.4	0.6	-2,008	-1,209
t2.5	0.7	-1,777	-1,053
t2.6	0.8	-1,679	-902
t2.7	0.9	-1,427	-761

Adapted from ref. [17]

F -alkyl, perfluorohexylpropylsiloxane-bonded silica, Fluophase-RP from Thermo Scientific; C_{18} , octadecylethyl-bridged hybrid-organic/inorganic, BEH- C_{18} from Waters; temperature, 298 K

213 opportunities that, in our opinion, will be offered by these
214 materials in several different fields of research.

215 The most important area in which we believe F -adsorbents
216 will contribute to the advancement of knowledge and technical
217 know-how is environmental chemistry. In particular, we
218 are thinking about the numerous classes of perfluoroalkyl and
219 polyfluoroalkyl substances and their several homologues and
220 isomers. Concern about the effects of F -compounds on the
221 environment and human health has dramatically increased in
222 recent years as these compounds are toxic, extremely resistant
223 to degradation, bioaccumulate in food chains, and can have
224 long half-lives in humans [31]. In spite of the complexity and
225 variety of these compounds, the interest of the scientific
226 community has focused almost exclusively on
227 perfluoroalkylcarboxylic and perfluoroalkylsulfonic acids.
228 Even though numerous studies have been published in the
229 literature and much information has been gathered about the
230 sources, fate, transport, and toxicity of these species, some
231 fundamental aspects of their physicochemical properties and

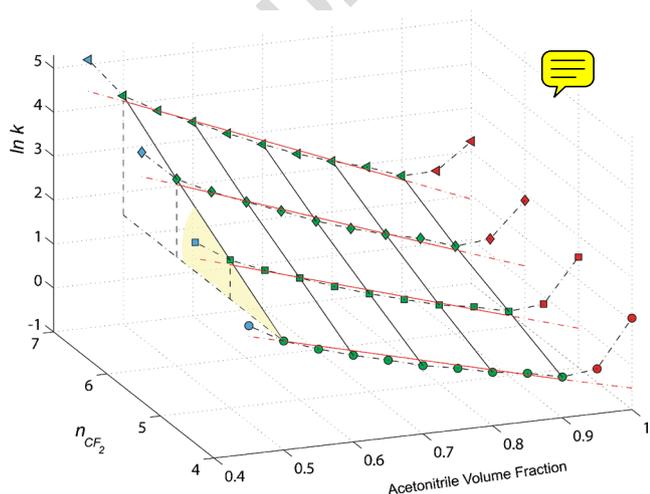


Fig. 2 Chromatographic separation of a mixture of benzene and five different fluorinated analogues on a C_{18} (top) and a C_8F_{17} (bottom) column. Taken with permission from ref. [19]

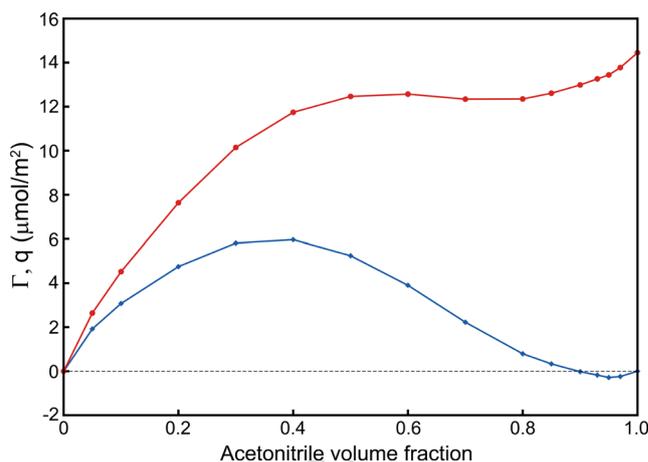


Fig. 3 Excess (Γ , blue line) and absolute (q , red line) adsorption isotherm of acetonitrile from water/acetonitrile binary mixtures on a straight-chain perfluorohexylethylsiloxane-bonded stationary phase. Temperature, 298 K. Adapted from ref. [27]

232 partitioning behavior are poorly understood and widely debat-
233 ed. As an example, the pK_a value of perfluorooctanoic acid is
234 reported to vary from 0 (i.e., a strong acid) to about 4 (rela-
235 tively weak acid) [32–34]. Since transport properties in the
236 environment are strictly dependent on the chemical form of
237 molecule (in this case ionic or neutral), and thus on pH, one
238 understands how fundamental research in the field is still
239 needed.

240 There are also many other F -compounds, whose presence
241 in the environment has been recently demonstrated (e.g.,
242 perfluoroalkane sulfinic acids and perfluoroalkyl phosphinic
243 and phosphonic acids [35]), for which the scenario is even
244 worse because practically no studies have been performed on
245 them.

246 Still another example is the class of fluorotelomers and
247 fluorotelomer-based products recently brought into the spot-
248 light [36]. This class includes, among others, many
249 antistaining and antiwetting agents, which are widely
250 employed in everyday life. Nevertheless, systematic studies
251 about their stability and degree of exposure in both humans
252 and the environment not only to them but also to their degra-
253 dation products are substantially missing.

254 As is illustrated by these examples, many questions about
255 F -compounds are unanswered and others will arise as more is
256 learned about these ubiquitous anthropogenic substances [31].
257 In the near future, it is reasonable to anticipate that there will
258 be an increasing demand by both the scientific community and
259 control and regulatory agencies for efficient, selective, and
260 easy-to-automate analytical methods and tools for the deter-
261 mination, monitoring, and removal of F -compounds in ma-
262 trixes of different origin (including biological samples). In all
263 these cases, the potential of F -adsorbents is evident. Owing to
264 their intrinsic affinity towards F -compounds, F -adsorbents
265 look like being the perfect counterpart for the separation and
266 capture of these species [15, 29, 30].

267 Apart from the already demonstrated use in proteomics
 268 [12] and metabolomics [13] for the separation of fluorourous-
 269 tagged molecules, another field where we consider the use of
 270 *F*-adsorbents to be potentially very useful is as stationary
 271 phases for bioaffinity chromatographic studies. This consid-
 272 eration comes from the evidence that *F*-compounds preferen-
 273 tially bioaccumulate in body compartments high in protein
 274 content (this property of *F*-compounds is known as
 275 proteinophilicity), such as the liver, kidney, and blood. For
 276 instance, it has been demonstrated that human serum albumin
 277 (HSA), the most abundant protein in blood plasma, binds
 278 through specific high affinity interactions with several *F*-com-
 279 pounds [37–39]. Thus, one might imagine using these adsor-
 280 bents either directly as supports for binding studies of proteins
 281 by means of nonlinear chromatographic techniques [40] or as
 282 a sort of pre-fractionation system (possibly in-line) for prote-
 283 ome analysis of low-abundance proteins [41]. Indeed these
 284 proteins, whose diagnostic potential is very relevant, are often
 285 extremely difficult to detect because of the masking presence
 286 of high-abundance serum proteins.

287 Finally, another field of application where *F*-adsorbents
 288 can be useful is heterogeneous catalysis for investigating and
 289 designing new recovery strategies of fluorourous catalysts and
 290 reagents without using fluorourous solvents. This is strictly con-
 291 nected to the possibility of studying the affinity of different
 292 solvents, including supercritical CO₂, or multicomponent sol-
 293 vent mixtures towards *F*-materials through dynamic
 294 (chromatographic) adsorption studies.

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