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Keywords: crocetin, monoolein, polarized light microscopy, small angle xray scattering, diffusion, tape-stripping

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Response to Reviewers: Dr. Xinyuan Zhu, Ph.D Editor Colloids and Surfaces B: Biointerfaces

Dear Dr. Xinyuan Zhu, thank you for the refereeing of the manuscript COLSUB-D-18-00627, "Monoolein liquid crystalline phases for topical delivery of crocetin" by F. Carducci, P. Mariani, N. Huang, F. Simelière, R. Cortesi, G. Romeo, C. Puglia and myself. We emended our manuscript accordingly to the comments raised by the Reviewers. The corrections are evidenced in yellow. Detailed list of the changes made: Reviewer #1

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Dear. Prof. John L. Brash,

Enclosed please find the manuscript "Monoolein liquid crystalline phases for topical delivery of crocetin" by

Elisabetta Esposito*, Federica Carducci, Paolo Mariani, Nicolas Huang, Fanny Simelière, Rita Cortesi*, Giuseppe Romeo and Carmelo Puglia

The paper is submitted for possible publication on "Colloids and Surfaces B".

We believe that the reported results give an interesting contribution to the field of colloids characterization and technology.

The mesophases resulting in monoolein/water systems are fascinating but currently underexploited for biological application. We have thoroughly characterized them by small angle x-ray scattering, polarized light microscopy and rheological analyses.

In addition, crocetin is a natural antioxidant molecule with several biological applications but very difficult to be solubilized in well tolerated biocompatible vehicles.

In vivo tape-stripping demonstrated the possibility to apply crocetin on the skin and to differently control its uptake using different monoolein concentrations.

For these reasons we think that publication of our findings can give valuable inputs to Colloids and Surfaces B readers.

Statistical summary

total number of words (excluding figure captions) 5995; Tables: 3, Figures: 5.

The manuscript is original, unpublished, it is not under consideration for publication elsewhere, and all authors have read and approved the text and consent to its publication.

I thank you in advance for consideration. Best regards

Dr. Elisabetta Esposito

Ferrara, 11 april 2018

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| 4 | Simelière ^c , Rita Cortesi ^{a*} , | Giuseppe Romeo ^d and Carmelo Puglia ^d | | | | | |
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| 24 | Statistical summary | | | | | | |
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42

Keywords: crocetin, monoolein, polarized light microscopy, small angle x-ray scattering,
 diffusion, tape-stripping.

45 **Abbreviations :** Crocetin (CRT) ; small angle x-ray scattering (SAXS).

47 **1. Introduction**

Amphiphilic polar lipids, such as monoglycerides, can form various crystalline phases in 48 the presence of different amounts of water. These lipids self-associate, depending on the 49 temperature and aqueous content, forming reversed micellar (L2), lamellar (L α), or 50 bicontinuous cubic phases (C) in which the hydrocarbon chains assume a liquid-like 51 conformation [1-2]. Particularly, glycerol monooleate (monoolein), а nontoxic. 52 biodegradable and biocompatible material commonly used as emulsifying agent and food 53 additive, is one of the monoglycerides most widely employed to form liquid crystalline 54 formulations [3-5]. Different studies have attributed to monoolein a penetration enhancer 55 activity when applied on skin, probably due to a temporary and reversible disruption of the 56 lamellar structure of the lipid bilayer in the stratum corneum caused by an increase in 57 intercellular lipid fluidity [6-8]. In the presence of tiny amounts of water (5-10%, w/w), 58 monoolein forms reversed micelles or lamellar phases, while when more water is added 59 (~15-40%) a cubic phase region dominates. This highly viscous isotropic phase is defined 60 bicontinuous, being constituted of a curved three-dimensional bilayer, separating two 61 congruent water channel networks [9, 10]. 62

Many authors have focused their attention on the relevance of the lipid crystalline phases for drug delivery [11-13]. Indeed, the presence of a lipid and an aqueous domain confers special properties to the crystalline phases, such as the ability to solubilize hydrophilic, hydrophobic, and amphiphilic substances [14-16]. In addition, liquid crystalline phases protect drugs from degradation, control drug release and possess hydrating power [17-20].

⁶⁸ Crocetin (CRT) is an active molecule originally discovered in dried stigma of <u>Crocus</u>
⁶⁹ Sativus (Saffron), recently taken in consideration in biomedical research because of its
⁷⁰ pharmacological activities, such as antitumoral, antioxidant, antihypertensive,
⁷¹ antiatherosclerotic and antidepressant [21-25].

In Crocus Sativus the chromoplast zeaxanthine cleavage dioxygenase first generates CRT 72 dialdehyde, then converted into CRT by an aldehyde oxydo-reductase and transformed to 73 the glucosylate derivative crocin by a glucosyl transferase [26,27]. Interestingly, recent 74 studies have indicated the potential of CRT against skin damage induced by UV-A. Indeed, 75 CRT reduces the oxidative stress by decreasing reactive oxygen species production and 76 cell apoptosis [28]. Nonetheless, the antioxidant power of CRT is unfortunately associated 77 to a low stability, especially in the presence of heat, oxygen, light and acids [29]. In the 78 present study, monoolein-water systems have been designed and investigated as vehicles 79 for CRT with the aim to find cutaneous systems suitable to treat skin pathologies and to 80 81 protect skin against UV-induced skin damage. The different monoolein mesophases have been characterized by small angle x-ray scattering (SAXS), polarized light microscopy and 82 rheological measurements. Tape stripping experiments enabled to shed light on the CRT 83 84 distribution on stratum corneum after application of different monoolein-water systems on the skin. 85

87 **2. Materials and methods**

88

89 2.1. Materials

90

91 Crocin and glucosidase enzyme were purchased from Merck KGaA (Darmstadt, 92 Germany). 2,3-Dihydroxypropyl oleate, Glyceryl monooleate, RYLO MG 19 (monoolein) 93 was a gift from Danisco Cultor (Grindsted, Denmark). Solvents and other chemicals were 94 purchased from Merck KGaA.

95

96 **2.2 Preparation of crocetin**

97 Crocetin, (2E,4E,6E,8E,10E,12E,14E)-2,6,11,15-Tetramethylhexadeca-2,4,6,8,10,12,14-

⁹⁸ heptaenedioic acid, CRT), was obtained in our laboratory by alkaline hydrolysis of crocin,

99 the protocol is reported in the supplementary materials section.

100

101 **2.3. Production of monoolein-water samples**

Monoolein based formulations were prepared by adding different amounts of water (ranging from 5 and 25% w/w) to molten monoolein at 42°C [30,31]. When a uniform mixture was formed under stirring, the containers were sealed, to avoid water evaporation, and placed in an oven at 42°C for 72 h. Afterwards the samples have been stirred by hand until uniform aspect. In the case of CRT containing monoolein-water systems, CRT (0.02 % w/v) has been added to the samples before placing in the oven. Sample compositions and acronyms are reported in Table 1.

Table 1: Composition of the studied monoolein based formulations

110

| Formulations | Monoolein (% w/w) | Water (% w/w) | Crocetin (% w/v)* |
|--------------|-------------------|---------------|-------------------|
| M-75 | 75 | 25 | - |
| M-80 | 80 | 20 | - |
| M-85 | 85 | 15 | - |
| M-90 | 90 | 10 | - |
| M-95 | 95 | 5 | - |
| M-75-CRT | 75 | 25 | 0.02 |
| M-80-CRT | 80 | 20 | 0.02 |
| M-85-CRT | 85 | 15 | 0.02 |
| M-90-CRT | 90 | 10 | 0.02 |
| M-95-CRT | 95 | 5 | 0.02 |

111 *with respect to the volume of the formulations

112

113

114 **2.4 X-ray characterization**

Small angle X-ray scattering (SAXS) experiments were performed at the Elettra synchrotron radiation facility (Basovizza, Trieste) at the AustroSAXS beamline. Samples were held on a flat watertight holder, to allow fixed composition studies and to avoid mechanical stress that could interfere or increase anisotropy. Experiments were performed at 37 °C on different monoolein formulations in the presence and in the absence of CRT. The investigated q-range (q = $4\pi \sin \theta/\lambda$, where 20 is the scattering angle and $\lambda = 1.54$ Å the X ray wavelength) was 1-4 nm⁻¹. The observed Bragg peaks were indexed considering
the different symmetries commonly observed in lipid polymorphism [32].

123

124 **2.5 Polarized light microscopy**

Monoolein-water samples were examined through a polarized light microscope (Ortholux POL-MK, Carl Zeiss, Oberkochen, Germany) to verify texture and anisotropy of the liquid crystalline phases. The prepared sample was deposited on a glass slide using a spatula, to avoid any possible mechanical stress that could force the alignment of the molecules inside the sample. Once the sample was loaded, the coverslip was sealed using silicone grease, to ensure the maintenance of the monoolein hydration. For every type of formulation, three different samples were observed.

132

133 **2.6 Rheological measurements**

Rheological measurements were performed with an AR-G2 controlled-stress rotational rheometer (TA Instruments, USA). The geometry used was an aluminium cone-plate (40 mm diameter, 1° cone angle, 28 μ m truncation gap). Flow curves were obtained by a flow sweep protocol: after a 2-min conditioning time, shear rate was increased from 0.1 to 2000 s⁻¹ for a total duration of 180 s. The temperature was maintained at 37 °C and controlled with a Peltier plate. Measurements were performed in triplicate at least for each sample, to ensure reproducibility.

141

142 2.7 Spreadability test

¹⁴³ The spreading capacity of selected formulations, namely M-90 and M-95, was evaluated

¹⁴⁴ as follows. One hundred mg of preparation was placed on a Petri dish (3 cm diameter) and

¹⁴⁵ pressed by another glass dish on which a 500-g mass was positioned. Taken the time by

which the formulation fills the entire dish, the following equation was used to calculate the
 spreadability (S) value.

148 $S = m \times I / I$

(1)

¹⁴⁹ where m is the weight (g) tied on the upper plate, I is the diameter (cm) of the glass plate,

and t is the time (s) taken for the gel to fill the entire diameter. The spreadability test was

¹⁵¹ performed thrice and the mean values ± standard deviations were calculated.

152

153 **2.8 CRT content**

The content of CRT in monoolein-water systems was determined by dissolving an aliquot of sample in dimethyl sulfoxide (1:10, w/w) under magnetic stirring (ARE-6 heating magnetic stirrer, VELP Scientifica, Usmate, Italy) for 1 hour in amber glass vials to avoid CRT photodegradation.

Samples were then filtered by nylon filters with 0.45 µm pore diameter (Whatman[™], Germany) and finally diluted with methanol (1:10, v/v). The filtrate was then analyzed by high performance liquid chromatography (HPLC). Determinations were performed using a quaternary pump (Agilent Technologies 1200 series, USA) an UV-detector operating at 423 nm, and a 7125 Rheodyne injection valve with a 50 µl loop. Samples were loaded on a stainless-steel C-18 reverse-phase column (15×0.46 cm) packed with 5 µm particles (Grace® - Alltima, Alltech, USA).

Elution was performed with a mobile phase containing methanol:water (80:20, v/v); at a flow rate of 0.8 ml/min, retention time was 3.8 min. The method was validated for linearity ($R^2 = 0.994$), repeatability (relative standard deviation 0.02%, n=6 injections) and limit of quantification (0.03 µg/ml).

169 CRT content was expressed as percentage of the total amount added to monoolein-water170 for system production.

171

172 **2.9 Prediction of long-term stability**

173 The stability of CRT was assessed in stored glass containers at 40°C for 3 months, 70-

174 <mark>75% RH.</mark>

Chemical stability was evaluated, determining CRT content by HPLC analyses as above
 reported. Shelf life values were calculated as below reported [33].

Log (CRT residual content, %) was plotted against time and the slopes (m) were calculated by linear regression.

The slopes (m) were then substituted into the following equation for the determination of k values:

181
$$k = m \times 2.303$$
 (2)

Shelf life values (the time for 10% loss, t_{90}) and half-life (the time for 50% loss, $t_{1/2}$) were then calculated by the following equations:

(4)

184
$$t_{90} = 0.105/k$$
 (3)

185
$$t_{1/2} = 0.693/k$$

186

187 **2.10 In vitro diffusion experiments**

CRT diffusion experiments were performed by Franz-type diffusion cells supplied by
 Vetrotecnica (Padova, Italy) associated to nylon membranes (Merck Millipore, 0.45 μm
 pore size).

The exposed membrane area was 0.78 cm² area (1 cm diameter orifice). The receptor compartment contained 5 ml of a mixture of phosphate buffer 60 mM pH 7.4 and ethanol (50:50, v/v). This solution was stirred with the help of a magnetic bar at 500 rpm and thermostated at $32 \pm 1^{\circ}$ C during all the experiments [34,35].

Approximately 1 g of M-75-CRT, M-90-CRT or M-95-CRT was placed on the membrane in the donor compartment and the latter was sealed to avoid evaporation. At predetermined

time intervals comprised between 0 and 24 h, samples (0.15 ml) of receptor phase solution 197 198 were withdrawn and the CRT concentration in the receptor phase was measured using HPLC. Each removed sample was replaced with an equal volume of fresh receptor phase. 199 The CRT concentrations were determined six times in independent experiments and the 200 mean values ± standard deviations were calculated. The mean values were then plotted 201 as a function of time. The flux coefficients were computed from the linear portion of the 202 accumulation curve calculating the curve slopes and dividing them by the CRT 203 concentration in the monoolein based formulation (expressed in mg/ml). 204

205

206 **2.11 Tape stripping**

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208 2.11.1 Volunteers recruitment

Ten volunteers of both sexes in the age range 25–55 years were recruited after medical screening, including the filling of a health questionnaire followed by physical examination of the application sites. Informed consent was obtained from all individual participants included in the study. The participants did not suffer from any ailment and were not on any medication at the time of the study. They were rested for 15 min prior to the experiments and room conditions were set at 22 ± 2 °C and 40–50% relative humidity.

215

216 2.11.2 Experimental protocol

The tape stripping protocol was approved by the Ethics Committee of the University of Ferrara, Italy (study number: 170986) and conducted in accordance with the Code of Ethics of the World Medical Association (Helsinki Declaration 1964) and its later amendments for experiments involving humans.

For each subject, ten sites on the ventral surface of forearms were defined using a rectangular template (2 cm^2) and demarcated with permanent ink. One of the ten sites of

each forearm was used as control, three sites were treated with 80 mg of M-75-CRT, three 223 sites with 80 mg of M-90-CRT and the remaining three with 80 mg of M-95-CRT. The 224 preparations were spread uniformly by means of a solid glass rod, thereafter the sites 225 were occluded for 1 h using rectangular plasters especially designed for skin occlusion. 226 Afterwards the residual formulations were removed by gently wiping with cotton balls 227 (different for each pretreated site). Ten individual 2 cm² squares of adhesive tape (Scotch 228 Book Tape 845, 3M) were utilized to sequentially tape-strip the stratum corneum on the 229 application sites. Particularly stratum corneum in each pretreated site was removed at 0.5 230 $(t_{0.5})$, 3 (t_3) and 6 (t_6) h after formulation removal. 231

Each adhesive square, before and after skin tape stripping, was weighed on a semimicrobalance (sensitivity 1mg, Sartorius model ME415S, Goettingen, Germany) to quantify the weight of removed *stratum corneum*. After each stripping, the tapes were put in the same vial containing 2 ml of the HPLC mobile phase methanol:water (80:20 v/v) and subjected to vortical stirring over 30 s. The extracted CRT was then quantified by HPLC. The recovery of CRT was validated by spiking tape-stripped samples of untreated *stratum corneum* with 2 ml of a mobile phase containing CRT 5 mg/ml [36].

239

240 2.11.3 Statistical Analysis

Statistical differences of *in vivo* data were determined using repeated-measures analysis of variance (ANOVA) followed by the Bonferroni-Dunn post hoc pairwise comparison procedure. The employed software was Prism 5.0, Graph Pad Software Inc. (La Jolla, CA -USA). A probability of less than 0.05 is considered significant in this study.

245 **3. Results**

246

247 **3.1. Synthesis and characterization of crocetin**

248 CRT is difficult to obtain in high yield and with a good degree of purity, thus it is hardly 249 available commercially. For this reason, the compound was obtained in our laboratory, 250 exploiting some experimental techniques described in supplementary materials section. 251 CRT was obtained by alkaline hydrolysis, its ¹H-NMR spectrum showed a doublet (d, δ =

252 7.21, *J* = 9.6 Hz) and a multiplet (m, δ = 6.90 – 6.44) related to hydrogens of the polyenic 253 chain and two singlets related to hydrogens of the methyl groups placed symmetrically at 254 the 2- and 15-positions (s, δ = 1.98) and at the 6- and 11-positions (s, δ = 1.92) of the 255 chain. The ¹H-NMR spectrum signals relative to the CRT sample coincided with those 256 estimated by the simulation program ACD/C + H NMR Predictors (version 11.01) and with 257 literature data [37].

258

3.2 Production of monoolein-water systems

A simple protocol was adopted to produce monoolein-water samples, notably the addition 260 of small percentage of water to melted monoolein followed by equilibration at 42°C 261 262 resulted in transparent systems with different consistency, depending on the amount of added water (Fig S1A, supplementary material). To assess CRT solubility, different 263 amounts of the drug (0.2-1 mg/ml) have been added to the monoolein-water systems 264 before equilibration at 42°C. The transparency of the systems enabled to detect the 265 presence of yellow crystals in the case of samples containing CRT>0.2 mg/ml, while a 266 homogeneous yellow colouring characterized samples with CRT 0.2 mg/ml. For all the 267 tested monoolein-water systems the CRT content, evaluated after disaggregation and 268 HPLC analyses, was 98 % \pm 0.2 with respect to the weight of CRT added to the 269 monoolein-water mixture. 270

271

3.4 X-ray scattering analysis

273 X-ray diffraction experiments were performed to investigate the structural properties of the 274 monoolein formulations. A few results are shown in Fig. 1, while details about phase 275 symmetry and unit cell values are reported in Table 2.

The X-ray diffraction scattering profiles for M-95 and M-95-CRT samples show a large 276 band centered at about 1.8 nm-1. In good agreement with the monoolein-water phase 277 diagrams [32,38], the large band confirms the presence of a micellar phase in both 278 systems. A narrow peak is indeed overlapped on this band in the case of the more 279 hydrated M-90 and M-90-CRT samples, indicating the formation of the lamellar phase. For 280 higher water concentrations, several Bragg peaks are observed; both in the absence and 281 in the presence of CRT, the spacing of the reflections has been indexed considering the 282 space group Q230. The characteristic profile then indicates the formation of the cubic la3d 283 phase in the more hydrated conditions investigated. A schematic representation of the 284 285 different lyotropic phases observed in the present systems is reported in Fig S1B (supplementary material). It should be mentioned that the la3d cubic phase is bicontinuous 286 and exhibits two 3-D networks of connected aqueous rods, co-planarly joined 3 by 3 [39]. 287 Notably, no differences in the X-ray diffraction profiles are detected in the presence of 288 CRT, disregarding the case of M-85-CRT sample. In this latest case, Bragg peaks appear 289 doubled, probably due to a low homogeneity of the sample, whose concentration 290 corresponds to the lamellar-to-la3d cubic phase boundary. Thus, CRT does not modify the 291 structural organization of monoolein. Slight changes have been however detected in the 292 unit cell parameters, which appear to be systematically reduced in the presence of CRT 293 (Table 2), suggesting a small dehydration of the lipid layer. 294

295



Figure 1. SAXS profiles observed on monoolein, both in the absence (A) and in the presence (B) of CRT
 0.02% w/v, measured at 37°C. The black arrow indicates the direction of the increasing concentration of
 water. Curves were separated using offset for clarity.

311

312 3.5 Polarized light microscopy characterization

Polarized light microscopy enables to easily characterize the sample quality and to screen 313 314 the lyotropic liquid crystalline phases in lipid-water systems. Indeed, lamellar and hexagonal phases evidence birefringence textures, as real crystals, while lack of 315 birefringence indicates that the phase is cubic or liquid [40]. Fig 2 shows polarized light 316 microscopy images of M-85-CRT and M-90-CRT samples. In the case of M-85-CRT (Fig. 317 1A), birefringent anisotropic textures can be appreciated, probably because of the 318 presence of phase coexistence. Instead, in the case of M-90-CRT (Fig. 1B) as well as M-319 95-CRT (not shown), flower-like structures typical of a lamellar phase organization can be 320 observed [41]. Similar images were taken in the case of samples produced in the absence 321

of CRT. In the case of the other more hydrated monoolein-water systems (from 85% to 75%), no textures have been detected, confirming the presence of a cubic phase.



324

Figure 2. Polarized light microscopy images of M-85-CRT (A), and M-90-CRT (B). Observations were made
 using a magnification 10x.

327 **3.6 Rheological analysis**

Rheology gives valuable information about the lipid crystalline phases [42-44]. A comprehensive rheological characterization can be very useful for their practical applications and for the development of suitable formulations.

Results of rheological measurements performed on plain monoolein and on monooleinwater systems, in the absence or in the presence of CRT, are shown in Fig 3, while viscosity values measured at 10 s⁻¹ are reported in Table 2.



Figure 3. Rheological flow curves of the indicated monoolein based formulations produced in the absence (A) or in the presence (B) of CRT. Measurements were performed at 37°C. The geometry used was an aluminium cone-plate. Flow curves were obtained by increasing the shear rate from 0.01 s-1 to 5000 s-1 with 5 points per decade, each point was maintained for a duration of 180 s to perform measurements in the permanent regime. Data are the means of 3 analyses on different batches of the same type of formulations.

Table 2. Macroscopic aspect, viscosity, phase symmetry and unit cell of monoolein
 based formulations measured at 37°C.

362

| Formulations | Macroscopic aspect | Viscosity* (Pa.s) ±5% | Phase symmetry ^a | Unit cell ^ª (Å) ± 0.5 |
|--------------|-----------------------|--------------------------|-----------------------------|-------------------------------------|
| M-75 | gel | 1435 | cubic la3d (Q230) | 120.7 |
| M-80 | gel | 3514 | cubic Ia3d (Q230) | 107.4 |
| M-85 | gel | 3819 | cubic la3d (Q230) | 99.9 |
| M-90 | viscous | 4.6 | lamellar/micellar | 37.6 |
| M-95 | liquid | 0.3 | micellar | 34 (broad) |
| M-75-CRT | gel | 1785 | cubic la3d (Q230) | 118.9 |
| M-80-CRT | gel | 3224 | cubic la3d (Q230) | 106.7 |
| M-85-CRT | gel | 3927 | cubic la3d (Q230) | 99.7 |
| M-90-CRT | viscous | 5.2 | lamellar/micellar | 37.4 |
| M-95-CRT | liquid | 0.3 | micellar | 34 (broad) |

363 -shear rate 10 ^{s-1}; a: as determined by X-ray scattering. Monoolein based formulations acronyms
 364 are explained in Table 1.
 365

Viscosity curves show that M-90, M-85, M-80 and M-75 were strongly shear-thinning.

Indeed, for these samples an appreciable decrease of the viscosity was detectable increasing the shear rate. Generally, an increase of water led to an increase of viscosity, particularly the increase was dramatic passing from M-95 to M-90, or from M-90 to M-85. Viscosity curves of M-85 and M-80 are alike and could be not distinguishable, indeed the structure for these two samples displayed the same flow behavior. The viscosity values of M-75 are slightly smaller than M-85 and M-80. M-95 and plain monoolein, employed as control, showed a slight shear-thinning behavior at low shear rate (below $\approx 1 \text{ s}^{-1}$) and a

Newtonian behavior at higher shear rates. The presence of CRT did not influence the viscosity, as indicated by superimposable profiles of Fig. 3A, 3B and viscosity data reported in Table 2. Upward and downward viscosity flow sweeps have been performed on M-80, M-90, M-95 and M-100 samples (supplementary material, Fig. S2). The samples were all weakly thixotropic and slightly more thixotropic in the case of M-90 (Fig. S2B) at low shear rates.

380

381 3.7 Spreadability of monoolein-water systems

Spreadability of monoolein-water systems was studied to select the systems suitable for 382 administration on skin. Indeed, this parameter is essential for cutaneous administration 383 since it influences extrudability from the package, uniform application and drug therapeutic 384 efficacy [45,46]. Among the systems, only those characterized by micellar or lamellar 385 mesophases, i.e. M-95 and M-90, were easily spreadable (spreadability ratio M-95/M-90 386 1.6:1) (Table S1, supplementary materials). Monoolein-water systems based on cubic 387 388 phases were not easily spreadable, apart from M-75, characterized by a viscosity lower than M-85 and M-80. The presence of CRT did not modify the system spreadability. 389 Noteworthily, further studies have been focused on M-95-CRT, M-90-CRT and on M-75-390 391 CRT, selected to compare the micellar, lamellar and cubic phase performances as delivery system for CRT. 392

393

394 **3.8 CRT stability**

To assess shelf life stability, CRT content of monoolein-water systems was determined as a function of time and expressed as percentage of the total amount used for the preparation (Fig. S3, supplementary materials). After 3 months CRT content followed the order M-75-CRT>M-90-CRT> M-95-CRT (CRT respectively 85, 71 and 54 % with respect

to the drug content detected after sample preparation). Table 3 reports shelf life (t_{90}) and

400 half-life $(t_{1/2})$ values calculated by equations (2) and (3).

401

402 **Table 3.** Shelf life data and CRT fluxes from the indicated monoolein based formulations

403

| Formulations | mª | к ^ь | t ₉₀ ^b (days) | t _{1/2} ^b (days) | flux ^c (cm/h*10 ³) |
|--------------|---------|-----------------------|-------------------------------------|--------------------------------------|---|
| M-75-CRT | -0.0007 | 0.0017 | 58.67 | 389.32 | 2.37 |
| M-90-CRT | -0.0016 | 0.0036 | 28.37 | 187.29 | 3.67 |
| M-95-CRT | -0.0027 | 0.0063 | 16.51 | 109.1 | 4.37 |

404 a: slope of the line of log (CRT residual content %) kinetic, calculated as the mean of 3 independent 405 determinations, s.d. $\leq 2\%$; b: K, t₉₀ and t_{1/2} were calculated following equations. 1, 2 and 3 406 respectively; c: calculated by Franz cell experiment, considering the slope of the line of CRT diffusion 407 and its concentration in the monoolein formulations. Monoolein based formulations acronyms are 408 explained in Table 1.

409

410 It was found that $t_{1/2}$ value of CRT exceeded 1 year in the case of M-75-CRT, 6 months in

the case of M-90-CRT and 3 months in the case of M-95-CRT. All data were statistically

412 significant (p<0.0001).

413 All monoolein/water systems maintained their physical appearance with time, without

414 phase separation phenomena also after six months from production.

415

416 **3.9 In vitro CRT diffusion**

The diffusion of CRT included in M-75-CRT, M-90-CRT and M-95-CRT was compared by Franz cell experiments. Notably the receptor phase was constituted of a phosphate buffer/ethanol 50:50 (v/v) mixture to allow the establishment of the sink conditions and to

sustain permeant solubilization [34]. Fig 4 shows the diffusion kinetics corresponding to

the linear part of the profile (from 0 to 8 h), while the flux values are reported in Table 3.

422 The diffusion of CRT was more controlled in the case of M-75-CRT with respect to the

other forms. Indeed, M-75-CRT flux value was almost half than M-95-CRT and 2/3 with
respect to M/90-CRT.

425



426

Figure 4. In vitro diffusion profiles of CUR from M-75-CRT (squares), M-90-CRT (circles) and M-95-CRT
 (crosses). Experiments were performed by Franz cell associated to nylon membranes. Data represent the
 mean of six independent experiments ± S.D.

430

431 **3.10 Tape-stripping evaluation**

A monocentric observational experiment was conducted by tape stripping for quantifying CRT presence in the *stratum corneum* after cutaneous administration of M-75-CRT, M-90-CRT and M-95-CRT [36,47]. Formulations have been applied on the forearms following the scheme depicted in data in supplementary material Fig. S4. Portions of *stratum corneum* have been stripped at $t_{0.5}$, t_3 and t_6 . Tape stripping results are summarized in Fig.5. A

general depletion in the amount of CRT in the stratum corneum was observed by time. 437 Notably, in the case of M-75-CRT, the CRT amount at t_{0.5} was 3.5 and 2-fold higher with 438 respect to M-90-CRT and M-95-CRT respectively (Fig 5A). Fig 5B shows a comparative 439 evaluation of the CRT level present in stratum corneum at t_3 and t_6 with respect to $t_{0.5}$. A 440 depletion of CRT more pronounced in the case of M-75-CRT was observed, followed by 441 M-90-CRT and M-95-CRT (Fig 5B). The CRT amounts detected in the stratum corneum at 442 $t_{0.5}$ and t_3 were significantly different. The extraction efficiency of CRT was 97.8 ± 0.4 % 443 (n=3).444

445

446 **4. Discussion and conclusions**

The powerful activity of CRT makes this molecule interesting both for pharmaceutical, as 447 well as for cosmeceutical applications. We succeeded to obtain this molecule by alkaline 448 hydrolysis of crocin. To find vehicles able to solubilize and deliver CRT through the skin, 449 monoolein-water systems were investigated. In these captivating systems, monoolein 450 disposes in various forms as a function of water content, resulting in different crystalline 451 mesophases. SAXS characterization of monoolein water-systems enabled to identify 452 micellar, lamellar or Q 230 phases. Namely these latest mesophases were found for 15-25 453 % water content. These results confirm previously findings of other authors and agree well 454 with polarized light microscopy observations and rheological measurements [30,43,44]. 455 Indeed, birefringence of M-90, M-90-CRT, M-95 and M-95-CRT samples allowed to 456 observe flower-like structures typical of lamellar phases, while samples with 20-25 % water 457 content did not transmit the light, being characterized by isotropic cubic phases. 458

Viscosity profiles are very intriguing, indeed monoolein-water samples showed a noncontinuous behavior under dilution (Fig. 3A and 3B). Namely, an almost Newtonian behavior was observed for 0-5% water content, while samples containing 10-25 % of water

were strongly shear-thinning, suggesting an important structure rearrangement under 462 shear. Such viscosity behavior can be directly related to the structure of the various 463 monoolein phases, having a different degree of entanglement. Indeed, the disordered 464 isotropic micellar phase (M-95), occurring in very dehydrated conditions, was very fluid 465 (viscosity<1 Pas), while an increase of water concentration led to lamellar phase formation 466 (M-90), characterized by a 1-D ordered structure and a dramatically higher viscosity (1-100 467 Pas). A second remarkable change in viscosity (100-100000 Pas) was observed in the 468 case of cubic la3d phase, showing a 3-D ordered structure, thus particularly viscous. 469 Notably, viscosity values of samples in the Ia3d cubic phase (M-85, M-80 and M-75) 470 471 slightly decreased, as a function of water content. It can be suggested that under hydration the cubic phase softens because of the changes in the monoolein structural parameters. 472 Indeed, with the same degree of entanglement, the lipid bilayer curvature, the area-per-473 474 molecule at the lipid/water interface and the hydrocarbon chain packing should play a key role on the mechanical properties of the phase [32]. 475

476 Regarding chemical stability, monoolein-water systems preparation did not induce
477 degradation of CRT, as indicated by HPLC analyses of systems after sample
478 disaggregation.

Shelf life and Franz cell studies evidenced that the system characterized by cubic phases better controlled stability and diffusion of CRT with respect to micellar and lamellar based systems. These differences could be attributed both to the viscosity and to the crystallographic structure of the systems, indeed, the viscosity of liquid crystalline phases can affect the diffusion kinetics of the solubilized active molecules. Notably, the bicontinuous cubic phases are more rigid than the lamellar ones, described as plastic fluids undergoing yielding [43,44].

Tape stripping experiments enabled to shed light on the performance of monoolein 486 mesophases as cutaneous delivery systems for CRT. At the first-time interval (t_{0.5}), the 487 higher CRT amount found in stratum corneum in the case of M-75-CRT could account for 488 its higher viscosity that could initially prolong its permanence on skin with respect to M-90-489 CRT and M-95-CRT. It should be considered that monoolein can alter the skin barrier 490 properties due to interactions with the intercellular lipids in the stratum corneum. These 491 alterations of lipid packing lead to hydration of stratum corneum and disorganization of the 492 lipid bilayers [7,18]. At t₃ and t₆, it is noteworthy that the CRT depletion was not ascribable 493 to high monoolein/water ratio, indeed in the case of micellar (M-95-CRT) or lamellar 494 495 phases (M-90-CRT), CRT depletion was less pronounced with respect to the cubic phase system (M-75-CRT). The more pronounced depletion of CRT found in the case of M-75-496 CRT could be justified by the hypothesis of a penetration enhancement effect, due to an 497 498 interaction between cubic phase system and stratum corneum lipids. This hypothesis is corroborated by some studies indicating a higher skin permeability exerted by cubic 499 phases with respect to lamellar ones [48] while others suggested a similarity between the 500 cubic phase structure and the structure of the stratum corneum [49]. Thus, it could be 501 suggested that monoolein organized in cubic mesophases could mix with stratum corneum 502 lipids and induce an intercellular lipid disorder, finally promoting skin uptake. 503

On the other hand, it can be asserted that the lower viscosity of M-90-CRT and M-95-CRT initially promotes CRT penetration through *stratum corneum* (t_{0.5}), afterwards the drug is slowly subtracted, suggesting the formation of a monoolein depot within the *stratum corneum* lipids. At this regard, it should be considered that CRT should long remain in the upper skin strata to exert its skin protection against UV damage, thus a prolong permanence is desirable, while a deeper CRT penetration should be avoided. Moreover, it is noteworthy that rheological properties (shear-thinning behaviour) and spreadability of

- 511 the lamellar phase make it more appropriate for cutaneous application with respect to the
- 512 cubic one.
- Eventually this study has demonstrated the suitability of monoolein-water systems as cutaneous vehicles for CRT, nonetheless further in vivo studies are needed (i) to verify the antioxidant activity and skin protection of the different CRT containing systems, (ii) to point out the mechanism of monoolein mesophases and CRT distribution in the different skin layers.

518

519 **Conflict of interest**

- 520 The authors declare that there is no conflict of interests.
- 521

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525 **References**

- 526 [1] J.M. Seddon, R.H. Templer, Polymorphism of lipid-water systems, in: R. Lipowsky, E.
- Sackmann (Eds.), Handbook of Biological Physics, Volume 1, Elsevier Science Publishers
 B.V., N.Y., United States, 1995.
- [2] R. Mezzenga, Physics of self-assembly of lyotropic liquid crystals, in: Nissim Garti,
 Ponisseril Somasundaran, Raffaele Mezzenga (Eds.), Self-assembled supramolecular
 architectures: lyotropic liquid crystals, John Wiley & Sons Inc. Hoboken, New Jersey 2012
 edited by, pp. 19-35.
- 533 [3] A. Ganem-Quintanar, D. Quintanar-Guerrero, P. Buri, Monoolein: a review of the 534 pharmaceutical applications, Drug. Dev. Ind. Pharm., 26 (2000) 809-820.
- [4] P. Mariani, F. Rustichelli, L. Saturni, L, Cordone, Stabilization of the monoolein Pn3m
 cubic structure on trehalose glasses, Eur. Biophys. J., 28 (1999) 294-301.
- 537 [5] V. Chandrashekhar, W.W. Kulkarni, I.-S. Guillermo, S. Engelskirchen, S. Ahuallia,
 538 Monoolein: a magic lipid?, Phys. Chem. Chem. Phys., 13 (2011) 3004-3021.
- [6] L.B. Lopes, J.H. Collett, M.V. Bentley, Topical delivery of cyclosporin A: an in vitro study
- using monoolein as a penetration enhancer, Eur. J. Pharm. Biopharm., 60 (2005) 25-30.
- 541 [7] L.B. Lopes, F.F. Speretta, M. Bentley, Enhancement of skin penetration of vitamin K
- using monoolein-based liquid crystalline systems, Eur. J. Pharm. Sci., 32 (2007) 209-215.
- [8] S. Milak, A. Zimmer, Glycerol monooleate liquid crystalline phases used in drug delivery
 systems, Int. J. Pharm., 478 (2015) 569-587.
- [9] K. Larsson, Cubic lipid–water phases: structures and biomembrane aspects, J. Phys.
 Chem., 93 (1989) 7304-7314.
- [10] K. Larsson, Aqueous dispersions of cubic lipid–water phases, Curr. Opin. Colloid
 Interface Sci., 5 (2000) 64-69.

- [11] M.G. Lara, M.V.L.B. Bentley, J.H. Collett, In vitro drug release mechanism and drug
 loading studies of cubic phase gels, Int. J. Pharm., 293 (2005) 241-250.
- [12] A.R. Ahmed, A. Dashevsky, R. Bodmeier, Drug release from and sterilization of in situ
 cubic phase forming monoglyceride drug delivery systems, Eur. J. Pharm. Biopharm., 75
 (2010) 375-380.
- [13] J.C. Shah, Y. Sadhale, D.M. Chilukuri, Cubic phase gels as drug delivery systems,
 Adv. Drug Deliv. Rev., 47 (2001) 229-250.
- [14] A. Zabara, R. Mezzenga, Controlling molecular transport and sustained drug release
 in lipid-based liquid crystalline mesophases, J. Control. Rel., 188 (2014) 31-43.
- [15] S.B. Rizwan, B.J. Boyd, T. Rades, S. Hook, Bicontinuous cubic liquid crystals as
 sustained delivery systems for peptides and proteins, Expert Opin. Drug Deliv., 7 (2010)
 1133-1144.
- [16] C. Guo, J. Wang, F. Cao, R.J. Lee, G. Zhai, Lyotropic liquid crystal systems in drug
 delivery, Drug Discov. Today, 15 (2010) 1032-1040.
- [17] H. Evenbratt, A. Strom, Phase behavior, rheology, and release from liquid crystalline
 phases containing combinations of glycerol monooleate, glyceryl monooleyl ether,
 propylene glycol, and water, RSC Adv., 7 (2017) 32966-32973.
- [18] L.B. Lopes, F.F. Speretta, M.V.Bentley, Enhancement of skin penetration of vitamin K
 using monoolein-based liquid crystalline systems, Eur. J. Pharm. Sci., 32 (2007) 209-215.
- [19] I. Amar-Yuli, D. Libster, A. Aserin, N. Garti, Solubilization of food bioactives within
 lyotropic liquid crystalline mesophases, Curr. Opin. Colloid Interface Sci., 14 (2009) 21–32.
 [20] F. Caboi, G.S. Amico, P. Pitzalis, M. Monduzzi, T. Nylander, K. Larsson, Addition of
 hydrophilic and lipophilic compounds of biological relevance to the monoolein/water
 system. Phase behaviour, Chem. Phys. Lipids, 109 (2001) 47-62.

573 [21] S.Z. Bathaie, R. Hoshyar, H. Miri, M. Sadeghizadeh, Anticancer effects of crocetin in
574 both human adenocarcinoma gastric cancer cells and rat model of gastric cancer.
575 Biochem. Cell Biol., 91 (2013) 397-403.

576 [22] F. Yoshino, A. Yoshida, N. Umigai, K. Kubo, MC. Lee, Crocetin reduces the oxidative
577 stress induced reactive oxygen species in the stroke-prone spontaneously hypertensive
578 rats (SHRSPs) brain, J. Clin. Biochem. Nutr., 49 (2011) 182-187.

- [23] E. Christodoulou, N.P. Kadoglou, N. Kostomitsopoulos, G. Valsami, Saffron: A natural
 product with potential pharmaceutical applications, J. Pharm. Pharmacol., 67 (2015) 16341649.
- [24] M. Giaccio, Crocetin from saffron: an active component of an ancient spice, Crit. Rev.
 Food Sci. Nutr., 44 (2004) 155-172.
- [25] L. Song, C. Kang, Y. Sun, W. Huang, W. Liu, Z. Qian, Crocetin Inhibits
 Lipopolysaccharide- Induced Inflammatory Response in Human Umbilical Vein Endothelial
 Cells, Cell Physiol. Biochem., 40 (2016) 443-452.
- [26] F. Bouvier, C. Suire, J. Mutterer, B. Camara, Oxidative remodeling of chromoplast
 carotenoids: identification of the carotenoid dioxygenase CsCCD and CsZCD genes
 involved in Crocus secondary metabolite biogenesis, Plant Cell., 15 (2003) 47-62.
- [27] O. Ahrazem, A. Rubio-Moraga, J. Berman, T. Capell, P. Christou, C. Zhu, L. GómezGómez, The carotenoid cleavage dioxygenase CCD2 catalysing the synthesis of crocetin
 in spring crocuses and saffron is a plastidial enzyme, New Phytol., 209 (2016) 650-663.
- [28] T. Ohba, M. Ishisaka, S. Tsujii, K. Tsuruma, M. Shimazawa, K. Kubo, N. Umigai, T.
 Iwawaki, H. Hara, Crocetin protects ultraviolet A-induced oxidative stress and cell death in
 skin in vitro and in vivo, Eur. J. Pharmacol., 789 (2016) 244-253.
- [29] M. Tsimidou, E. Tsatsaroni, Stability of saffron pigments in aqueous extracts, J. Food
 Sci., 58 (1993) 1073-1075.

- [30] P. D'Antona, W.O. Parker, M.C. Zanirato, E. Esposito, C. Nastruzzi, Rheological and
 NMR characterization of monoglyceride-based formulations, J. Biomed. Mater. Res., 52
 (2000) 40-52.
- [31] E. Esposito. V. Carotta, A. Scabbia, L. Trombelli, P. D'Antona, E. Menegatti, C.
 Nastruzzi, Comparative analysis of tetracycline-containing dental gels: Poloxamer- and
 monoglycerides-based formulations. Int. J. Pharm., 142 (1996) 9-23.
- [32] S. Mazzoni, L.R.S. Barbosa, S.S. Funari, R. Itri, P. Mariani, Cytochrome-c Affects the
 Monoolein Polymorphism: Consequences for Stability and Loading Efficiency of Drug
 Delivery Systems, Langmuir, 32 (2016) 873-881.
- [33] A. Garg, D. Aggarwal, S. Garg, A.K. Singla, Spreading of semisolid formulations. An
 update, Pharm. Technol., 26 (2002) 84-105.
- [34] W.J. Pugh, Kinetics of product stability. In: Aulton ME (Ed). Aultons's Pharmaceutics.
- The design and manufacture of the medicines, 3rd ed. London: Churchil Livingstone Elsevier (2007) pp:99-107
- [35] M. Siewert, J. Dressman, C.K. Brown, V.P. Shah, FIP/AAPS guidelines to
 dissolution/in vitro release testing of novel/special dosage forms, AAPS PharmSciTech, 4
 (2003) E7.
- [36] L.M. Andrade, C. de Fátima Reis, L. Maione-Silva, J.L.V. Anjos, A. Alonso, R.I.C.
 Serpa, R.N. Marreto, E. M. Lima, S. F. Taveira, Impact of lipid dynamic behavior on
 physical stability, in vitro release and skin permeation of genistein-loaded lipid
 nanoparticles, Eur. J. Pharm. Biopharm., s 88 (2014) 40–47
- [37] E. Esposito, M. Sguizzato, M. Drechsler, P. Mariani, F. Carducci, C. Nastruzzi, R,
 Cortesi, Progesterone lipid nanoparticles: Scaling up and in vivo human study, Eur. J.
 Pharm. Biopharm., 119 (2017) 437-446.
- [38] M.-R. Van Calsteren, M. C. Bissonnette, F. Cormier, C. Dufresne, T. Ichi, J.C.Y.
 LeBlanc, D. Perreault, I. Roewer, Spectroscopic characterization of crocetin derivatives
 - 28

from Crocus sativus and Gardenia jasminoides, J. Agric. Food Chem., 45 (1997) 10551061.

[39] J. Briggs, H. Chung, M. Caffrey, The temperature-composition phase diagram and
mesophase structure characterization of the monoolein/water system, J Phys II, EDP Sci.,
6 (1996) 723-751.

[40] P. Mariani, V. Luzzati, H. Delacroix, Cubic phases of lipid-containing systems.
Structure analysis and biological implications, J. Mol. Biol., 204 (1988) 165-189.

[41] C. C. Muller-Goymann, Physicochemical characterization of colloidal drug delivery
systems such as reverse micelles, vesicles, liquid crystals and nanoparticles for topical
administration, Eur. J. Pharm. Biopharm., 58 (2004) 343-356.

[42] Y. Iwashita, H. Tanaka, Self-organization in phase separation of a lyotropic liquid
 crystal into cellular, network and droplet morphologies, Nat. Mater., 5 (2006) 147-152.

[43] L. Sagalowicz, M. Michel, M. Adrian, P. Frossard, M. Rouvet, H.J. Watzke, A.
Yaghmur, L. de Campo, O. Glatter, M.E. Leser. Crystallography of dispersed liquid
crystalline phases studied by cryo-transmission electron microscopy, J. Microsc., 221
(2006) 110-121.

[44] E. Esposito, M. Sguizzato, C. Bories, C. Nastruzzi, R. Cortesi, Production and
characterization of clotrimazole liposphere gel for candidiasis treatment, Polymers, 10
(2018) 160-175.

[45] E. Esposito, M. Drechsler, N. Huang, G. Pavoni, R. Cortesi, D. Santonocito, C. Puglia,
Ethosomes and organogels for cutaneous administration of crocin, Biomed. Microdevices,
18 (2016) 1-12.

[46] R. Mezzenga, C. Meyer, C. Servais, A.I. Romoscanu, L. Sagalowicz, R.C. Hayward,
Shear rheology of lyotropic liquid crystals: a case study, Langmuir, 21 (2005) 3322-3333.

[47] G. Bonacucina, G.F. Palmieri, D.Q.M. Craig, Rheological and dielectric
characterization of monoolein/water mesophases in the presence of a peptide drug, J.
Pharm. Sci., 94 (2005) 2452-2462.

[48] D.G. Lim, W.-W. Jeong, N.A. Kim, J.Y. Lim, S.-H. Lee, W.S. Shim, N.-G. Kang, S.H.
Jeong. Effect of the glyceryl monooleate-based lyotropic phases on skin permeation using

- in vitro diffusion and skin imaging. Asian J. Pharm. Sci., 9 (2014) 324-329.
- [49] N. Lars, A.-A. Ashraf, Stratum corneum keratin structure, function, and formation: the
 cubic rod-packing and membrane templating model, J. Invest. Dermatol., 4 (2004) 715732.

658 **FIGURE LEGENDS**

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Figure 1. SAXS profiles observed on monoolein, both in the absence (A) and in the presence (B) of CRT 0.02% w/v, measured at 37°C. The black arrow indicates the direction of the increasing concentration of water. Curves were separated using offset for clarity.

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Figure 2. Polarized light microscopy images of M-85-CRT (A), and M-90-CRT (B).
 Observations were made using a magnification 10x.

667

Figure 3. Rheological flow curves of the indicated monoolein based formulations produced in the absence (A) or in the presence (B) of CRT. Measurements were performed at 37°C. The geometry used was an aluminium cone-plate. Flow curves were obtained by increasing the shear rate from 0.01 s⁻¹ to 5000 s⁻¹ with 5 points per decade, each point was maintained for a duration of 180 s to perform measurements in the permanent regime. Data are the means of 3 analyses on different batches of the same type of formulations.

Figure 4. In vitro diffusion profiles of CUR from M-75-CRT (squares), M-90-CRT (circles)
 and M-95-CRT (crosses). Experiments were performed by Franz cell associated to nylon
 membranes. Data represent the mean of six independent experiments ± S.D.

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Figure 5. Tape stripping evaluation. A: CRT amount in the *stratum corneum* after M-90CRT, M-95-CRT and M-75-CRT application, removal and tape stripping. Tape stripping
was performed t_{0.5} (white), t₃ (light) and t₆ (dark) from formulation removal. B: comparative 31

evaluation of the CRT level present at t_3 (light bars) and t_6 (dark bars) in *stratum corneum*. The reported levels represent the percentage of CRT with respect to that present in *stratum corneum* at $t_{0.5}$. Data represent the mean for ten subjects ± S.D., p<0.001.

1 Table 1: Composition of the studied monoolein based formulations

2

| Formulations | Monoolein (% w/w) | Water (% w/w) | Crocetin (% w/v)* |
|--------------|-------------------|---------------|-------------------|
| M-75 | 75 | 25 | - |
| M-80 | 80 | 20 | - |
| M-85 | 85 | 15 | - |
| M-90 | 90 | 10 | - |
| M-95 | 95 | 5 | - |
| M-75-CRT | 75 | 25 | 0.02 |
| M-80-CRT | 80 | 20 | 0.02 |
| M-85-CRT | 85 | 15 | 0.02 |
| M-90-CRT | 90 | 10 | 0.02 |
| M-95-CRT | 95 | 5 | 0.02 |

3 *with respect to the volume of the formulations

Table 2. Macroscopic aspect, viscosity, phase symmetry and unit cell of monoolein based formulations measured at 37°C.

| Formulations | Macroscopic aspect | Viscosity* (Pa.s) ±5% | Phase symmetry ^a | Unit cell ^a (Å) ± 0.5 |
|--------------|-----------------------|--------------------------|-----------------------------|-------------------------------------|
| M-75 | gel | 1435 | cubic Ia3d (Q230) | 120.7 |
| M-80 | gel | 3514 | cubic Ia3d (Q230) | 107.4 |
| M-85 | gel | 3819 | cubic Ia3d (Q230) | 99.9 |
| M-90 | viscous | 4.6 | lamellar/micellar | 37.6 |
| M-95 | liquid | 0.3 | micellar | 34 (broad) |
| M-75-CRT | gel | 1785 | cubic la3d (Q230) | 118.9 |
| M-80-CRT | gel | 3224 | cubic la3d (Q230) | 106.7 |
| M-85-CRT | gel | 3927 | cubic Ia3d (Q230) | 99.7 |
| M-90-CRT | viscous | 5.2 | lamellar/micellar | 37.4 |
| M-95-CRT | liquid | 0.3 | micellar | 34 (broad) |

*shear rate 10 ^{s-1}; a: as determined by X-ray scattering. Monoolein based formulations acronyms are explained in Table 1.

Table 3. Shelf life data and CRT fluxes from the indicated monoolein based 10 formulations 11

12

| Formulations | m ^a | K ^b | t ₉₀ ^b (days) | t _{1/2} ^b (days) | flux ^c (cm/h*10 ³) |
|--------------|----------------|----------------|-------------------------------------|--------------------------------------|---|
| M-75-CRT | -0.0007 | 0.0017 | 58.67 | 389.32 | 2.37 |
| M-90-CRT | -0.0016 | 0.0036 | 28.37 | 187.29 | 3.67 |
| M-95-CRT | -0.0027 | 0.0063 | 16.51 | 109.1 | 4.37 |

13 a: slope of the line of log (CRT residual content %) kinetic, calculated as the mean of 3 independent

14 determinations, s.d. $\leq 2\%$; b: K, t₉₀ and t_{1/2} were calculated following equations. 2, 3 and 4 15 respectively; c: calculated by Franz cell experiment, considering the slope of the line of CRT diffusion 16 and its concentration in the monoolein formulations. Monoolein based formulations acronyms are

17 explained in Table 1.



Figure 1



Figure 2







Figure 4



Figure 5

Supplementary Material Click here to download Supplementary Material: Supplemental materials coll Surf B-R.docx **Graphical Abstract**



Highlights

The natural antioxidant crocetin is very difficult to be solubilized in biocompatible vehicles
Crocetin can be solubilized in monoolein/water systems leading to different mesophases
Micellar, lamellar and Q230 phases have been found as a function of added water
Viscosity and spreadability of monoolein systems allow crocetin application on skin
Monoolein/water systems differently control crocetin diffusion through skin