

Journal of Medical Entomology

Scanning Electron Microscopy Investigations of Third Instar Larva of Cordylobia rodhaini (Diptera: Calliphoridae), Agent of Furuncular Myiasis

Journal:	Journal of Medical Entomology
Manuscript ID:	JME-2014-0010.R1
Manuscript Type:	Research Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Pezzi, Marco; University of Ferrara, Life Sciences and Biotechnology Cultrera, Rosario; University of Ferrara, Medical Sciences, Section of Dermatology and Infectious Diseases Chicca, Milvia; University of Ferrara, Life Sciences and Biotechnology Leis, Marilena; University of Ferrara, Life Sciences and Biotechnology
Please choose a section from the list :	Morphology, Systematics, Evolution
Field Keywords:	Medical Entomology
Organism Keywords:	Calliphoridae

SCHOLARONE™ Manuscripts Pezzi et al.: Scanning electron microscopy of *Cordylobia rodhaini*

Journal of Medical Entomology Morphology, Systematics, Evolution M. Leis

Dep. of Life Sciences and Biotechnology

University of Ferrara

Via Luigi Borsari 46, 44121 Ferrara (Italy)

Phone: ++39 532-455313 Fax: ++39 532-249761

E-mail: marilena.leis@unife.it

Scanning Electron Microscopy Investigations of Third Instar Larva of *Cordylobia rodhaini* (Diptera: Calliphoridae), Agent of Furuncular Myiasis

M. Pezzi¹, R. Cultrera², M. Chicca¹ and M. Leis¹

¹ Department of Life Sciences and Biotechnology, University of Ferrara, Via Luigi Borsari 46, 44121, Ferrara, Italy

² Department of Medical Sciences, Section of Dermatology and Infectious Diseases, University of Ferrara, Via Fossato di Mortara 64/B, 44121, Ferrara, Italy

1	Abstract
2	A scanning electron microscopy study of the 3 rd larval instar of <i>Cordylobia rodhaini</i> Gedoelst
3	(Diptera: Calliphoridae), causing obligatory furuncular myiasis, is presented here for the first time.
4	The larvae were collected from a patient exposed to them in the tropical rainforest of Kibale
5	National Park (Uganda). Distinctive features are described in sequence from the anterior region to
6	the posterior region, highlighting the morphological features of antennae, maxillary palps, structures
7	related to mouth opening, sensory structures, thoracic and abdominal spines, anterior and posterior
8	spiracles. The results are compared with those of other Calyptrata flies, mainly from the family
9	Calliphoridae and, when possible, with C. anthropophaga Blanchard (Diptera: Calliphoridae), the
10	only other species of genus Cordylobia investigated by scanning electron microscopy.
11	
12	only other species of genus <i>Cordylobia</i> investigated by scanning electron microscopy. Keywords Cordylobia rhodaini, SEM, 3 rd instar larva, morphology, furuncular myiasis
13	
14	
15	Keywords
16	Cordylobia rhodaini, SEM, 3 rd instar larva, morphology, furuncular myiasis
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	

27	Introduction
28	Cordylobia rodhaini Gedoelst (Diptera: Calliphoridae) is a fly species causing an obligatory
29	furuncular myiasis. C. rodhaini is distributed in tropical Africa, mostly in rainforest. The species
30	typically parasitizes mammals, especially monkeys, rodents and small antelopes. Humans are
31	occasionally infested but the myiasis by C. rodhaini is far less common than that caused by C.
32	anthropophaga Blanchard (Diptera: Calliphoridae) (Zumpt 1965). These two species, with C.
33	ruandae Fain (Diptera: Calliphoridae) are the only three species of the genus Cordylobia Gruenberg
34	(Veraldi et al. 2014). Since 1902 only 25 cases of human furuncular myiasis by C. rodhaini have
35	been reported (Pezzi et al. 2014). Studies on morphology of 3 rd instar <i>C. rodhaini</i> were previously
36	performed by stereomicroscopy and light microscopy (Gedoelst 1909, Rodhain 1915, Bertram
37	1938, Fain 1953, Zumpt 1965, Kremer et al. 1970, Pampiglione et al. 1991): however,
38	ultrastructural analyses of body surface by scanning electron microscopy (SEM) have never been
39	performed. Here we describe by SEM the 3 rd instar larva of <i>C. rodhaini</i> to provide a more detailed
40	description of this dipteran of medical and veterinary importance.
41	
42	Materials and Methods
43	Five larvae identified as 3 rd instar <i>C. rodhaini</i> based on previous stereomicroscopic observations
44	were collected from a patient who was likely exposed to them in June 2014 while traveling in the
45	tropical rainforest of Kibale National Park (Uganda) (Pezzi et al. 2014). The larvae were prepared
46	according to the following procedure: they were fixed in 3% glutaraldehyde in sodium cacodylate
47	buffer 0.125 M, pH 7.3, for 24 hours, rinsed three times for 15 min in the same buffer and postfixed
48	in 2% osmium in sodium cacodylate buffer for 24h. After postfixation the samples were rinsed three
49	times for 15 min in the buffer, then dehydrated in a graded ethanol series (12 h for each grade) and
50	critical point dried in a Balzers CPD 030 dryer (Leica Microsystems, Wetzlar, Germany). The
51	samples were then mounted on stubs and coated with gold-palladium in a Sputter Coated S150
52	Edwards (HHV Ltd, Crawley, UK). All SEM observations were conducted at the Electronic

Microscopy Centre of University of Ferrara on a Zeiss EVO 40 SEM (Zeiss, Milan, Italy). For each morphological aspect such as thoracic and abdominal spines at least 20 samples were examined in all five specimens. Two of the larvae were dissected for detailed observations of the anterior and posterior ends. The area of posterior spiracles and cuticular depressions was measured in the posterior end.

58

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

53

54

55

56

57

59 Results

The 3rd instar larva of *C. rodhaini* is barrel-shaped with a tapering anterior end, a cylindrical pseudocephalon and truncated posterior end with two posterior spiracles (Fig. 1A-D), in agreement with previous light and stereomicroscopy data (Gedoelst 1909, Rodhain 1915, Bertram 1938, Fain 1953, Zumpt 1965, Kremer et al. 1970, Pampiglione et al. 1991). The pseudocephalon shows two prominent lobes (Fig. 1B, C), the pseudocephalic lobes, each one bearing an antero-lateral maxillary palpus and a dorsal antenna (Fig. 2A). The maxillary palpus contains a cluster of sensilla located at the top of a conic structure: each cluster has a knob sensillum, a pit sensillum, four papillary sensilla and a bilobed protrusion. Behind the cluster of sensilla the maxillary palpus has a basiconic sensillum and a coeloconic sensillum in dorsal position (Fig. 2B). The antenna has a blunt conic shape composed of two structures, a conic basal ring and a distal dome (Fig. 2C). Two mouth hooks ("denticles") with broad tips are visible in the anterior ventral part of the pseudocephalon (Fig. 1B, C). On both sides of the mouth hooks there is a group of oral ridges with 9-10 lobes, each one covered by tiny conic denticles with 1-3 tips (Fig. 1B and 2D, E). In the centre of the oral ridges there is the ventral organ composed of a circular pit sensillum, a small pit sensillum with an opening partially surrounded by a ridge, and a button-shaped papillary sensillum (Fig. 2F). Between the mouth hooks, along the dorsal-ventral axis there is a cluster of conic preoral denticles with down curved tips (Fig. 2G). The mouth opening has a crescent shape with a triangular labium (Fig. 2H), showing in the lower part two trichoid sensilla with one seta and 3-4 dome-shaped papillary sensilla (Fig. 2H-J).

The first segment of the thoracic region is divided into two bands: the anterior one is covered by
cuticular spines, while the posterior one is thinly folded (Fig. 3A). The spines, pointing towards the
posterior end of the body, are drop-shaped and $60.8 \pm 11.0~\mu m$ long in the dorsal part (Fig. 3B) and
sharply conic and $40.1 \pm 12.6~\mu m$ long in the ventral part (Fig. 3C). In the posterior band devoid of
spines there are several sensilla, either coeloconic (Fig. 3D) or campaniform (Fig. 3E). In the
second thoracic segment there are some campaniform sensilla similar to those of the first thoracic
segment (Fig. 3F). On both sides of the first thoracic segment, near the border with the second
segment, there is an anterior spiracle composed of a ridge with 5-6 domes. (Fig. 3G). As for the first
thoracic segment, the second and third thoracic segments are also covered by cuticle spines pointing
towards the posterior end. However, these spines are all conic and $150.3 \pm 18.5 \mu m$ long (Fig. 1A,
B). Concerning the abdominal segments, they are all completely covered by conic cuticle spines,
$167.2 \pm 12.7 \ \mu m$ long (Fig. 3H). However, the spines of the last two abdominal segments are fewer
and smaller in size: those of the second to last abdominal segment are $69.4 \pm 9.5 \mu m$, while those of
the last segment are $36.6 \pm 11.3~\mu m$ long (Fig. 4A). The last two segments have coeloconic sensilla,
either isolated or in pairs (Fig. 4B, C). There are two fan-shaped posterior spiracles (Fig. 1D and
4D), with area $1.1 \pm 0.3 \times 10^5 \mu m^2$. Each one of them is slightly raised, with three sinuous slits
branching off from a respiratory button partially covered by an operculum (Fig. 4E). Between the
respiratory button and the slits there are 7-8 papillae arranged in a dorsal-ventral row (Fig. 4E and
inlay). Above and below the posterior spiracles there are several cuticular depressions, with area 6.6
$\pm~1.3~x~10^3~\mu m^2$ (Fig. 4F) with deeply corrugated and perforated bottoms (Fig. 4G).

100 Discussion

The morphological studies by SEM of the larvae of the genus *Cordylobia* are very limited (Gall et al. 1987, Bettoli et al. 1993) and the present one is the first on *C. rodhaini*. The only two studies available deal with *C. anthropophaga*: the first one describes the cephalic region, cuticle spines and posterior region of a 2nd instar (Bettoli et al. 1993) and the second one the cephalic region and the

first thoracic segment of an unidentified instar (probably the third one) (Gall et al. 1987).
Comparing the results of SEM observations on C. rodhaini 3 rd instar to those obtained in C.
anthropophaga, it is possible to detect several differences. For example, in C. rodhaini the mouth
hooks are more rounded and no cluster of conic denticles or oral grooves are found in C .
anthropophaga, which in place of the oral grooves exhibits triangular spines similar to those of the
thoracic segments (Gall et al. 1987). The cluster of conic denticles appears typical of C. rodhaini,
although all three larval instars of Cochlyomyia macellaria Fabricius (Diptera: Calliphoridae) have
dental sclerites in the same position (Mendonça et al. 2014a). The grouping of oral grooves in C.
rodhaini resembles that of the 3 rd instar of Wohlfahrtia magnifica (Schiner) (Diptera:
Sarcophagidae) (Ruíz-Martínez et al. 1990). In C. rodhaini these accessory structures of the mouth
hooks, together with thoracic and abdominal spines, probably help the larva to feed and later exit
the host and burrow in the soil to pupate. Moreover, the morphology of antennae of 3^{rd} instar C .
rodhaini appears similar to that of the 2 nd and 3 rd instar of C. anthropophaga (Gall et al. 1987) and
of 3 rd instars of other species of the family Calliphoridae (Sandeman et al. 1987; Leite et al. 1993;
Sukontason et al. 2005, 2006, 2008; Mendonça et al. 2010, 2014a, 2014b; Ubero-Pascal et al. 2012;
Jang et al. 2013). The cluster of maxillary palp sensilla on the top of a conic structure found in C .
rodhaini is also present in C. anthropophaga (Gall et al. 1987). Concerning the ventral organ, no
mention or description of this structure is found in previous works describing the ultrastructural
morphology of C. anthropophaga, therefore the presence of this organ and its morphology is
described for the first time in the genus Cordylobia. In C. rodhaini the ventral organ is different in
comparison to 3 rd instars of other Calliphoridae, characterized by short spines (Sukontason et al.
2003a, Ubero-Pascal et al. 2012). No description of the labium is available in C. anthropophaga but
the labium of <i>C. rodhaini</i> is not divided in lobes as in 3 rd instar <i>Chrysomya rufifacies</i> (Macquart)
(Diptera: Calliphoridae) (Sukontason et al. 2003b).
The SEM investigation confirms previous data on 3 rd instar C. rodhaini showing a different
morphology of cuticle spines according to their position (Bertram 1938, Fain 1953, Pampiglione et

al. 1991). However, the SEM images show two types of cuticle spines in the first thoracic segment.
one drop-shaped in the dorsal region and the other sharply conic in the ventral region. Moreover,
comparing the cuticle spines of the first thoracic segment of 3 rd instar C. rodhaini with those of the
same segment in C. anthropophaga, the C. anthropophaga spines are all conic with a wider base
(Gall et al. 1987).
The cuticular depressions found in the last two abdominal segments of 3 rd instar C. rodhaini have
not been described in C. anthropophaga but similar structures have been described in the second
thoracic segment of 3 rd instar <i>Gasterophilus nasalis</i> (L.) (Diptera: Oestridae) (Leite et al. 1999).
Similar structures have also been found in the last abdominal segment of 2 nd instar Calliphora
vicina Robineau-Desvoidy (Diptera: Calliphoridae) and, less evident, in the 3 rd instar of the same
species (Ubero-Pascal et al. 2012). Concerning the sensilla, the campaniform ones found in the first
and second thoracic segment of 3 rd instar C. rodhaini are similar to the "papillae" of 3 rd instar
Lucilia sericata (Meigen) (Diptera: Calliphoridae) (Shaheen et al. 2004) while the coeloconic ones
are similar to those described in 2 nd instar C. anthropophaga (Bettoli et al. 1993). Besides
confirming the data obtained during previous observations on 3 rd instar <i>C. rodhaini</i> (Gedoelst 1909,
Bertram 1938, Fain 1953, Zumpt 1965, Kremer et al. 1970, Pampiglione et al. 1991), the SEM
analyses of posteriors spiracles shows the presence of an "operculum" in proximity of the button
spiracles of the respiratory plates and of a row of basiconic sensilla between the respiratory button
and the sinous slits. No ultrastructural description of posterior spiracles is available for 3^{rd} instar C .
anthropophaga. This study provides ultrastructural information useful to discriminate 3 rd instar C.
rodhaini from the same instar of C. anthropophaga. No data by SEM are presently available on the
other species of the genus Cordylobia, C. ruandae. More ultrastructural investigations are required
on larval instars of this genus in order to extend the morphological details able to discriminate
among its three species responsible of furuncular myiasis.

Acknowledgemen	nts
----------------	-----

We would like to thank Gilberto Grandi, Professor of Zoology at the Department of Life Sciences and Biotechnology, University of Ferrara (Italy) for critically reviewing the manuscript and providing relevant suggestions. We owe thanks also to Dr Elisa Rota, Dr Maria Rita Bovolenta and Jhm. Miss Paola Boldrini for kind technical assistance. This study was funded by the municipality of Ferrara (Ferrara, Italy).

162

156

157

158

159

160

163	References cited
164	Bertram, D. S. 1938. A note upon myiasis due to the larvae of Cordylobia rodhaini Gedoelst. Ann
165	Trop. Med. Parasit. 32: 431-345.
166	Bettoli, V., S. Trimurti, M. Leis, and A. Virgili. 1993. Miasi cutanea da tumbu fly (Cordylobia
167	anthropophaga): studio ultrastrutturale. G. Ital. Dermatol. Venereol. 128: 243-246.
168	Fain, A. 1953. Notes sur Cordylobia rodhaini Ged. Etude des terminalia des Mouches adultes et
169	pieces chitineuses de la partie antérieure de la larve. Rev. Zool. Bot. Afr. 48: 306-312.
170	Gall, Y., C. Pecquet, P. Litoux, and H. Barrière. 1987. Myase furonculoïde à Cordylobia
171	anthropophaga. Examen en microscopie électronique à balayage. Ann. Dermatol. Venereol. 114:
172	59-63.
173	Gedoelst, L. 1909. Cordylobia rodhaini nova species, diptère congolais à larve cuticole. Arch.
174	Parasit. 13: 538-547.
175	Jang, M., S. M. Ryu, S. C. Kwon, J. O. Ha, Y. H. Kim, D. H. Kim, S. M. Jung, S. I. Lee, W. M
176	Sohn, H. J. Cha, and M. Ock. 2013. A case of oral myiasis caused by Lucilia sericata (Diptera:
177	Calliphoridae) in Korea. Korean J. Parasitol. 51: 119-123.
178	Kremer, L., J. Lenys, M. Basset, H. Rombourg, and B. Molet. 1970. Deux cas de myiase à
179	Cordylobia rodhaini contractée au Cameroun et diagnostiquée en Alsace. Bull. Soc. Pathol. Exot.
180	63: 592-596.
181	Leite, A. C. R., and J. D. E. Guevara. 1993. Scanning electron microscopy of the larval instars of
182	Cochliomyia hominivorax. Med. Vet. Entomol. 7: 263-270.
183	Leite, A. C. R., F. B. Scott, L. G. Evangelista. 1999. Scanning electron microscope observations
184	on third-instar Gasterophilus nasalis (Diptera: Oestridae). J. Med. Entomol. 36: 643-648.
185	Mendonça, P. M., J. R. dos Santos-Mallet, and M. M. Queiroz. 2010. Ultramorphological
186	characteristics of immature stages of <i>Chrysomya albiceps</i> (Wiedemann 1819) (Diptera:
187	Calliphoridae), a fly specie of forensic importance. Microsc. Res. Tech. 73: 779-784.

- Mendonça, P. M., R. R. Barbosa, L. B. Cortinhas, J. R. dos Santos-Mallet, and M. M. de
- 189 Carvalho Queiroz. 2014a. Ultrastructure of immature stages of *Cochliomyia macellaria* (Diptera:
- 190 Calliphoridae), a fly of medical and veterinary importance. Parasitol. Res. 113: 3675-3683.
- 191 Mendonça, P. M., R. R. Barbosa, C. Carriço, L. B. Cortinhas, J. R. dos Santos-Mallet, and M.
- 192 **M. Queiroz. 2014b.** Ultrastructure of immature stages of *Lucilia cuprina* (Diptera: Calliphoridae)
- using scanning electron microscopy. Acta Trop. 136: 123-128.
- 194 Pampiglione, S., S. Schiavon, G. Candiani, and M. L. Fioravanti. 1991. Osservazioni cliniche e
- parassitologiche su di un caso di miasi foruncolosa disseminata da Cordylobia rodhaini nell'uomo
- in Etiopia. Parassitologia 33: 159-167.
- 197 **Pezzi, M., R. Cultrera, M. Chicca, and M. Leis. 2014.** Furuncular myiasis maused by *Cordylobia*
- 198 rodhaini (Diptera: Calliphoridae): a case report and a literature review. J. Med. Entomol. (in press),
- 199 doi 10.1093/jme/tju027.
- 200 **Rodhain, J. 1915.** Sur la biologie de *Stasisia rodhaini* Gedoelst (*Cordylobia rodhaini*). C.R. Acad.
- 201 Sci. Paris 161: 323-325.
- 202 Ruíz-Martínez, I., M. D. Soler-Cruz, R. Benítez-Rodríguez, J. M. Pérez-Jiménez, C. Adalid-
- Fuentes, M. Díaz-López. 1990. Scanning electron microscope study of Wohlfahrtia magnifica
- 204 (Schiner, 1862) (Diptera: Sarcophagidae). I. Structures with parasitic and possible taxonomic
- meaning. Scanning Microsc. 4: 103-109.
- Sandeman, R. M., B. J. Collins, and P. R. Carnegie. 1987. A scanning electron microscope study
- of *L. cuprina* larvae and the development of blowfly strike in sheep. Int. J. Parasitol. 17: 759-765.
- Shaheen, M. S., A. I. Hamza, and S. M. Abdel-Rahman. 2004. Light and scanning electron
- 209 microscopy of third-instar larva of *Lucilia sericata* (Diptera: Calliphoridae) recovered from a case
- of human intestinal myiasis. Assiut Vet. Med. J. 51: 127-135.
- 211 Sukontason, K. L., K. Sukontason, S. Piangjai, N. Boonchu, T. Chaiwong, R. C. Vogtsberger,
- B. Kuntalue, N. Thijuk, and J. K. Olson. 2003a. Larval morphology of *Chrysomya megacephala*
- 213 (Fabricius) (Diptera: Calliphoridae) using scanning electron microscopy. J. Vector Ecol. 28: 47-52.

214	Sukontason.	, K. L.	, K. Sukontason	, S.	Lertthamnongtham,	B.	. Kuntalue	, N.	Thij	uk,	R.	C.

- Vogtsberger, and J. K. Olson. 2003b. Surface ultrastructure of *Chrysomya rufifacies* (Macquart)
- 216 larvae (Diptera: Calliphoridae). J. Med. Entomol. 40: 259-267.
- 217 Sukontason, K. L., R. C. Vogtsberger, N. Boonchu, T. Chaiwong, D. Sripakdee, R. Ngern-
- Klun, S. Piangjai, and K. Sukontason. 2005. Larval morphology of *Chrysomya nigripes* (Diptera:
- Calliphoridae), a fly species of forensic importance. J. Med. Entomol. 42: 233-240.
- 220 Sukontason, K. L., S. Piangjai, W. Boonsriwong, N. Bunchu, R. Ngern-klun, R. C.
- Vogtsberger, and K. Sukontason. 2006. Observations of the third instar larva and puparium of
- 222 Chrysomya bezziana (Diptera: Calliphoridae). Parasitol. Res. 99: 669-674.
- 223 Sukontason, K. L., P. Sribanditmongkol, T. Chaiwong, R. C. Vogtsberger, S. Piangjai, K.
- 224 **Sukontason. 2008.** Morphology of immature stages of *Hemipyrellia ligurriens* (Wiedemann)
- (Diptera: Calliphoridae) for use in forensic entomology applications. Parasitol. Res. 103: 877-887.
- Ubero-Pascal, N., R. López-Esclapez, M. D. García, and M. I. Arnaldos. 2012. Morphology of
- preimaginal stages of *Calliphora vicina* Robineau-Desvoidy, 1830 (Diptera, Calliphoridae): A
- comparative study. Forensic Sci. Int. 219: 228-243.
- Veraldi, S., S. M. Serini, and L. Süss. 2014. Three cases of cutaneous myiasis caused by
- 230 Cordylobia rodhaini. J. Infect. Dev. Ctries. 8: 249-251.
- **Zumpt, F. 1965.** Myiasis in man and animals in the world. Butterworth & Co., London, UK.

233	Figure Legends
234	Fig. 1. Scanning electron micrographs of 3 rd instar <i>Cordylobia rodhaini</i> . (A) Entire body in ventral
235	view, an asterisk indicates the second to last segment and the diamond indicates the last segment.
236	(B) Anterior region in lateral-ventral view. (C) Anterior region in dorsal view. (D) Posterior end of
237	body. AD: abdomen; AO: anal opening; AS: anterior spiracle; CAS: campaniform sensillum; CCD:
238	conic preoral denticles; CS: coeloconic sensillum; LA: labium; MH: mouth hooks; OR: oral ridges;
239	PL: pseudocephalic lobe; POS: posterior spiracles; PSE: pseudocephalon; TR: thorax; TR1: first
240	thoracic segment; TR2: second thoracic segment.
241	Fig. 2. Scanning electron micrographs of 3 rd instar <i>C. rodhaini</i> . (A) Pseudocephalic lobe: the dashed
242	line outlines the maxillary palpus. (B) Maxillary palpus: the dashed line outlines the cluster of
243	sensilla at the top of the conic structure. (C) Antenna: the asterisk indicates the distal dome and the
244	diamond indicates the conic basal ring. (D) Oral ridges and ventral organ. (E) Details of the oral
245	ridges with tiny conic denticles. (F) Details of the ventral organ. (G) Details of the conic preoral
246	denticles. (H) Mouth opening: the arrowheads indicate the dome-shaped papillary sensilla and the
247	arrows indicate the trichoid sensilla. (I) Trichoid sensillum in the lower part of the labial lobe. (J)
248	Dome-shaped papillary sensilla of the lower labial lobe. AN: antenna; BP: bilobed protrusion; BS:
249	basiconic sensillum; KS: knob sensillum; PIS: pit sensillum; PIS1: circular pit sensillum; PIS2: pit
250	sensillum with ridge; PS: papillary sensillum; PSC: papillary sensilla within a cluster; VO: ventral
251	organ. Other abbreviations as in Fig. 1.
252	Fig. 3. Scanning electron micrographs of 3 rd instar <i>C. rodhaini</i> . (A) First thoracic segment, dorsal
253	view: the dashed line outlines the two bands of the first thoracic segment, and the arrowheads
254	indicate the campaniform sensilla. (B) Details of cuticular spines of the first thoracic segment,
255	dorsal region. (C) Details of cuticular spines of the first thoracic segment, ventral region. (D)
256	Coeloconic sensillum of the first thoracic segment. (E) Campaniform sensillum of the first thoracic
257	segment. (F) Campaniform sensillum of the second thoracic segment. (G) Anterior spiracle: the

258	asterisk indicates the dome line. (H) Conic cuticular spines covering the abdomen with exception of
259	the second to last and last segments. AB: anterior band; PB: posterior band.
260	Fig. 4. Scanning electron micrographs of 3 rd instar <i>C. rodhaini</i> . (A) Details of the lateral region of
261	the second to last abdominal segment (asterisk) and of the last abdominal segment (diamond). (B)
262	Coeloconic sensillum of the last abdominal segment. (C) A pair of coeloconic sensilla. (D) Posterior
263	region of the last abdominal segment: the rectangle outlines the region enlarged in F and the
264	arrowheads indicate the cuticular depressions. (E) Respiratory button: the arrowheads indicate the
265	papillary sensilla, the inlay shows an enlarged papillary sensillum. (F) Detail of Fig. 4D showing
266	the cuticular depressions. (G) Details of the bottom of a cuticular depression. CD: cuticular
267	depression; OP: operculum; RB: respiratory button; SS: sinuous slit. Other abbreviations as in Fig.
268	1.
269	

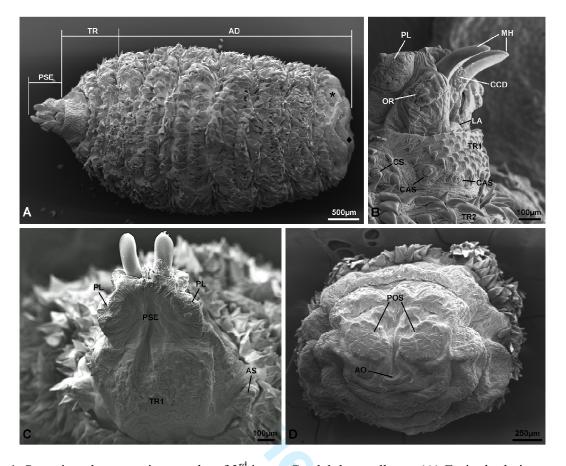


Fig. 1. Scanning electron micrographs of 3rd instar *Cordylobia rodhaini*. (A) Entire body in ventral view, an asterisk indicates the second to last segment and the diamond indicates the last segment. (B) Anterior region in lateral-ventral view. (C) Anterior region in dorsal view. (D) Posterior end of body. AD: abdomen; AO: anal opening; AS: anterior spiracle; CAS: campaniform sensillum; CCD: conic preoral denticles; CS: coeloconic sensillum; LA: labium; MH: mouth hooks; OR: oral ridges; PL: pseudocephalic lobe; POS: posterior spiracles; PSE: pseudocephalon; TR: thorax; TR1: first thoracic segment; TR2: second thoracic segment.

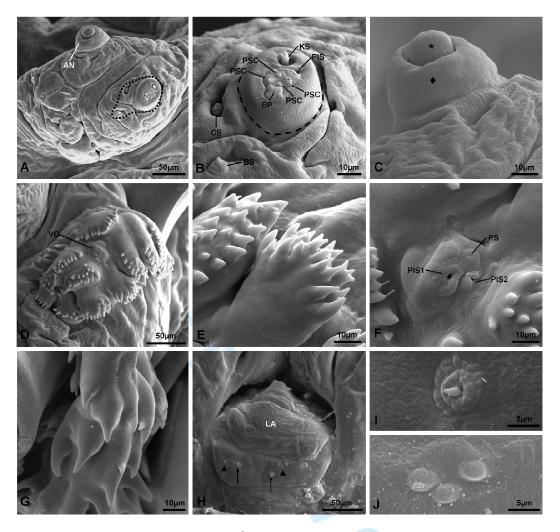


Fig. 2. Scanning electron micrographs of 3rd instar *C. rodhaini*. (A) Pseudocephalic lobe: the dashed line outlines the maxillary palpus. (B) Maxillary palpus: the dashed line outlines the cluster of sensilla at the top of the conic structure. (C) Antenna: the asterisk indicates the distal dome and the diamond indicates the conic basal ring. (D) Oral ridges and ventral organ. (E) Details of the oral ridges with tiny conic denticles. (F) Details of the ventral organ. (G) Details of the conic preoral denticles. (H) Mouth opening: the arrowheads indicate the dome-shaped papillary sensilla and the arrows indicate the trichoid sensilla. (I) Trichoid sensillum in the lower part of the labial lobe. (J) Dome-shaped papillary sensilla of the lower labial lobe. AN: antenna; BP: bilobed protrusion; BS: basiconic sensillum; KS: knob sensillum; PIS: pit sensillum; PIS1: circular pit sensillum; PIS2: pit sensillum with ridge; PS: papillary sensillum; PSC: papillary sensilla within a cluster; VO: ventral organ. Other abbreviations as in Fig. 1.

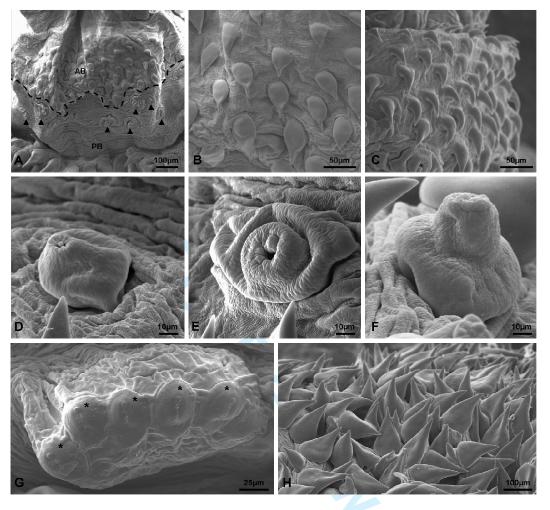


Fig. 3. Scanning electron micrographs of 3rd instar *C. rodhaini*. (A) First thoracic segment, dorsal view: the dashed line outlines the two bands of the first thoracic segment, and the arrowheads indicate the campaniform sensilla. (B) Details of cuticular spines of the first thoracic segment, dorsal region. (C) Details of cuticular spines of the first thoracic segment, ventral region. (D) Coeloconic sensillum of the first thoracic segment. (E) Campaniform sensillum of the first thoracic segment. (F) Campaniform sensillum of the second thoracic segment. (G) Anterior spiracle: the asterisk indicates the dome line. (H) Conic cuticular spines covering the abdomen with exception of the second to last and last segments. AB: anterior band; PB: posterior band.

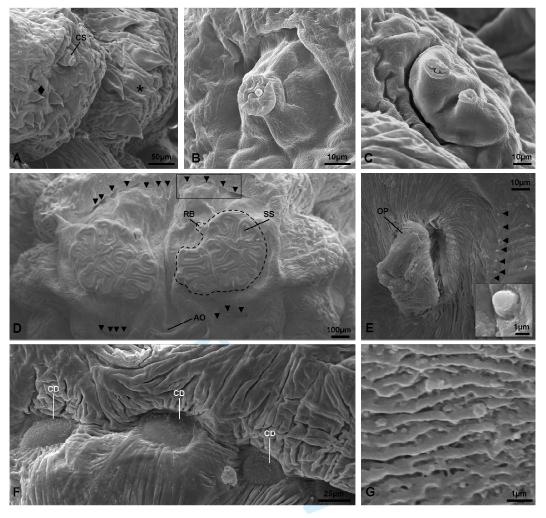


Fig. 4. Scanning electron micrographs of 3rd instar *C. rodhaini*. (A) Details of the lateral region of the second to last abdominal segment (asterisk) and of the last abdominal segment (diamond). (B) Coeloconic sensillum of the last abdominal segment. (C) A pair of coeloconic sensilla. (D) Posterior region of the last abdominal segment: the rectangle outlines the region enlarged in F and the arrowheads indicate the cuticular depressions. (E) Respiratory button: the arrowheads indicate the papillary sensilla, the inlay shows an enlarged papillary sensillum. (F) Detail of Fig. 4D showing the cuticular depressions. (G) Details of the bottom of a cuticular depression. CD: cuticular depression; OP: operculum; RB: respiratory button; SS: sinuous slit. Other abbreviations as in Fig. 1.