- 1 Expanding the potential of chiral chromatography
- ² for high-throughput screening of large compound
- ³ libraries by means of sub–2µm Whelk-O 1

⁴ stationary phase in supercritical fluid conditions

5 Luca Sciascera¹, Omar Ismail¹, Alessia Ciogli^{1,*}, Dorina Kotoni^{1,2}, Alberto Cavazzini³, Lorenzo

6 Botta⁴, Ted Szczerba⁵, Jelena Kocergin⁵, Claudio Villani¹ and Francesco Gasparrini^{1,*}

- ¹ Dipartimento di Chimica e Tecnologie del Farmaco, "Sapienza" Università di Roma, P.le Aldo
 Moro 5, 00185 Roma, Italy;
- 9 ² current address: Chemical and Analytical Development, Novartis Pharma AG, CH–4002, Basel,
- 10 Switzerland;
- ³ Department of Chemistry and Pharmaceutical Sciences, University of Ferrara, Via L. Borsari
- 12 46, 44121Ferrara, Italy;
- ⁴ Department of Pharmacy, University of Naples Federico II, Via D. Montesano 49, 80131,
- 14 Naples, Italy;
- ⁵ Regis Technologies, Inc., 8210 Austin Avenue, Morton Grove, IL 60053, USA.

16

17 Keywords

sub-2 µm Whelk-O 1; ultra fast chiral separations; eUHPSFC; enantioselective high throughput
screening.

20	* Corresponding authors.
----	--------------------------

- 21 1) Tel.: +39 06 49912776; fax: +39 06 49912780.
- *E-mail address*: francesco.gasparrini@uniroma1.it (F. Gasparrini).
- 23 2) Tel.: +39 06 49912799; fax: +39 06 49912780.
- *E-mail address*: alessia.ciogli@uniroma1.it (A. Ciogli).

39 Abstract

With the aim of exploring the potential of ultrafast chiral chromatography for high-throughput
analysis, the new sub-2 micron Whelk-O 1 chiral stationary phase (CSP) has been employed in
supercritical fluid conditions to screen 129 racemates, mainly of pharmaceutical interest.

By using a 5-cm long column (0.46 cm internal diameter), a single co-solvent (MeOH) and a 7min gradient elution, 85% of acidic and neutral analytes considered in this work have been successfully resolved, with resolution (Rs) larger than 2 in more than 65% of cases. Moreover, almost a half of basic samples that, for their own characteristics, are known to be difficult to separate on Whelk–O 1 CSP, have shown Rs greater than 0.3. The screening of the entire library could be accomplished in less than 24 h (single run) with 63% of positive score.

For well-resolved enantiomers (Rs roughly included between 1 and 3), we show that method transfer from gradient to isocratic conditions is straightforward. In many cases, isocratic ultrafast separations (with analysis time smaller than 60 sec) have been achieved by simply employing, as isocratic mobile phase, the eluent composition at which the second enantiomer was eluted in gradient mode.

54 By considering the extension and variety of the library in terms of chemico-physical and 55 structural properties of compounds and numerousness, we believe that this work demonstrates 56 the real potential of the technique for high-throughput enantioselective screening.

57

58

59

60 **1. Introduction**

61 The production of new chiral molecules to be evaluated as potential drug candidates has 62 experienced, over the last 15 years, an impressive growth in medicinal and pharmaceutical 63 chemistry. To assess the true potency of each single enantiomer, highly-efficient and fast 64 separation methods are needed. Among them, direct enantioselective chromatography, thanks to 65 the broad availability of Chiral Stationary Phases (CSPs) and modes of operation, predated the 66 field becoming *de facto* an essential tool in the area of drug discovery [1-6]. Depending on the 67 stage in chiral drug discovery, goals and needs are different. At the initial stages of the process, 68 indeed, the possibility of a quickly screening of large libraries of chiral molecules which is 69 fundamental [7]. High throughput methods need enantioselective systems able to separate the 70 largest number of structurally and chemically different molecules with minimum changes of 71 experimental conditions. On the other hand, once a racemic candidate has been selected for 72 successive steps of drug development, the optimization of the separation (possibly under 73 isocratic preparative conditions) is strongly facilitated by the possibility of transferring all or part 74 of the information gathered in the screening phase with great practical and economic advantages.

75 Enantioselective Supercritical Fluid Chromatography (eSFC) is nowadays a preferred solution 76 for the development of fast-screening methods [8–15]. Supercritical or subcritical CO₂-based 77 mobile phases are indeed characterized by lower viscosity and faster mass transfer properties 78 than typical liquid chromatography eluents, which allows for the reduction of analysis time 79 without losing efficiency. In addition, as they permit significant reduction of organic solvent 80 consumption (estimable up to 20-30% in volume), SFC methods are considered to be greener 81 than both normal- and reversed-phase chromatography, which is significant especially from the 82 industrial point of view. The fast eSFC screening of chiral molecules has been most of times

83 realized by using polysaccharide-based CSPs with gradient elution [16-17]. Analysis time in the 84 order of 15-20 min or longer were considered very satisfactory in chiral separations until 5-10 85 years ago, when a barrier to the development of very fast separations is essentially represented 86 by the 5 µm particle size of the packing bed [16]. Reduced analysis time can be obtained by 87 using the chiral stationary phases developed on 3 µm particle size [18-21]. However, the recent 88 development of CSPs in sub-2 µm format has however opened new frontiers in the field of 89 enantioselective ultra-high performance SFC (eUHPSFC) [22]. As a matter of fact, however, this 90 transition has so far effectively affected only brush-type CSPs, in part because of practical 91 difficulties to adapt the surface modification chemistry of classical chiral stationary phases to 92 smaller particles (a very common problem encountered during the preparation of sub-2µm CSPs 93 is their tendency to aggregate during synthesis, which provokes the attainment of broad particle 94 size distribution unsuitable for efficient packing) and, in part, due to the scarcity of fundamental 95 studies in mass-transfer mechanisms in CSPs [22, 23]. On the other hand, brush type CSPs and, 96 in particular, Whelk-O 1 type have been prepared in sub-2 μ m format with excellent results [24-97 26]. Indeed, phenomena such as particle aggregation and clogging or the non-uniformity/excess 98 of selector coating are not frequently encountered during the preparation of these phases, which 99 on the other hand exhibit fast adsorption/desorption kinetics possibly due to the monomeric 100 nature of selectors.

In this work, we report about the use of a Whelk-O 1 CSP, prepared by employing 1.7 micron high-surface totally porous spherical particles, to screen a very large library made of 129 racemic compounds in eUHPSFC. Very promising results have been obtained under fast-gradient conditions (7 min total analysis time) for compounds with significantly different chemicophysical properties, including acidic, neutral and even basic ones, which are not ideal molecules

106 to be separated on these stationary phases [27]. In the light of the numerousness of the compound 107 library and its variety, this study represents, to the best of our knowledge, the first example 108 demonstrating the real feasibility of high-throughput screening of enantiomers in eUHPSFC. We 109 also show that, when the separation of enantiomers is reasonably satisfactory (1 < Rs < 3), a 110 simple criterion to achieve very fast and efficient chiral separations under isocratic conditions is 111 that of operating the column at a mobile phase composition close to that in which the second 112 enantiomer has been eluted under gradient mode. In cases of smaller Rs, this proof-of-concept finding has been in any case particularly important, as it has provided reliable initial conditions 113 114 for optimization of the isocratic separation of enantiomers.

115 **2. Experimental**

116 **2.1 Materials and chemicals**

117 All reagents and solvents were purchased from Sigma Aldrich (Milano, Italy) and used without 118 further purification. Grade 5.5 carbon dioxide was from Gruppo SAPIO (Milano, Italy). HPLC 119 gradient grade methanol was further filtered on 0.2 µm Omnipore filters (Merck Millipore, 120 Darmstadt, Germany), prior to use in the UHPSFC system. Syncronis silica 1.7 µm (pore size 120 Å, particle size 1.7 μ m and specific surface area 320 m² g⁻¹) was a gift from Thermo 121 122 Scientific (Waltham, MA, USA). The (S,S) Whelk-O 1 selector and the HPLC (S,S) Whelk-O 1 123 analytical column (particle size 5µm, 150 mm x 4.6 mm ID) were donated by Regis Technologies Inc.[®] (Morton Grove, IL, USA). Chiral samples were available from previous 124 125 studies or were provided by Regis Technologies Inc.[®] (Morton Grove, IL, USA). The complete 126 list of analyzed samples is reported in Table 1. Empty stainless steel column was from IsoBar 127 Systems by Idex (Wertheim-Mondfeld, Germany).

128 **2.2 Instrumentations**

129 UHPLC Instrumentation. The UHPLC chromatographic system used was an UltiMate 3000 130 RS system from Thermo Fisher Dionex (Sunnyvale, California), consisting of a dual gradient RS 131 pump (pressure up to 1034 bar under reversed phase conditions, up to 800 bar under normal 132 phase conditions; flow rates up to 8.0 mL/min), an in-line split loop Well Plate Sampler, a 133 thermostatted RS Column Compartment (temperature range 5-110°C) and DAD detector with a 134 2.5 µL flow cell. The DAD was set at a filter time constant of 0.002 s, a data collection rate of 135 100 Hz and a response time of 0.025 s. Viper capillaries and fittings were used, with the two 136 capillary Viper tubes (350 mm x 0.13 mm I.D.) producing an extra-column volume of 9.3 µL. 137 Data acquisition and processing was performed with Chromeleon 6.8 software from Thermo 138 Fisher. Detection of all tested analytes was carried out at two different wavelengths (214 nm and 139 254 nm). The injection volume ranged between $1 - 2 \mu L$ on the eUHPLC 1.7 μm columns and 140 between 5 - 10 µL on the eHPLC columns. The UHPLC system was characterized prior to the 141 analysis, yielding a total extra-column volume of 19 μ L (variance, $\mu_{2,extra,f0.5}$ = 12.2 μ L²), 142 obtained by injecting toluene and using a zero dead volume connector.

UHPSFC Instrumentation. A Waters Acquity UPC² (Ultra Performance Convergence 143 144 Chromatography) was used to perform SFC analyses. The system was equipped with a binary 145 solvent delivery pump compatible with mobile phase flow rates up to 4 mL/min and maximum 146 system pressure of 414 bar. A 250 µL mixing chamber is present in the delivery system. The 147 system also comprised an autosampler with a 10 µL loop, a column oven compatible with 148 temperatures up to 90 °C, an UV detector equipped with an 8 µL flow-cell and a backpressure 149 regulator (BPR). The injector/column inlet and column/detector connection tubes were 600 mm 150 long and had an I.D. of 0.175 mm. The extra-column and dwell volumes of this instrument were

- estimated to be 60 µL and 440 µL respectively [28]. Data acquisition and control of the
 UHPSFC system was performed with the Empower 3.
- 153

154 2.3 Chiral Whelk–O 1 stationary phase and columns

The (*S*,*S*) Whelk-O 1, 1.7- μ m chiral stationary phase (CSP) was prepared as described in reference [29, 30]. A 50 x 4.6 mm L x I.D. column was used in this study. The unoptimized packing procedure consists of acetone slurry (composition 10% w/v), 900 bar packing pressure and hexane as flushing solvent. End frits of 0.5 μ m were used. To comparative purpose, a Regis (*S*,*S*) Whelk-O 1, 5 μ m analytical column (150 mm x 4.6 mm ID) was employed.

160

161 **2.4 Methodology**

162 The van Deemter equation (H = A + B/ ϕ + C* ϕ) was used to fit the experimental data, allowing 163 to compare the efficiency of the new (S,S) Whelk-O 1, 1.7 µm column with the commercially 164 available (S,S) Whelk-O 1 columns packed with 5 μ m (eHPLC column). Data fitting of the van 165 Deemter curves was performed using Origin 6.0 software, while further calculations were 166 performed with MS Excel 2010 software. Van Deemter plots were produced by inspection of 167 column efficiencies using trans-stilbene oxide (TSO) as chiral probe. The data obtained were not 168 corrected for the extra-column peak broadening. For both 5 µm and 1.7 µm columns, the mobile 169 phase consisted of 90/10 *n*-hexane/EtOH +1% MeOH in Normal Phase Liquid Chromatography, 170 and of CO₂/MeOH, 80/20 in SFC conditions. A backpressure of 1800 psi was set in the UHPSFC 171 system. The temperature of the column was set at 25 °C in all experiments, both in LC and SFC. 172 UV detection was performed at 240 nm. The number of theoretical plates N was calculated for every sample according to the European Pharmacopeia using the peak width at half height as implemented in the Chromeleon 6.8 software. An average of two measurements was used for each determination. For the present work, the HETP (H) values were not corrected for the extracolumn volume as, although theoretically correct, it would, nevertheless represent a different situation from the one experimentally observed in the lab.

178 The chiral samples were resolved by using methanol as organic solvent in linear gradient elution 179 (9 min total cycle time): starting from 2% to 30% of MeOH in 5 min, maintaining 30% MeOH 180 for 2 min. The re-equilibration step required 2 min. Mobile phase composition in isocratic 181 elution for each sample was optimized starting from resolution versus retention time scatter plot 182 as explained in result and discussion section. Trifluoroacetic acid (TFA), for neutral and acidic 183 compounds, and ammonia (NH₃) 7N ca. solution in methanol, for basic samples, were used as 184 additives in the organic solvent (0.1 % v/v). For SFC measurements, the following conditions 185 were employed: 124 bar backpressure, 35°C temperature, 3.5 ml/min flow rate and 210–300 nm 186 UV detection. The system pressure ranged from 263 bar at initial condition of gradient to 298 bar 187 at the end. Measurements were repeated twice and average values were used for calculations. 188 Hold-up time was simply estimated from the first negative deviation of the signal. For each 189 enantioseparation, the resolution (Rs) and plate number (N) were calculated according to the 190 European Pharmacopeia using peak width at half height $(w_{0.5})$. In particular, resolution was 191 calculated as:

192 $Rs = 2[(t_r)_2 - (t_r)_1]/(w_{0.5})_2 + (w_{0.5})_1$

where t_r is the retention time. Subscripts 1 and 2 refer to the firstly and secondly eluted peak of racemic sample. The retention time of eluted peaks was not corrected for the dwell volume. The percentage of methanol in the gradient ramp was estimated by proportion considering the maximum value of methanol content as 28% and adding the 2% of initial conditions. That's means: %MeOH $_{2nd peak} = (28 \text{ x tr}_2/5) + 2$. In the result evaluation, peaks with Rs > 2.0 are considered to be "largely separated", those with 1.0 < Rs < 2.0 are "baseline separated" and, finally, peaks with $0.3 < \text{Rs} \le 1.0$ are "partially resolved". Rs = 0 means no separation.

200 **3. Results and Discussion**

3.1 Kinetic performance evaluation.

202 The (S,S) Whelk-O 1, 1.7 μ m CSP has been previously fully characterized in terms of kinetic and 203 thermodynamic properties in normal phase and reversed phase eUHPLC [26]. Before developing 204 the fast gradient screening, an evaluation of the kinetic performance of this column was 205 performed in SFC conditions. Furthermore, by including the 5-µm column in this evaluation, a 206 direct comparison between eHPLC, eUHPLC, eSFC and eUHPFC was obtained and is 207 summarized in Figure 1. Mobile phases were chosen in order to have as similar as possible 208 retention factors between the two columns on the two different systems. Van Deemter curves 209 were then plotted using the first eluting enantiomer of TSO. The H_{min} in eUHPLC is reached at a 210 flow rate twice higher than in eHPLC: 2.04 mL/min and 0.84 mL/min respectively. A H_{min} of 211 4.44 µm and 13.9 µm were respectively obtained for the eUHPLC and eHPLC columns. In 5 µm 212 particle domain, the optimal flow rate was as expected higher in SFC compared to HPLC, 213 corresponding to a two-fold increase in supercritical conditions. Interestingly, the minimum of 214 the van Deemter curve could not be obtained for the 1.7 µm column in eUHPSFC: at the 215 maximum flow rate permitted (4 mL/min) the column efficiency continued increasing, as can be 216 also seen by the descending curve in Figure 1. The number of theoretical plates per meter was in

217 this case 235000 plates at 4 mL/min, but a prediction of the minimum of the van Deemter curve 218 obtained using data fitting as incorporated in Origin 6.0 yielded a flow rate of approximately 219 5.00 mL/min (improved speed factor of 2.4). This would be in agreement with the generally 220 observed that an increase of the optimal flow rate of a factor of 2-2.5 times is to be expected 221 when transitioning from LC to SFC. As a consequence higher linear velocities can be used in 222 UHPSFC without loss in efficiency, leading to a reduced analysis time, which is a basic principle 223 in the transfer from ultra-fast to ultra-high performance conditions. In addition to this, the extra-224 benefit of the use of sub-2-µm particles, advantageous in kinetic terms, allows to further 225 decrease analysis time while gaining in efficiency and thus in resolution achieving the ultra-high 226 performance separation. The chromatographic traces, reported in figure S1, showed that in a 227 quasi-optimal UHPSFC conditions (flow 4.0 mL/min), due to the instrumental limitation, the Rs 228 values were greater than those recorded in UHPLC. The enantiomers of TSO and haloxyfop-2-229 ethoxyethyl (a and c in figure S1) were separated with a gain in Rs of 1.31 and 1.34 factors 230 respectively. For the enantiomers of benzoin (b, figure S1) the slight Rs increase is offset by a 3-231 fold reduced analysis time.

232 **3.2 Fast gradient screening**

Chiral compounds considered in this work have been selected as to cover a broad range of chemically different samples. They include drugs and chiral intermediates of pharmaceutical, biological and medicinal interest (see Table 1 for the complete list of compounds). The chemical structure of all samples has been reported in the Supporting Information (Table S1). As it can be seen the ensemble of compounds considered in this study is extremely heterogeneous both as physico-chemical and structural properties. In the histogram chart of Figure 2, all compounds have been classified according to their principal employment on human and plant diseases. The most represented classes are beta-blockers (18 compounds), agrochemicals (10), antidepressants (10) and antihypertensives (9), covering in total 36% of cases. Samples with unknown activity have been generically classified as "organic compounds" and are 32% of racemates. Of the 129 samples, a useful classification of racemates was done based on a primary chemical property: 33 were acidic (bearing a carboxylic group), 38 neutral and 58 basic (with at least an amino group).

The geometry of the column employed for the fast screening of these compounds, 50×4.6 mm I.D., has been chosen to reduce the effect of extra-column band broadening and simultaneously to allow for short analysis time [25, 31].

248 **Figure 2**

249 The organic modifier employed for the fast gradient screening of compound library was 250 methanol as it combines low viscosity and high polarity and is characterized by a low boiling 251 point (useful behavior for scaling-up to preparative level) if compared to other modifiers often 252 employed in SFC, such as ethanol and isopropanol. In all cases, the gradient program was 253 performed by linearly increasing the amount of methanol in mobile phase from 2 to 30% v/v in 5 254 min and therefore the sub-critical conditions are reached. However, since numerous studies have 255 reported continuity of all chromatographic properties between sub- and super-critical domains 256 [32, 33], in this work we will only speak in terms of super-critical conditions. Depending of 257 compound's nature and to improve peak shape, trifluoroacetic acid (0.1% v/v) was added to the 258 mobile phase for the separation of acidic samples and ammonia (0.1% v/v) was employed for the 259 separation of basic ones. As the presence of additives does not affect the shape of neutral 260 compounds, it was decided to use the same mobile phase to elute both acidic and neutral 261 analytes. This choice is essentially practical and reflects the attempt of performing the entire

screening procedure with minimal changes of experimental conditions. The screening with combined acid and basic additives in a single mobile phase has been reported in literature, but its use is still not widespread [12, 13]. Therefore, the screening of all 129 compounds was achieved with a same gradient program and only two mobile phases (one with methanol/trifluoroacetic acid and the other with methanol/ammonia).

267

268 **Table 1**

269 A flow rate of 3.5 ml/min allowed for ultra-high performance SFC condition, meaning a 270 combination of very fast and highly efficient separations. In fact, most of samples being were 271 eluted and successfully separated in less than 4 min, as shown in Table 1 where the retention 272 time of the latter eluted enantiomer (t_{r2}) , the percentage of methanol when the more retained 273 enantiomer is eluted (%B) and the observed resolution (Rs) are reported. As expected, best 274 results were obtained with acidic and neutral compounds. As an example, two chromatograms 275 showing the enantioseparation of neutral compounds Praziquantel (entry 41 of Table 1, resolved in 3 min with Rs = 5.08), and of Kavain (entry 36, less than 2.5 min analysis time and Rs = 1) and 276 277 the basic compound Mianserine (entry 77 of Table 1, resolved in 3.5 min with Rs = 1.81) are 278 reported on the right side of Figure 3.

However, it is remarkable to notice that the (*S*,*S*) Whelk-O 1, 1.7 μ m CSP performed well also with basic enantiomers. Indeed, 47% of basic compounds was separated with Rs about 0.3. These findings not only confirm the ability of Whelk-O 1 selectors to separate acidic species but somehow show that this CSP can be successfully employed also with basic enantiomers. In terms of efficiency of sample screening, 63% of the analytes (81 of 129 enantiomeric pairs) were resolved and 47 compounds, corresponding to 58% of positive score, have been separated with Rs > 2.0 (Figure 3 left).

286 **Figure 3**

287 For the sake of clarity, the information of the screening has been condensed in Figure 4. The plot 288 on the left refers to the separation of acidic and neutral compounds (mobile phase modifier: 289 methanol plus 0.1% v/v trifluoroaceitc acid), that on the right is for the basic ones (mobile phase 290 modifier: methanol plus 0.1% v/v ammonia). Each point in a plot corresponds to an entry of 291 Table 1 and essentially contains three information: retention time of secondly eluted enantiomer 292 (bottom x-axis), resolution of separation (y-axis) and percentage of mobile phase modifier 293 required for the elution of the more retained enantiomer (top x-axis). Very high values of 294 resolution (Rs >13) were obtained for trans-stilbenoxide (entry 50), Benzoin (entry 51) and 295 Naproxen (entry 11). Most separations were characterized by 1.00 < Rs < 5.00. The zoomed area 296 in Figure 4 (bottom left) refers to the zone characterized by Rs < 4.00 and retention time < 2.5297 min, where a large number of acidic compounds was eluted.

298 **Figure 4**

299 **3.2 Method transfer from gradient to isocratic conditions**

300 Isocratic conditions are usually preferred for purity control (i.e., determination of enantiomeric 301 excess) and purification of chiral samples (i.e., under nonlinear conditions). Going from gradient 302 to isocratic conditions however is not always straightforward, especially if the resolution is low. 303 For samples included in the zoomed area of Figure 4 and with a 1.5 < Rs < 5, we show that a 304 simple empirical criterion can be employed to find suitable conditions for ultrafast separation in

305 isocratic mode. This proof of concept finding is that one should use, in isocratic mode, a mobile 306 phase composition that corresponds to that of the elution of the more retained enantiomer under 307 gradient mode. When Rs becomes larger than roughly 5, this concept helps to get reliable initial 308 conditions from which to start to achieve an ultrafast enantioseparation. For example, the 309 enantiomers of Abscisic acid (entry 4 in Table 1) have been baseline resolved (Rs = 2.21) in less 310 than 60 sec by using a mobile phase containing 15% v/v organic modifier, that is the 311 composition at which the second enantiomer of 4 is eluted in gradient mode (see Table 1). Figure 312 5 (top) reports the corresponding chromatogram. The other chromatograms shown in Figure 5 313 refer to the ultrafast separation of other two racemates, Acenaphthenol (entry 57, middle 314 chromatogram) and Bendroflumethiazide (entry 105, bottom chromatogram). In particular, 315 Acenaphtenol exhibited a very large resolution under gradient conditions (Rs 4.5). To perform an 316 ultrafast separation (less than 1 min) of its enantiomers, we had to use twice the amount of 317 modifier at which the more retained enantiomer was eluted in gradient mode. In this case the 318 information coming from gradient elution was important to set up reliable initial conditions. In 319 cases where resolution is low (Rs < 1.5), separations took a longer time which is in any case very 320 short if compared to that of typical enantioseparations. As an example, the enantiomers of 321 Ketoprofen (entry 6) were *quasi*-baseline separated in less than 2.5 min (Rs = 1.25, see Figure 322 S2-a of Supporting Information) while those of Kavain (entry 36) needed less than 4 min (Rs 323 =0.9, Figure S2-b of Supporting Information).

Figure 5

As a final comment, we want to point out that in cases of low Rs, the use of longer column can help to improve separation. As an example of the very efficient separations that can be achieved in eUHPSFC, the chromatograms for the elution of the enantiomers of two pesticides (Quizalofop-methyl, entry 49, and Flamprop-methyl, entry 48) have been presented in Figure S3
of Supporting Information. In these cases, efficiency larger than 193,000 theoretical plates per
meter were reached by using a 10 cm long column packed with the Whelk-O 1 CSP.

331 4. Conclusions

332 In this proof of concept study we have shown that the high-throughput screening of large 333 compound library by enantioselective SFC is reality. In less than 24 h, by using a 5 cm long 334 column packed with 1.7 µm totally porous Whelk-O 1 particles and fast gradient elution (total 335 analysis time 9 min, including column re-equilibration), 129 racemates with significantly 336 different chemico-physical properties could be screened with a positive result of 63%. In 337 particular, 85% of acidic and neutral analytes have been effectively resolved. Of these 66% have 338 been separated with Rs > 2.0. Moreover, the CSP performed very well also towards the 339 separation of basic compounds, which have been considered not to be good candidates for 340 separation on this chiral selector.

341

342 Acknowledgment

343 This study was supported by PRIN contract no. 2012ATMNJ_003 and by Sapienza University

344 contract no. C26H13ZSR4.

345

346 Conflict of interest

347 Authors declare no conflict of interest.

348

349 **References**

[1] H.–J Federsel, Facing chirality in the 21st century: Approaching the challenges in the
pharmaceutical industry, Chirality 15 (2003) 128–142.

- 352 [2] N.M. Maier, P. Franco, W. Lindner, Separations of enantiomers: needs, challenges,
 353 perspectives, J. Chromatogr. A 906 (2001) 3–33.
- 354 [3] S. Ahuya, Chiral separation methods for pharmaceutical and biotechnological products, John
- 355 Wiley & Sons Inc., Hoboken, USA, 2011.
- 356 [4] Y. Zhang, D.R. Wu, D.B. Wang-Iverson, A.A. Tymiak, Enantioselective chromatography in
- drug discovery, Drug Discov. Today 10 (2005) 571–577.
- 358 [5] E.R. Francotte, Enantioselective chromatography as a powerful alternative for the preparation
- 359 of drug enantiomers, J. Chromatogr. A 906 (2001) 379–397.
- [6] T.J. Ward, K.D. Ward, Chiral separations: a review of current topics and trends, Anal. Chem.
 84 (2012) 626–635.
- 362 [7] D. Mangelings, Y. Vander Heyden, Screening approaches for chiral separations in: E.
- Grushka, N. Grinberg (Eds.), Advances in chromatography, vol. 46, CRC Press, New York, 2007, pp. 175–211.
- [8] L.T. Taylor, Supercritical fluid chromatography for the 21st century, J. Supercrit. Fluids 47
 (2009) 566–573.
- [9] N. Matthijs, M. Maftouh, Y. Vander Heyden, Chiral separation strategy in polar organic
 solvent chromatography and performance comparison with normal–phase liquid and
 supercritical–fluid chromatography, J. Sep. Sci. 29 (2006) 1353–1362.
- 370 [10] M. Maftouh, C. Granier–Loyaux, E. Chavana, J. Marini, A. Pradines, Y. Vander Heyden, C.
- 371 Picard, Screening approach for chiral separation of pharmaceuticals. Part III. Supercritical fluid
- 372 chromatography for analysis and purification in drug discovery, J. Chromatogr. A 1088 (2005)373 67–81.
- [11] K. De Klerck, G. Parewyck, D. Mangelings, Y. Vander Heyden, Enantioselectivity of
 polysaccharide–based chiral stationary phase in supercritical fluid chromatography using
 methanol–containing carbon dioxide mobile phases, J. Chromatogr. A 1269 (2012) 336–345.
- 377 [12] K. De Klerck, Y. Vander Heyden, D. Mangelings, Generic chiral method development in
- 378 supercritical fluid chromatography and ultra-performance supercritical fluid chromatography, J.
- 379 Chromatogr. A 1363 (2014) 311–322.
- 380 [13] K. De Klerck, D. Mangelings, D. Clicq, F. De Boever, Y. Vander Heyden, Combined use of
- 381 isopropylamine and trifluoroacetic acid in methanol-containing mobile phases for chiral
- 382 supercritical fluid chromatography, J. Chromatogr. A 1234 (2012) 72–79.

- 383 [14] J.O. DaSilva, B. Coes, L. Frey, I. Mergelsberg, R. McClain, L. Nogle, C.J. Welch,
- Evaluation of non-conventional polar modifiers on immobilized chiral stationary phases for
 improved resolution of enantiomers by supercritical fluid chromatography, J. Chromatogr. A
- 386 1328 (2014) 98–103.
- 387 [15] K. De Klerck, C. Tistaert, D. Mangelings, Y. Vander Heyden, Updating a generic screening
- 388 approach in sub– or supercritical chromatography for the enantioresolution of pharmaceuticals, J.
- 389 Supercrit. Fluids 80 (2013) 50–59.
- 390 [16] D. Mangelings, Y. Vander Heyden, Chiral separations in sub– and supercritical fluids
 391 chromatography, J. Sep. Sci. 31 (2008) 1252–1273.
- 392 [17] K. De Klerck, D. Mangelings, Y. Vander Heyden, Supercritical fluid chromatography for
- the enantioseparation of pharmaceuticals, J. Pharm. and Biomed Analysis 69 (2012) 77–92.
- 394 [18] C. Hamman, M. Wong, M. Hayes, P. Gibbons, A high throughput approach to purifying
- 395 chiral molecules using $3\mu m$ analytical chiral stationary phases via supercritical fluid
- 396 chromatography, J. Chromatogr. A, 1218 (2011) 3529–3536.
- 397 [19] W. Schafer, T. Chandrasekaran, Z. Pirzada, C. Zhang, X. Gong, M. Biba, E. L. Regalado, C.
- J. Welch, Improved chiral SFC screening for analytical method development, Chirality 25 (2013)
 799–804.
- 400 [20] M. Biba, E. L. Regalado, N. Wua, C. J. Welch, Effect of particle size on the speed and
- 401 resolution of chiral separations using supercritical fluid chromatography, J. Chromatogr A, 1363
- 402 (2014) 250–256.
- 403 [21] C. J. Welch, E. L. Regalado, Estimating optimal time for fast chromatographic separations,
 404 J. Sep. Sci. 2014, *37*, 2552–2558.
- 405 [22] C. Hamman, M. Wong, I. Aliagas, D.F. Ortwine, J. Pease, D.E. Schmidt jr., J. Victorino, J.
- 406 Chromatogr. A 1305 (2013) 310–319.
- 407 [23] A. Cavazzini, N. Marchetti, R. Guzzinati, M. Pierini, A. Ciogli, D. Kotoni, I. D'Acquarica,
- 408 C. Villani, F. Gasparrini, Enantioseparation by ultra-high-performance liquid chromatography,
- 409 TrAC (2014), doi: 10.1016/j.trac.2014.06.026.
- 410 [24] G. Cancelliere, A. Ciogli, I. D'Acquarica, F. Gasparrini, J. Kocergin, D. Misiti, M. Pierini,
- 411 H. Ritchie, P. Simone, C. Villani, Transition from enantioselective high performance to ultra-
- 412 high performance liquid chromatography: A case study of a brush-type chiral stationary phase
- 413 based on sub-5-micron to sub-2-micron silica particles, J. Chromatogr. A 1217 (2010) 990–999.

- 414 [25] D. Kotoni, A. Ciogli, C. Molinaro, I. D'Acquarica, J. Kocergin, T. Szczerba, H.
- 415 Ritchie, C. Villani, F. Gasparrini, Introducing enantioselective UHPLC: theoretical inspections
- 416 and ultra-fast separations on a new sub-2 μm Whelk-O 1 stationary phase, Anal. Chem. 84
 417 (2012) 6805–6813.
- 418 [26] D. Kotoni, A. Ciogli, C. Molinaro, I. DAcquarica, J. Kocergin, T. Szczerba, H. Ritchie, C.
- 419 Villani, F. Gasparrini, Enantioselective ultra-high and high performance liquid chromatography:
- 420 A comparative study of columns based on the Whelk-O 1 selector. J. Chromatogr. A 1269 (2012)
 421 226–241.
- 422 [27] C. J. Welch, T. Szczerba, S.R. Perrin, Some recent high-performance liquid chromatography
- 423 separations of the enantiomers of pharmaceuticals and other compounds using the Whelk-O 1
- 424 chiral stationary phase, J. Chromatogr. A, 758 (1997) 93-98.
- 425 [28] A. Grand–Guillaume Perrenoud, J.–L. Veuthey, D. Guillaume, Comparison of ultra–high
- 426 performance supercritical fluid chromatography and ultra-high performance liquid
 427 chromatography for the analysis of pharmaceutical compounds, J. Chromatogr. A 1266 (2012)
 428 158–167.
- 429 [29] W.H. Pirkle, C.J. Welch, B. Lamm, Design, synthesis, and evaluation of an improved
 430 enantioselective naproxen selector, J. Org. Chem. 57 (1992) 3854–3860.
- [30] W.H. Pirkle, C.J. Welch, An improved chiral stationary phase for the chromatographic
 separation of underivatized naproxen enantiomers, J. Liq. Chromatogr. 15 (1992) 1947–1955.
- 433 [31] S. Fekete, I. Kohler, S. Rudaz, D. Guillarme, Importance of instrumentation for fast liquid
 434 chromatography in pharmaceutical analysis, J. Pharm. Bio. Analysis 87 (2014) 105–119
- [32] T.A. Berger, Density of methanol-carbon dioxide mixtures at three temperatures:
 Comparison with vapor-liquid equilibria measurements and results obtained from
 chromatography, J. High Resolut. Chromatogr. 14 (1991) 312–316.
- 438 [33] J.A. Blackwell, R.W. Stringham, J.D. Weckwerth, Effect of mobile phase additives in
- 439 packed-column subcritical and supercritical fluid chromatography, Anal. Chem. 69 (1997) 409–
- 440 415.
- 441
- 442
- 443
- 444

445 Figure captions.

Figure 1. Comparison of van Deemter plots in LC and SFC at 25 °C for the first eluting enantiomer of TSO. Columns: A) Whelk-O 1, 150 mm x 4.6 mm I.D., 5 μ m, red and orange curves; B) Whelk-O 1, 50 mm x 4.6 mm I.D., 1.7 μ m, blue and green curves. Data not corrected for extra-column band broadening effect. (For interpretation of the colors in this legend, please refer to the web version of the article).

Figure 2. Classification of investigated samples based on their chemical properties (acidic, neutral, basic) and pharmacological activity. For those compounds whose pharmaceutical activity is unknown/unspecified the generic expression "organic compounds" has been employed.

Figure 3. Left: pie chart representing numerical proportion of resolved enantiomers with Rs>2,
1<Rs<2 and 0.3<Rs<1.0. Right: Examples of experimental chromatograms in gradient elution.
Top: Praziquantel, entry 41 of Table 1; middle: Kavain, entry 36; bottom: Mianserine, entry 77.

(For interpretation of the colors, please refer to the web version of the article).

Figure 4. Scatter plots showing the results of the screening. Left: acidic and neutral samples. Zoomed area (bottom left) corresponds to the zones where the largest part of acidic compounds has been eluted. Right: basic samples. Limit values of Rs (0.9 for acidic and neutral compounds and 0.3 for basic ones) have been represented in the plots by dotted lines. See text for details.

463 Figure 5. Examples of ultrafast enantioseparations in isocratic conditions. Experimental details464 in the text.

465

458