

# Cutting-edge biotechnological advancement in islet delivery using pancreatic and cellular approaches

Magdy Elnashar\*, 1,2, Mauro Vaccarezza & Hani Al-Salami

- <sup>1</sup>Biotechnology & Drug Development Research Laboratory, School of Pharmacy & Biomedical Sciences, Curtin Health Innovation Research Institute, Curtin University, Perth, Western Australia, Australia
- <sup>2</sup>Centre of Excellence, Department of Polymers, National Research Centre, Cairo, Egypt
- <sup>3</sup>School of Pharmacy & Biomedical Science, Faculty of Health Sciences, Curtin University, Bentley, Perth, WA 6102, Australia
- \*Author for correspondence: magmel@gmail.com

There are approximately 1 billion prediabetic people worldwide, and the global cost for diabetes mellitus (DM) is estimated to be \$825 billion. In regard to Type 1 DM, transplanting a whole pancreas or its islets has gained the attention of researchers in the last few decades. Recent studies showed that islet transplantation (ILT) containing insulin-producing  $\beta$  cells is the most notable advancement cure for Type 1 DM. However, this procedure has been hindered by shortage and lack of sufficient islet donors and the need for long-term immunosuppression of any potential graft rejection. The strategy of encapsulation may avoid the rejection of stem-cell-derived allogeneic islets or xenogeneic islets. This review article describes various biotechnology features in encapsulation-of-islet-cell therapy for humans, including the use of bile acids.

Lay abstract: The global cost for diabetes mellitus (DM) is estimated to be \$825 billion. With regard to Type 1 DM, transplanting pancreas whole/islets have gained the attention of researchers in the last few decades. Recent studies have demonstrated that islet transplants containing insulin-producing  $\beta$  cells is the most notable advancement cure for Type 1 DM. However, this procedure has been hampered by islet donor shortage and the need for long-term immunosuppressive therapy. An encapsulation strategy could overcome these shortcomings. Here, we discuss various biotechnology features in the encapsulation of islet cell therapy for humans including the use of bile acids.

First draft submitted: 13 June 2020; Accepted for publication: 29 October 2020; Published online: 23 November 2020

**Keywords:** auto-transplantation • bile acid • diabetes treatment • encapsulation • human pancreas • pancreatic islets • stem cells • transplantation • Type 1 diabetes

According to the World Health Organization, diabetes occurs when the pancreas produces too little insulin for the body's requirements, or when the body cannot use insulin effectively. Chronic diabetic complications impair the metabolic processing of fats, carbohydrates and electrolytes, which disrupts the vascular system [1]. Examples of microvascular complications are myocardial infarction and retinopathy (e.g., blindness), and for vascular complications are nephropathy and neuropathy (e.g., foot amputation). Type 2 diabetes mellitus (T2DM) is the most prevalent (~85%), and it occurs due to tissue resistance or lack of insulin. These issues can be managed by lifestyle changes (physical exercise + diet), insulin, oral antidiabetic drugs and noninsulin injections. Patients suffering from T1DM are insulin-dependent, and rely on insulin injection.

The number of diabetic patients dramatically increased from 1980 to 2014, from 108 to 422 million [2] and between 2012 and 2015, there were 3.8 million diabetes-related deaths. That number could increase greatly, as there are now approximately one billion pre-diabetic people. The diagnosis and treatment of T1DM is more costly than T2DM, and the global cost for DM is estimated to be \$825 billion [3,4].

Despite ongoing research in multiple laboratories worldwide, islet transplantation (ILT) remains confined to clinical trials rather than common practice, using robust commercially available therapeutics. The latest studies



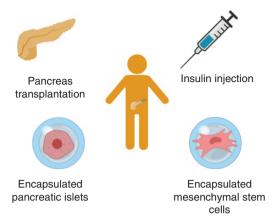


Figure 1. Possible strategies for the treatment and management of Type 1 diabetes mellitus: insulin injection, pancreas transplantation, encapsulated pancreatic islets and encapsulated mesenchymal stem cells.

on transplanting the pancreas (whole organ or islets) in patients with T1DM with severe glycemic variability found it possible to restore the endocrine functions of the pancreas [5]. Combining total pancreatectomy with islet autotransplantation was another strategy [6]. The application of bile acids (BAs) in islet encapsulation and transplantation has gained interest, and laboratory results show significant promise, not only in maintaining high cellular viability but also rejuvenation postencapsulation, cell bioenergetics, insulin release and cell-cell interaction necessary for acceptable performance, short and long term [7,8].

Recent advancements in the treatment/management of T1DB also include encapsulated pancreatic islets and encapsulated mesenchymal stem (MSCs), as shown in Figure 1.

# Pancreatic cell transplantation compared with whole-pancreas transplantation Pancreas transplantation

Transplanting the whole pancreas from a healthy donor to a diabetic patient requires major surgery. The process is invasive, and over the past 30 years, research activity has led to some standardized protocols to achieve this process without increasing the rate of hypoglycemic events in the recipient [9–12]. Since 1966, the rate of implant postsurgery survival has been well maintained at 76% after 1 year, 62% after 3 years and 50–70% after 5 years [13]. Over the last decade, the number of solitary operations has remained stable with a steady rate of graft failure, especially in the last 5 years [14].

## Limitations of pancreatic transplantation

According to Ramesh & Brayman, the procedure of pancreatic transplantation (PCT), which is generally performed concurrently with kidney transplantation, includes some surgical and postsurgical complications. The major complications involve graft pancreatitis/thrombosis, and formation of pseudocysts and pancreatic fistulae [15]. Finding a donor is another issue.

# Islet transplantation

ILT comes as an alternative, and is considered as an upgraded way to treat T1DM, in contrast to injection with insulin or whole-pancreas transplantation. It requires approximately 2% (w/w) of the total pancreas (endocrine part) that contains β cells in a healthy person to be transplanted into a diabetic person with reasonable success. ILT can be infused using a catheter through the portal venous access, which represents a minimally invasive process and is also associated with marginal or no complications [16]. Another positive aspect of ILT is that it can deliver glycemic control without the risk of hypoglycemia and the use of exogenous insulin. For instance, Shapiro has explored ILT and its potential use, and has demonstrated that ILT does possess the ability and validity to correct HbA1c concentrations to certain values that may reverse the secondary consequences of diabetes [17], while insulin administration via pump or similar set-ups may result in fewer or attenuated effects on concentrations of the diabetic biomarker HbA1C [18]. Thompson *et al.* found that ILTs are more effective in decreasing the progression of microvascular diseases (e.g., retinopathy) and vascular diseases (e.g., nephropathy) related to diabetes than is rigorous medical therapy [19]. Although this method is effective, seems simpler and less invasive, it does have some limitations.

A recent study of the Clinical Islet Transplantation (CIT) Consortium protocol 07 (CIT-07) has shown the results in 48 adults with T1D for more than 5 years [20–22]. Patients were suffering from severe hypoglycemic events (SHEs) and impaired awareness of hypoglycemia (IAH). Purified human pancreatic islets (PHPI) were transplanted at eight centres in North America, and each patient had been administered an immunosuppressive and PHPI. The median HbA1c level was 5.6% at 1–2 years with no death or disability noted [21]. The overall effectiveness of the CIT-07 trial [20,22] has been further demonstrated by the results by Foster *et al.*, showing that we can potentially maintain blood glucose control without experiencing severe cases of hypoglycemia, and hence improved quality of life via successful ILT that is safe and robust [21]. Of note, large decreases in diabetes-related distress and fear of hypoglycemia were observed. Furthermore, quality of life and functional health status did not worsen, despite ongoing immunosuppression. General measurements of health status, such as Short Form 36 Health Survey (SF-36) and mental summary score (MCS), demonstrated statistically significant differences from baseline status. The CIT-07 result gives a new perspective on ILT: a prospective assessment of health utility and health status outcomes among transplant recipients in a Phase-III study.

#### Limitations of ILT

Long-term causes of graft loss can be listed as:

# Immunosuppression-associated factors

Patients with ILT are normally prescribed immune-suppressive medications (systematic immunosuppressive treatment) to avoid allogeneic rejections. Immunosuppressive medication should be taken in optimized doses to avert graft loss. However, even the optimized medications can be directly toxic to the ILT or cause dysfunction of other tissue organs after long-term usage [23]. In addition, systemic immune-suppressive medications such as cyclosporine and tacrolimus may also increase the patient's risk of developing cancer, organ damage or infection. Histological studies found that ILT triggers recurrent autoimmune effects that can result in cell destruction [24,25]. The use of a specific antigen to inhibit the immune response through the initiation of regulatory T cells, has shown some promising results in avoiding the development of the response [26]. However, in the USA from 1999 to 2007, less than 400 ILTs were performed out of  $\sim$ 1.25 M cases nationwide, and that was linked to the high risk of chronic immunosuppression which presently outweighs the potential benefits of ILT [27,28].

#### Nonimmunosuppression-associated factors

ILTs are engrafted in the liver through the portal system, and the process may encounter islet-graft loss due to hypoxia and/or instant blood-mediated inflammatory factors (INMIR), which may lead to reduced islet mass and poor islet quality and function.

- Hypoxia [29]
  - Islet cells are damaged due to hypoxia, as the process of isolation causes de-vascularization in the islet cells. Also, islets are implanted into the liver in low oxygen tension (Carlsson *et al.*). There is the indirect effect of the hypoxic environment stimulating the innate immune system, resulting in the release of the inflammatory cytokines interferon (IFN), IL-1 and TNF [30].
- Instant blood-mediated inflammatory reaction (IBMIR) [31] IBMIR causes activation of the coagulation system, resulting in serious problems by disruption [32] and destruction of 60% of transplanted islets [33]. In general, poor clinical outcomes of ILT are frequently related to high IBMIR [30].

The expertise in both ILT and pancreas transplantation is not widely available. This issue renders problematic any decision in regard to therapy, based on the pros and cons of islet versus pancreas transplantation. As a result, diagnosis and management of patients with clinical diabetic symptoms has been guided mostly by local expertise [5]. Recently, Maffi and Secchi, have proposed a protocol based on Shapiro *et al.* [34] to define when to carry out islet or pancreas transplantation and in many scenarios, there was a degree of overlap. Pros and cons for pancreas versus ILT are summarized in Table 1.

In summary, they suggest that ILT is preferred for patients with:

- Severe glycemic problems that cannot be controlled using insulin therapy
- Unstable Type 1 diabetes

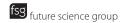


Table 1. Comparison between the pros and cons of whole-pancreas transplantation versus islet transplantation.		
	Islet transplantation	Pancreas transplantation
Biological cost (organ consumption)	Needs several donors	Needs one donor
Biological cost (technical complications during follow-up)	No major complications (~3.1%)	Major complications ( $\sim$ 10%): bleeding, thrombosis, morbidity and duodenal leaks
Insulin-independence	Delayed process (months-years). 44% is achieved at 3 years	Speedy process. 61% is achieved at 3 years
Surgical complications	No risk	High risk (e.g., pre-existing of cardiovascular disease)

- Hypoglycemia unawareness
- Cardiovascular disease

The most notable advancement cure for T1DM is ILT containing insulin-producing  $\beta$  cells [35,36]. ILT has shown the potential to be one of the most promising solutions for the treatment of T1DM, and that has encouraged many ILT Centres to start license applications for clinical allogeneic ILT. The inflammation of pancreatic  $\beta$  cells and the paucity of cell viability after ILT are the biggest challenges to researchers [37–39].

# Total pancreatectomy & islet autotransplantation

Another potential possible therapeutic approach could be islet autotransplantation which considers only patients with chronic pancreatitis (CP), and is usually preceded by total removal of pancreas (pancreatectomy) [6]. Recent observations by Berman *et al.* show that diabetic candidates with severe painful CP are valid candidates for pancreatectomy [40]. This observation is of paramount significance, especially among candidates and patients with autogenous pancreatic islet transplantation (TP-IAT), in that it has not been readily accessible until recent years before which patients were reported to transplant centres during later stages of their disease. While there has been a growing utilization of TP-IAT for patients with refractory CP over many years, there remains a lack of consensus clinical guidelines to inform the counselling and management of patients undergoing TP-IAT and its eventual utilization as a valuable method to control diabetes and glycemia.

As highlighted recently by Al-Sofiani *et al.*, there is a need to develop uniform practice guidelines and to standardize clinical measurement protocols on how to assess patients before and after surgery; moreover, future research should improve islet isolation and engraftment techniques, and validate new biomarkers and imaging tools able to monitor the function, viability and location of the islet cells after engraftment [6,41]. The future could be even more promising in a similar scenario with encapsulated ILT, avoiding immunosuppressive drug treatment (non-systematic immune-suppressive treatment).

# Nonsystematic immune-suppressive treatment

Nonsystematic immunosuppressive (NSI) treatment uses different approaches to reduce local inflammation and generate immune-privileged sites, for example by engineering materials that release factors (prostaglandins [PGE2], IL-10, transforming growth factor-b, SDF and chemokine MCP1) to reduce inflammation [42–44]. This approach also includes stem cell (autologous  $\beta$  cells), immunomodulation (antigen-specific T cells), immune-protective devices, coatings and capsules [45–47]. Immunoprotection using engineered materials such as encapsulation of ILT has received the greatest consideration.

## Bio-nano- & micro-encapsulation of pancreatic islets

Encapsulation of cells is one way to protect these viable cells from the host immune system, post-transplantation. Islet bio-nano- and micro-encapsulation can be carried out by encapsulating the viable, living and functional components within various forms of semipermeable cases/membranes [48–51]. Encapsulation is the ability of the capsule to envelop the cells/islets and protect them from being recognized/identified by the body's own immune system. Entrapment of pancreatic ILT has been widely researched and aimed to protect the pancreatic islets from the environment and to reduce  $\beta$  cells inflammation and death [52,53]. Encapsulation embraces the approach of isolation and shielding of the islet-grafts from the recipient's immune system, which is different to the conventional strategy of ongoing immunosuppressive treatments. That potentially represents a suitable approach for effectively supporting graft functionality and survival [54].

 However, it is obvious that implanting and/or encapsulating biomaterial may still trigger a foreign-body reaction, and may affect the safety of the implanted device. This caveat can considerably impact the short-/long-term tissue responses comprising proteins, cells and other biological components used in regenerative medicine or tissue engineering. Anderson *et al.* [55] have studied the mechanism of foreign-body reaction and the effect of adherent macrophages on the overall inflammatory response to biomaterials. They found that surface chemistry can influence the behavior of macrophages such as cytokine secretion, adhesion, fusion and apoptosis.

In order to encapsulate viable cells or biologically active components including organoids, specialized biomaterials are needed. These biomaterials need to provide support as well as biocompatibility to the encapsulated active moieties including pancreatic islets. The biomaterials need to exhibit a wide range of features including:

- Being porous enough to faciliate and allow nutrients and oxygen permeation into the graft, and also allow waste to leave the graft
- Having the capacity to encapsulate high concentrations of the target molecule
- Acting as an immunostatic or biological barrier to control its core active moieties, while protecting the functionality and performance of the graft
- Being available and from a reliable source, ready for scale-up production
- Having good physical and mechanical stability and regularity
- Not allowing fibrotic growth and subsequent graft failure
- Maintaining long-term cell viability

Scharp and Marchetti have published a detailed review article on the encapsulation of ILT. The review article comprises different types of capsules/materials and devices [47].

# Preparation of macro-/micro-capsules

The most common size and shape for islet entrapment and encapsulation are in the form of disks or tablets, beads or, less commonly, fibers. Gel microcapsules are the most commonly used form of islet encapsulation, since nano-capsules would ultimately provide a huge surface area and more islet interaction with the outside microenvironment. The tiny micro- and nanocapsules can be fabricated by a wide range of technologies such as the Innotech Encapsulator, ionic-gelation methods, Vibrational Jet-Flow Technology, dripping and interphase technique method [56–58]. The Buchi-supported systems have the advantage of strong and comprehensive control over various encapsulating parameters including production rate, being 50–3000 beads/second, based on the blend of rheological and non-Newtonian parameters [59].

## Biopolymers commonly researched for islet encapsulation

# Hydrogels & water-soluble polymers

There is a wide range of various biomaterials and biopolymers currently available for encapsulation of biomolecules, including hydrogels such as alginate, chitosan, cellulose, hyaluronic acid (HA), collagen and carrageenan [60–75].

# Alginate-based capsules

Among all these hydrogels, alginate has been dominating, and is the most commonly used hydrogel for encapsulation of ILT due to its low cost, high availability and durability, nontoxicity to host organisms and its mechanism for encapsulation is well-established [76–78]. Multiple polymers such as alginates and the like have the appropriate properties to form suitable matrices when multivalent cations such as Ca<sup>2+</sup> in aqueous medium are present [57,79].

Encapsulation of cells using alginate has been proven to be efficient [80]. For example, Ludwig et al. and Prochorov et al. have introduced heterologous islets without the use of immunosuppressive drug protocols [81,82]. The main message from these data is the importance of cell viability in the islet graft, which greatly influences the outcomes of the procedure. In particular, the proposed device by Ludwig et al. couples an alginate immune-isolated preparation with an oxygenated chamber system. Oxygenation is pivotal for cell survival and correct cell metabolism: in fact, the graft was effective and well-tolerated without immunosuppression. This contribution demonstrates that encapsulation with regulated/controlled oxygenation (avoiding hypoxia at the level of the encapsulated cells) could be a viable option to improve the efficacy of encapsulated ILT without the use of immunosuppressive drugs. It is worth noting that the use of this oxygenated alginate-encapsulation strategy was applied to allogeneic human ILT in humans for the first time by Ludwig et al. [82].

In another example by Jacobson-Tulleneers-Thevissen *et al.*, they implanted alginate-encapsulated human islet cells into the peritoneal cavity of mice. The results showed that intraperitoneal transplantation had a better result compared with the free implants under the kidney capsule [83]. Free-floating capsules in the peritoneum, without direct contact with host tissue or metabolic correction, and immediately corrected hyperglycemia, were maintained until the end of the study. Recovered floating capsules demonstrated excellent cell viability and secretion, without signs of inflammation/fibrosis in the recipient rodents. These results prompted the authors to translate the concept in a pilot study run on one T1D patient: the main goal of this study was to assess the survival and function of  $\beta$  cells in the encapsulated environment transplanted in the peritoneum. The function and efficacy were confirmed (as in the rodent study), but signs of fibrosis were present, and most of the capsules were found together in large clumps, sticking to the abdominal wall [83].

However, alginate micro- or nanocapsules alone, without any additives, do not seem to act as good-quality micro- and nano-capsules, given their weak physical stability and loss of the mechanical resistance to shear force over long periods of time, that is required and expected post-transplantation [59,84,85]. This is possibly due to alginate's poor ability to adhere to the crosslinking processes and not having sufficient interactive force to form a solid surface resistant to damage or stress [86]. The literature also suggests that alginate gels are not stable in various types of buffers [87], and like other seaweeds, they are susceptible to fibrotic overgrowth when used *in vivo*, resulting in necrosis of the encapsulated cells and pre-mature graft failure [78,88–90]. Many modifications have been made to improve alginate capsules such as ultra-purification, co-encapsulation and surface treatment, but the final results are not yet satisfactory enough to be used at the clinical level [77,89,91–93].

## **HA-based** capsules

HA, on the other hand, and especially HA-based hydrogels, have shown to be very suitable for cell-therapy applications and tissue engineering because of their distinctive biological and mechanical properties [94–96]. Formulations based on HA and other polymers, such as denatured collagen (DCOL), have been found to be durable, with a high shear strength of up to 3500 Pa. A commercial version of HA-DCOL is known as HyStem-C, and it has been used *in vivo* to repair osteochondral defects in rabbits and myocardial infarcts in SCID mice [97–99]. HA is used as a replacement for alginate for the encapsulation of ILT, especially to address graft failure associated with fibrosis [88].

#### Biomimetics of transplantable materials

Biomimetic materials are hybrids/composites of natural materials (e.g., peptides, amino acids, saccharides and proteins) developed using motivation from nature. They have a lower probability of inducing fibrotic growth and higher biocompatibility, which make them suitable for cell encapsulation [100,101]. Unfortunately, these advanced biomimetic materials have low durability and a high degradation rate, which make them unsuitable for ILT.

# BAs as anti-inflammatory compounds

BAs formed from cholesterol catabolism and are metabolically active compounds. The process of bile-acid synthesis involves two main biological pathways and the metabolic and enzymatic activation of approximately 17 hepatic enzymes [102]. After cholesterol catabolism in the liver and their synthesis, BAs are metabolized by the gut microbiota. In the liver, they are conjugated with taurine or glycine amino acids, and the corresponding sodium or potassium salts are known as bile salts. Bile salts are usually stored in the gallbladder and secreted postprandially. Once in the gut, BAs and salts are metabolized further by the gut microbiome, to produce secondary BAs which undergo further re-absorption in a process known as enterohepatic recirculation. The types of BAs and salts exceed one hundred, including deoxycholic acid (DCA), ursodeoxycholic acid (UDCA) and lithocholic acid (LCA). They facilitate the gut uptake of many dietary lipids by their surfactant-like effects [103].

BAs are known to produce various biochemical and biological effects in the body, such as removal of cholesterol and bilirubin, emulsification and solubilization of lipid-soluble vitamins, facilitating their gut and oral uptake [104]. About 0.6 g of BAs/salts are metabolized and produced daily to replace lost BAs excreted in the feces. Among commonly researched BAs is the secondary BA, UDCA, which has been used for many centuries by traditional Chinese therapies [105]. UDCA is currently prescribed for cholestasis and liver cirrhosis [106]. UDCA's mechanism of action and pharmacology are anticipated to be because of its anti-inflammatory and cytoprotective effects [107].

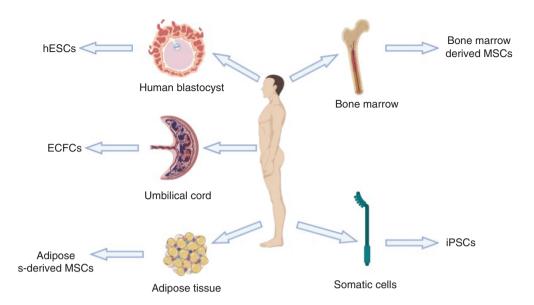


Figure 2. Sources of stem/progenitor cells. Umbilical cord blood can generate ECFCs; human blastocytes can generate hESCs; somatic cells can generate iPSCs; bone marrow can generate bone-marrow-derived mesenchymal stem cells; adipose tissue can generate adipose-derived mesenchymal stem cells.

ECFC: Endothelial colony forming cell; hESC: Human embryonic stem cell; iPSC: Induced pluripotent stem cell; MSC: Mesenchymal stem cell.

# Encapsulation of stem cells

Encapsulation of stem cells (SC) is the latest advancement in encapsulation technology. SCs would be a resolution to our distressing deficiency of donated organs. However, encapsulating SCs is a fairly novel area of research, and this is explains why there are so few studies on encapsulated islets based on SCs for the adjustment of diabetes [108]. Researchers have used embryonic and adult SCs to provide islets for cell-replacement therapy. The ethical issues of using SCs for cell replacement have been widely discussed in the review article by Street *et al.* [109].

Isolated islets in combination with mesenchymal SCs (MSCs), also known as adult SCs, can be isolated from different tissues such as bone marrow, somatic cells and adipose tissue (Figure 2). Isolated islets-co-MSCs have been explored by Shafiee *et al.* due to their potential to differentiate into different cell types. It was found that MSCs have the potential to produce paracrine factors promoting the function and growth of neighbouring cells. Pathak *et al.* have suggested bone marrow and adipocytes, as effective sources of MSCs, to be good options for allogenic MSC cotransplantation with isolated islets [110]. Micro-capsules of co-transplantations of islets with MSCs have been successful in mice, improving islet grafting [111]. This has encouraged many authors in the encapsulation of stem cells [112–114].

## **Conclusion**

Management of T1D using daily insulin injection might fail to achieve optimum glycemic control, and result in acute complications. Transplantations of whole pancreases or islets are presently considered potential options. Some authors have proposed protocols to define when to carry out islet or pancreas transplantation, and, in many scenarios, there is a degree of overlap. However, both methods have their pros and cons. Out of the two scenarios, ILT is considered the future and the more promising solution, therefore many strategies, including the encapsulation of islets andthe use of BAs or stem cells, are under investigation to overcome the limitations associated with ILT. Encapsulation of stem cells (SC) is the latest advancement in encapsulation technology in the past few years, and it seems to be encouraging, albeit still at the mouse level.

## **Future perspective**

ILT to treat diabetes have been widely researched for many years, and despite innovative ideas and dedicated researchers, no commercialable widely available product has made it to the market. The source of viable functional suitable islets remain challenging, and an appropriate matrices remain to be developed. Future perspective needs to focus on revolutionizing our approach to consider avenues of cell reprogramming, cell differentiation, bio-printing,

gene editing, bio-nanotechnological scalable procedures that build on synthetic, semisynthetic and endogenously produced BAs and other compounds, and patient-specific approaches to enable the success of ILTs and wide applications in the clinic. Future studies need to focus on improved delivery systems and matrices that better resemble the human pancreas.

#### **Executive summary**

- Islet transplantation remains limited to Phase1/2 clinical trials with no widely used commercially available products.
- Main challenges to islet transplantation include body immune rejection and lack of appropriate delivery matrices.
- Bile acids have recently been proposed to complement matrices, suitable for islet transplantation.
- Bile acids are endogenously produced in humans and many are considered safe.

#### Acknowledgments

The authors thank Dr D Chaliha for English proofreading.

#### Financial & competing interests disclosure

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

No writing assistance was utilized in the production of this manuscript.

#### Open access

This work is licensed under the Creative Commons Attribution 4.0 License. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/

## References

Papers of special note have been highlighted as: • of interest; •• of considerable interest

- Kolluru GK, Bir SC, Kevil CG. Endothelial dysfunction and diabetes: effects on angiogenesis, vascular remodeling, and wound healing. Int. J. Vasc. Med. 2012, 918267 (2012).
- 2. Mathers CD, Loncar D. Projections of global mortality and burden of disease from 2002 to 2030. PLoS Med. 3(11), e442 (2006).
- 3. Lotfy M, Adeghate J, Kalasz H, Singh J, Adeghate E. Chronic complications of diabetes mellitus: a mini review. *Curr. Diabetes Rev.* 13(1), 3–10 (2017).
- 4. Worldwide trends in diabetes since 1980: a pooled analysis of 751 population-based studies with 4.4 million participants. *Lancet (London, England)* 387(10027), 1513–1530 (2016).
- 5. Maffi P, Secchi A. Islet transplantation alone versus solitary pancreas transplantation: an outcome-driven choice? *Curr. Diabetes Rep.* 19(5), 26 (2019).
- 6. Ong SL, Gravante G, Pollard CA, Webb MA, Illouz S, Dennison AR. Total pancreatectomy with islet autotransplantation: an overview. HPB 11(8), 613–621 (2009).
- Mooranian A, Zamani N, Takechi R et al. An in vivo pharmacological study: variation in tissue-accumulation for the drug probucol as the result of targeted microtechnology and matrix-acrylic acid optimization and stabilization techniques. PloS One 14(4), e0214984 (2019).
- 8. Mooranian A, Raj Wagle S, Kovacevic B et al. Bile acid bio-nanoencapsulation improved drug targeted-delivery and pharmacological effects via cellular flux: 6-months diabetes preclinical study. Sci. Rep. 10(1), 106 (2020).
- Article on the use of bile acids during the nanoencapsulation process of islet transplantation.
- 9. Gruessner AC, Sutherland DE. Pancreas transplant outcomes for United States (US) and non-US cases as reported to the United Network for Organ Sharing (UNOS) and the International Pancreas Transplant Registry (IPTR) as of June 2004. *Clin. Transplant.* 19(4), 433–455 (2005).
- Gruessner AC, Sutherland DE. Pancreas transplant outcomes for United States (US) cases as reported to the United Network for Organ Sharing (UNOS) and the International Pancreas Transplant Registry (IPTR). Clin. Transplant. 45–56 (2008).
- 11. Mora M, Ricart MJ, Casamitjana R et al. Pancreas and kidney transplantation: long-term endocrine function. Clin. Transplant. 24(6), E236–240 (2010).
- 12. Demartines N, Schiesser M, Clavien PA. An evidence-based analysis of simultaneous pancreas-kidney and pancreas transplantation alone. *Am. J. Transplant.* 5(11), 2688–2697 (2005).

- 13. Cohen DJ, St Martin L, Christensen LL, Bloom RD, Sung RS. Kidney and pancreas transplantation in the United States, 1995–2004. Am. J. Transplant. 6(5 Pt 2), 1153–1169 (2006).
- 14. Kandaswamy R, Stock PG, Gustafson SK et al. OPTN/SRTR 2017 Annual Data Report: Pancreas. Am. J. Transplant. 19(Suppl. 2), 124–183 (2019).
- 15. Ramesh A, Chhabra P, Brayman KL. Pancreatic islet transplantation in Type 1 diabetes mellitus: an update on recent developments. Curr. Diabetes Rev. 9(4), 294–311 (2013).
- 16. Gaba RC, Garcia-Roca R, Oberholzer J. Pancreatic islet cell transplantation: an update for interventional radiologists. *J. Vasc. Intervent. Radiol. JVIR* 23(5), 583–594 (2012).
- 17. Shapiro AM. Strategies toward single-donor islets of Langerhans transplantation. Curr. Opin. Organ Transplant. 16(6), 627-631 (2011).
- 18. Ly TT, Nicholas JA, Retterath A, Lim EM, Davis EA, Jones TW. Effect of sensor-augmented insulin pump therapy and automated insulin suspension vs standard insulin pump therapy on hypoglycemia in patients with Type 1 diabetes: a randomized clinical trial. *Jama* 310(12), 1240–1247 (2013).
- 19. Thompson DM, Meloche M, Ao Z et al. Reduced progression of diabetic microvascular complications with islet cell transplantation compared with intensive medical therapy. *Transplantation* 91(3), 373–378 (2011).
- 20. Hering BJ, Clarke WR, Bridges ND *et al.* Phase 3 trial of transplantation of human islets in Type 1 diabetes complicated by severe hypoglycemia. *Diabetes Care* 39(7), 1230–1240 (2016).
- 21. Foster ED, Bridges ND, Feurer ID, Eggerman TL, Hunsicker LG, Alejandro R. Improved health-related quality of life in a Phase 3 islet transplantation trial in Type 1 diabetes complicated by severe hypoglycemia. *Diabetes Care* 41(5), 1001–1008 (2018).
- Ricordi C, Goldstein JS, Balamurugan AN et al. National Institutes of Health-Sponsored Clinical Islet Transplantation Consortium Phase 3 Trial: manufacture of a complex cellular product at eight processing facilities. *Diabetes* 65(11), 3418–3428 (2016).
- 23. Ryan EA, Paty BW, Senior PA et al. Five-year follow-up after clinical islet transplantation. Diabetes 54(7), 2060-2069 (2005).
- 24. Rossini AA. Autoimmune diabetes and the circle of tolerance. Diabetes 53(2), 267-275 (2004).
- Sharma V, Andersen D, Thompson M et al. Autoimmunity after islet-cell allotransplantation. New Engl. J. Med. 355(13), 1397–1399 (2006).
- 26. Wing K, Sakaguchi S. Regulatory T cells exert checks and balances on self tolerance and autoimmunity. *Nat. Immunol.* 11(1), 7–13 (2010)
- 27. Dufrane D, Gianello P. Macro- or microencapsulation of pig islets to cure Type 1 diabetes. World J. Gastroenterol. 18(47), 6885–6893 (2012).
- 28. Elliott RB, Escobar L, Tan PL, Muzina M, Zwain S, Buchanan C. Live encapsulated porcine islets from a Type 1 diabetic patient 9.5 yr after xenotransplantation. *Xenotransplantation* 14(2), 157–161 (2007).
- Carlsson PO, Palm F, Andersson A, Liss P. Markedly decreased oxygen tension in transplanted rat pancreatic islets irrespective of the implantation site. *Diabetes* 50(3), 489–495 (2001).
- Johansson H, Lukinius A, Moberg L et al. Tissue factor produced by the endocrine cells of the islets of Langerhans is associated with a negative outcome of clinical islet transplantation. Diabetes 54(6), 1755–1762 (2005).
- 31. Nilsson B, Ekdahl N, Korsgren O. Control of instant blood-mediated inflammatory reaction to improve islets of Langerhans engraftment. *Curr. Opin. Organ Transplant.* 16(6), 620–626 (2011).
- 32. Bennet W, Sundberg B, Groth CG et al. Incompatibility between human blood and isolated islets of Langerhans: a finding with implications for clinical intraportal islet transplantation? Diabetes 48(10), 1907–1914 (1999).
- Barshes NR, Wyllie S, Goss JA. Inflammation-mediated dysfunction and apoptosis in pancreatic islet transplantation: implications for intrahepatic grafts. *Journal of leukocyte biology* 77(5), 587–597 (2005).
- 34. Shapiro AM, Pokrywczynska M, Ricordi C. Clinical pancreatic islet transplantation. Nat. Rev. Endocrinol 13(5), 268-277 (2017).
- 35. Rother KI, Harlan DM. Challenges facing islet transplantation for the treatment of Type 1 diabetes mellitus. *J. Clin. Investig.* 114(7), 877–883 (2004).
- 36. Ricordi C. Islet transplantation: a brave new world. Diabetes 52(7), 1595-1603 (2003).
- Mooranian A, Negrulj R, Takechi R, Mamo J, Al-Sallami H, Al-Salami H. The biological effects of the hypolipidaemic drug probucol
  microcapsules fed daily for 4 weeks, to an insulin-resistant mouse model: potential hypoglycaemic and anti-inflammatory effects. *Drug Deliv. Transl. Res.* 8(3), 543–551 (2018).
- 38. Karaoz E, Tepekoy F, Yilmaz I, Subasi C, Kabatas S. Reduction of inflammation and enhancement of motility after pancreatic islet derived stem cell transplantation following spinal cord injury. *J. Korean Neurosurg. Soc.* 62(2), 153–165 (2019).
- 39. Wang J, Sun Z, Gou W et al. alpha-1 antitrypsin enhances islet engraftment by suppression of instant blood-mediated inflammatory reaction. Diabetes 66(4), 970–980 (2017).
- 40. Berman A, Wszola M, Gorski L et al. Islet autotransplantation in diabetic patients. Transplant. Proceed. 51(8), 2781–2786 (2019).

- 41. Al-Sofiani ME, Quartuccio M, Hall E, Kalyani RR. Glycemic outcomes of islet autotransplantation. *Curr. Diabetes Rep.* 18(11), 116 (2018).
- 42. Qi M. Transplantation of encapsulated pancreatic islets as a treatment for patients with Type 1 diabetes mellitus. Adv. Med. 2014, 429710 (2014).
- 43. Tooley JE, Waldron-Lynch F, Herold KC. New and future immunomodulatory therapy in Type 1 diabetes. *Trends Mol. Med.* 18(3), 173–181 (2012).
- 44. Desai T, Shea LD. Advances in islet encapsulation technologies. Nat. Rev. Drug Discov. 16(5), 338-350 (2017).
- Article for a better understanding of encapsulated islet cell therapy for humans.
- 45. Graham JG, Zhang X, Goodman A et al. PLG scaffold delivered antigen-specific regulatory T cells induce systemic tolerance in autoimmune diabetes. Tissue Eng. Part A 19(11–12), 1465–1475 (2013).
- Maehr R, Chen S, Snitow M et al. Generation of pluripotent stem cells from patients with Type 1 diabetes. Proc. Natl Acad. Sci. USA 106(37), 15768–15773 (2009).
- Scharp DW, Marchetti P. Encapsulated islets for diabetes therapy: history, current progress, and critical issues requiring solution. Adv. Drug Deliv. Rev. 67–68, 35–73 (2014).
- 48. Groboillot A, Boadi DK, Poncelet D, Neufeld RJ. Immobilization of cells for application in the food industry. *Crit. Rev. Biotechnol.* 14(2), 75–107 (1994).
- 49. Vojtisek V, Jirku V. Immobilized cells. Folia Microbiol. 28(4), 309-340 (1983).
- 50. Dolgin E. Diabetes: Encapsulating the problem. Nature 540(7632), S60-s62 (2016).
- 51. Elnashar MM, Kahil T. Biopolymeric formulations for biocatalysis and biomedical applications. Biomed. Res. Int. 2014, 418097 (2014).
- 52. Wilson JT, Chaikof EL. Challenges and emerging technologies in the immunoisolation of cells and tissues. *Adv. Drug Deliv. Rev.* 60(2), 124–145 (2008).
- Cao R, Avgoustiniatos E, Papas K, de Vos P, Lakey JRT. Mathematical predictions of oxygen availability in micro- and macro-encapsulated human and porcine pancreatic islets. J. Biomed. Mater. Res. BAppl. Biomater. 08(2), 343–352 (2019).
- 54. Ryan AJ, O'Neill HS, Duffy GP, O'Brien FJ. Advances in polymeric islet cell encapsulation technologies to limit the foreign body response and provide immunoisolation. *Curr. Opin. Pharmacol.* 36, 66–71 (2017).
- 55. Anderson JM, Rodriguez A, Chang DT. Foreign body reaction to biomaterials. Semin. Immunol. 20(2), 86-100 (2008).
- Danial EN, Elnashar MMM, Awad GEA. Immobilized inulinase on grafted alginate beads prepared by the one-step and the two-steps methods. Ind. Eng. Chem. Res. 49(7), 3120–3125 (2010).
- 57. Elnashar MMM, Danial EN, Awad GEA. Novel carrier of grafted alginate for covalent immobilization of inulinase. *Ind. Eng. Chem. Res.* 48(22), 9781–9785 (2009).
- 58. Elnashar MMM. Review article: immobilized molecules using biomaterials and nanobiotechnology. *Biomater. Nanobiotechnol.* 1(01), 61–76 (2010).
- Article about the different polymers/hydrogels used for immobilization (encapsulation) of cells.
- Patil JS, Kamalapur MV, Marapur SC, Kadam DV. Ionotropic gelation and polyelectrolyte complexation: the novel techniques to design hydrogel particulate sustained, modulated drug delivery system: a review. *Dig. J. Nanomater. Bios.* 5(1), 241–248 (2010).
- Elnashar MM, Yassin AM. Covalent immobilization of β-galactosidase on carrageenan coated chitosan. J. Appl. Polym. Sci. 114, 17–24 (2009).
- 61. Elnashar MM. Carriers for immobilization and reusability of enzymes. WO Patent WO2009155937A1 (2009).
- Elnashar MM, Yassin MA. Lactose hydrolysis by beta-galactosidase covalently immobilized to thermally stable biopolymers. Appl. Biochem. Biotechnol. 159(2), 426–437 (2009).
- 63. Elnashar M. Carrageenan treated with synthetic polymer as a carrier for biotechnological applications. Egypt (2009).
- 64. Awad GE, Amer H, El-Gammal EW, Helmy WA, Esawy MA, Elnashar MM. Production optimization of invertase by Lactobacillus brevis Mm-6 and its immobilization on alginate beads. *Carbohydr. Polym.* 93(2), 740–746 (2013).
- 65. Wahba MI, Soliman T. Whey protein isolate for the preparation of covalent immobilization beads. *Biocat. AgriBiotechnol* 14, 328–337 (2018).
- Elnashar MM HM, Awad GEA. Grafted carrageenan gel disks and beads with polyethylenimine and glutaraldehyde for covalent immobilization of penicillin G acylase. J. Colloid Sci. Biotechnol. 2, 1–7 (2013).
- 67. Elnashar MM, Mostafa H, Morsy NA, Awad GEA. Biocatalysts: isolation, identification and immobilization of thermally stable lipase onto novel supports. *Industrial & Engineering Chemistry Research* 52, 14760–14767 (2013).
- Elnashar M, Wahba M, Amin M, Eldiwany A. Application of Plackett- Burman screening design to the modeling of grafted alginatecarrageenan beads for the immobilization of penicillin G acylase. J. Appl. Polym. Sci. 131 (2014).
- 69. Elnashar MM, Hassan ME. Novel epoxy activated hydrogels for solving lactose intolerance. Biomed. Res. Int. 2014, 817985 (2014).

- Gawad R, Kattab, Strabel et al. Encapsulation method to protect unsaturated fatty acids from rumen biohydrogenation in vitro. J. Innov. Pharm. Biol. Sci. 2, 240–251 (2015).
- 71. Gawad R, Kattab, Strabel et al. Effect of different levels from linseed oil and linseed oil beads on rumen fermentation and microbial parameters using gas production system and rumen simulation technique. Asian J. Animal and Vet. Adv. 10 (2015).
- 72. Awad GEA, Abd El Aty AA, Shehata AN, Hassan ME, Elnashar MM. Covalent immobilization of microbial naringinase using novel thermally stable biopolymer for hydrolysis of naringin. 3 Biotech 6(1), 14 (2016).
- Esawy MA, Awad GEA, Wahab WAA et al. Immobilization of halophilic Aspergillus awamori EM66 exochitinase on grafted k-carrageenan-alginate beads. 3 Biotech 6(1), 29 (2016).
- Elnashar MM, Awad GE, Hassan ME, Mohy Eldin MS, Haroun BM, El-Diwany AI. Optimal immobilization of beta-galactosidase onto kappa-carrageenan gel beads using response surface methodology and its applications. Scientific World Journal 2014, 571682 (2014).
- 75. Elnashar MM. The art of immobilization using biopolymers, biomaterials and nanobiotechnology. *J. Biomater. Nanobiotechnol.* 1, 61–76 (2010).
- Calafiore R, Basta G. Clinical application of microencapsulated islets: actual prospectives on progress and challenges. Adv. Drug Deliv. Rev. 67–68, 84–92 (2014).
- 77. Wang T, Adcock J, Kuhtreiber W et al. Successful allotransplantation of encapsulated islets in pancreatectomized canines for diabetic management without the use of immunosuppression. *Transplantation* 85(3), 331–337 (2008).
- 78. Tuch BE, Keogh GW, Williams LJ *et al.* Safety and viability of microencapsulated human islets transplanted into diabetic humans. *Diabetes care* 32(10), 1887–1889 (2009).
- Elnashar M, Yassin M, Abdel Moneim A, Abdel Bary E. Surprising performance of alginate beads for the release of low-molecular-weight drugs. J. Appl. Polym. Sci. 116, 3021–3026 (2010).
- Elnashar MM, Abdallah MS, Hamed RR. Covalent immobilization of penicillin G acylase onto grafted alginate beads. J. Colloid Sci. Biotechnol. 1, 7 (2012).
- 81. Prochorov AV, Tretjak SI, Goranov VA, Glinnik AA, Goltsev MV. Treatment of insulin dependent diabetes mellitus with intravascular transplantation of pancreatic islet cells without immunosuppressive therapy. *Adv. Med. Sci.* 53(2), 240–244 (2008).
- 82. Ludwig B, Reichel A, Steffen A et al. Transplantation of human islets without immunosuppression. Proc. Natl Acad. Sci. USA 110(47), 19054–19058 (2013).
- •• This is the first time for the oxygenated alginate-encapsulation strategy to be applied to allogeneic human islet transplantation in
- 83. Jacobs-Tulleneers-Thevissen D, Chintinne M, Ling Z et al. Sustained function of alginate-encapsulated human islet cell implants in the peritoneal cavity of mice leading to a pilot study in a Type 1 diabetic patient. Diabetologia 56(7), 1605–1614 (2013).
- 84. de Vos P, Faas MM, Strand B, Calafiore R. Alginate-based microcapsules for immunoisolation of pancreatic islets. *Biomaterials* 27(32), 5603–5617 (2006).
- 85. Elnashar MM, Abdel Moneim A, Abdel Bary EM. Surprising performance of alginate beads for the release of low-molecular-weight drugs. J. Appl. Polym. Sci. 116, 3021 (2010).
- Shoichet MS, Li RH, White ML, Winn SR. Stability of hydrogels used in cell encapsulation: an in vitro comparison of alginate and agarose. Biotechnol. Bioeng. 50(4), 374

  –381 (1996).
- 87. Mateo C, Abian O, Fernandez-Lafuente R, Guisan JM. Increase in conformational stability of enzymes immobilized on epoxy-activated supports by favoring additional multipoint covalent attachment\*. Enzyme Microbial Technol. 26(7), 509–515 (2000).
- 88. Harrington S, Williams J, Rawal S, Ramachandran K, Stehno-Bittel L. Hyaluronic acid/collagen hydrogel as an alternative to alginate for long-term immunoprotected islet transplantation. *Tissue Eng. A* 23(19–20), 1088–1099 (2017).
- 89. Dufrane D, Goebbels RM, Saliez A, Guiot Y, Gianello P. Six-month survival of microencapsulated pig islets and alginate biocompatibility in primates: proof of concept. *Transplantation* 81(9), 1345–1353 (2006).
- 90. Gotfredsen CF, Stewart MG, O'Shea GM, Vose JR, Horn T, Moody AJ. The fate of transplanted encapsulated islets in spontaneously diabetic BB/Wor rats. *Diabetes Res. (Edinburgh, Scotland)* 15(4), 157–163 (1990).
- 91. Langlois G, Dusseault J, Bilodeau S, Tam SK, Magassouba D, Halle JP. Direct effect of alginate purification on the survival of islets immobilized in alginate-based microcapsules. *Acta Biomater.* 5(9), 3433–3440 (2009).
- 92. Hillberg AL, Kathirgamanathan K, Lam JB, Law LY, Garkavenko O, Elliott RB. Improving alginate-poly-L-ornithine-alginate capsule biocompatibility through genipin crosslinking. *J. Biomed. Mater. Res. B Appl. Biomater.* 101(2), 258–268 (2013).
- 93. Campanha-Rodrigues AL, Grazioli G, Oliveira TC, Campos-Lisboa AC, Mares-Guia TR, Sogayar MC. Therapeutic potential of laminin-biodritin microcapsules for Type 1 diabetes mellitus. *Cell Transplant*. 24(2), 247–261 (2015).
- 94. Prestwich GD. Hyaluronic acid-based clinical biomaterials derived for cell and molecule delivery in regenerative medicine. *J. Control. Release* 155(2), 193–199 (2011).
- 95. Burdick JA, Prestwich GD. Hyaluronic acid hydrogels for biomedical applications. *Adv. Mater. (Deerfield Beach, Fla.)* 23(12), H41–56 (2011).

- 96. Prestwich GD. Engineering a clinically-useful matrix for cell therapy. Organogenesis 4(1), 42-47 (2008).
- 97. Vanderhooft JL, Alcoutlabi M, Magda JJ, Prestwich GD. Rheological properties of cross-linked hyaluronan-gelatin hydrogels for tissue engineering. Macromol. Biosci. 9(1), 20-28 (2009).
- 98. Cheng K, Blusztajn A, Shen D et al. Functional performance of human cardiosphere-derived cells delivered in an in situ polymerizable hyaluronan-gelatin hydrogel. Biomaterials 33(21), 5317-5324 (2012).
- 99. Liu Y, Shu XZ, Prestwich GD. Osteochondral defect repair with autologous bone marrow-derived mesenchymal stem cells in an injectable, in situ, cross-linked synthetic extracellular matrix. Tissue Eng. 12(12), 3405-3416 (2006).
- 100. Mohan N, Gupta V, Sridharan B, Sutherland A, Detamore MS. The potential of encapsulating "raw materials" in 3D osteochondral gradient scaffolds. Biotechnol. Bioeng 111(4), 829-841 (2014).
- 101. Murali R, Ponrasu T, Cheirmadurai K, Thanikaivelan P, Biomimetic hybrid porous scaffolds immobilized with platelet derived growth factor-BB promote cellularization and vascularization in tissue engineering. J. Biomed. Mater. Res. A 104(2), 388-396 (2016).
- 102. Shapiro H, Kolodziejczyk AA, Halstuch D, Elinav E. Bile acids in glucose metabolism in health and disease. J. Exp. Med. 215(2), 383-396 (2018).
- 103. Hofmann AF, Borgstroem B. The intraluminal phase of fat digestion in man: the lipid content of the micellar and oil phases of intestinal content obtained during fat digestion and absorption. J. Clin. Investig. 43, 247-257 (1964).
- 104. Hofmann AF. The continuing importance of bile acids in liver and intestinal disease. Arch. Int. Med. 159(22), 2647-2658 (1999).
- 105. Feng Y, Siu K, Wang N et al. Bear bile: dilemma of traditional medicinal use and animal protection. J. Ethnobiol. Ethnobed. 5, 2 (2009).
- 106. Williamson KD, Chapman RW. Primary sclerosing cholangitis. Digest. Dis. (Basel, Switzerland) 32(4), 438-445 (2014).
- 107. Vang S, Longley K, Steer CJ, Low WC. The Unexpected uses of urso- and tauroursodeoxycholic acid in the treatment of non-liver diseases. Global Adv. Health Med. 3(3), 58-69 (2014).
- 108. Robles L, Storrs R, Lamb M, Alexander M, Lakey JR. Current status of islet encapsulation. Cell Transplant. 23(11), 1321–1348 (2014).
- 109. Street CNK, G S. Stem cells as a source of islets for transplantation in Type 1 diabetes: scientific and ethical considerations. Can. J. Diabetes 27, 262-270 (2003).
- 110. Pathak V, Pathak NM, O'Neill CL, Guduric-Fuchs J, Medina RJ. Therapies for Type 1 diabetes: current scenario and future perspectives. Clin. Med. Insights Endocrinol. Diabetes 12, 1179551419844521 (2019).
- 111. Kerby A, Jones ES, Jones PM, King AJ. Co-transplantation of islets with mesenchymal stem cells in microcapsules demonstrates graft outcome can be improved in an isolated-graft model of islet transplantation in mice. Cytotherapy 15(2), 192-200 (2013).
- 112. Alipio Z, Liao W, Roemer EJ et al. Reversal of hyperglycemia in diabetic mouse models using induced-pluripotent stem (iPS)-derived pancreatic beta-like cells. Proc. Natl Acad. Sci. USA 107(30), 13426-13431 (2010).
- 113. Kroon E, Martinson LA, Kadoya K et al. Pancreatic endoderm derived from human embryonic stem cells generates glucose-responsive insulin-secreting cells in vivo. Nat. Biotechnol. 26(4), 443-452 (2008).
- 114. Matveyenko AV, Georgia S, Bhushan A, Butler PC. Inconsistent formation and nonfunction of insulin-positive cells from pancreatic endoderm derived from human embryonic stem cells in athymic nude rats. Am. J. Physiol. Endocrinol. Metab. 299(5), E713-720 (2010).

future science group fsg 10.2144/fsoa-2020-0105 Future Sci. OA (2020) FSO660