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# **ORIGINAL ARTICLE** Prognostic and biologic significance of *DNMT3B* expression in older patients with cytogenetically normal primary acute myeloid leukemia

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*DNMT3B* encodes a DNA methyltransferase implicated in aberrant epigenetic changes contributing to leukemogenesis. We tested whether *DNMT3B* expression, measured by NanoString nCounter assay, associates with outcome, gene and microRNA expression and DNA methylation profiles in 210 older ( $\geq$ 60 years) adults with primary, cytogenetically normal acute myeloid leukemia (CN-AML). Patients were dichotomized into high versus low expressers using median cut. Outcomes were assessed in the context of known CN-AML prognosticators. Gene and microRNA expression, and DNA methylation profiles were analyzed using microarrays and MethylCap-sequencing, respectively. High *DNMT3B* expressers had fewer complete remissions (CR; P = 0.002) and shorter disease-free (DFS; P = 0.02) and overall (OS; P < 0.001) survival. In multivariable analyses, high *DNMT3B* expression remained an independent predictor of lower CR rates (P = 0.04) and shorter DFS (P = 0.04) and OS (P = 0.001). High *DNMT3B* expression associated with a gene expression profile comprising 363 genes involved in differentiation, proliferation and survival pathways, but with only four differentially expressed microRNAs (*miR-133b*, *miR-148a*, *miR-122*, *miR-409-3p*) and no differential DNA methylation regions. We conclude that high *DNMT3B* expression independently associates with adverse outcome in older CN-AML patients. Gene expression analyses suggest that *DNMT3B* is involved in the modulation of several genes, although the regulatory mechanisms remain to be investigated to devise therapeutic approaches specific for these patients.

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# INTRODUCTION

Acute myeloid leukemia (AML) is a heterogeneous disease presenting with a wide spectrum of prognostically relevant cytogenetic aberrations, gene mutations and abnormal expression of genes and microRNAs. Cytogenetically normal AML (CN-AML) patients, constituting 40 to 50% of all AML patients,<sup>1</sup> are the largest and molecularly best characterized cytogenetic subset in primary (*de novo*) AML.<sup>1–3</sup> Although leukemic blasts of these patients do not contain microscopically detectable chromosome abnormalities, they harbor prognostically relevant mutations and aberrantly expressed genes and microRNAs.<sup>2–16</sup> In addition to these genetic alterations, epigenetic changes have recently been shown to participate in myeloid leukemogenesis and be pharmacologically targetable.<sup>17,18</sup> Notably, some genes whose mutations are prognostic in CN-AML encode proteins that are

implicated in epigenetic regulation of gene transcription, namely *IDH2, ASXL1* and *DNMT3A*. The latter is among the most frequently mutated genes in primary CN-AML patients, being found mutated in 29 to 34% of the patients.<sup>9,19</sup>

*DNMT3A* encodes DNA methyltransferase 3A (DNMT3A), which is involved in epigenetic gene silencing through DNA hypermethylation.<sup>20</sup> In addition to DNMT3A, DNMT1 and DNMT3B also mediate DNA methylation in normal and malignant cells, and may represent potential therapeutic targets in cancer and leukemia.<sup>21–24</sup> However, in contrast to *DNMT3A*, no recurrent mutations of *DNMT1* and *DNMT3B* genes have been reported in AML.<sup>25</sup> Instead, one study has indicated that higher expression of *DNMT3B* is associated with worse outcome in AML.<sup>26</sup> However, the patient cohort analyzed was cytogenetically diverse and heterogeneous for clinical features and treatment received. Thus, it is

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unknown whether *DNMT3B* expression is an independent prognostic factor and can be used for stratification guidance in CN-AML.

Thus, we analyzed the clinical significance of *DNMT3B* expression in the context of a comprehensive panel of molecular prognosticators in a relatively large cohort of older (aged  $\ge$  60 years) patients with CN-AML who were similarly treated on cytarabine/daunorubicin-based protocols. To gain biologic insights, we also derived genome-wide *DNMT3B*-associated gene and microRNA expression and DNA methylation profiles. We studied older patients because both the incidence of AML and the role of epigenetics increase with age. Moreover, we have recently reported a favorable clinical response to hypomethylating agents in this age group of AML patients.<sup>27</sup>

# PATIENTS AND METHODS

#### Patients, treatment and cytogenetic studies

Pretreatment bone marrow or blood samples were obtained from 210 patients with primary CN-AML aged 60 to 83 years (median, 68 years) who received intensive first-line therapy on Cancer and Leukemia Group B (CALGB) trials.<sup>28–32</sup> All patients received cytarabine–daunorubicin-based induction chemotherapy, and no patient received allogeneic hematopoietic stem cell transplantation during first complete remission (CR). For details regarding treatment protocols and sample collection, see Supplementary Information. All patients were enrolled on companion CALGB/Alliance protocols: 8461 (cytogenetic analyses), 9665 (tissue banking) and 20 202 (molecular analyses).

Cytogenetic analyses were performed in institutional CALGB/Alliance cytogenetics laboratories. For the patient's karyotype to be considered normal,  $\ge 20$  metaphase cells from short-term cultures of pretreatment bone marrow specimens had to have been analyzed and the normal result confirmed by central karyotype review.<sup>33</sup> All patients provided written informed consent for participation in these studies; study protocols were in accordance with the Declaration of Helsinki and approved by local Institutional Review Boards.

#### Single-gene expression analyses

The expression of *DNMT3B* transcript was assessed by NanoString nCounter assays (NanoString Technologies, Seattle, WA, USA; Supplementary Information).<sup>34</sup> These assays measured global expression of the *DNMT3B* gene, and did not allow for quantification of isoform-specific expression of *DNMT3B*. *DNMT3B* expression levels were normalized using *ABL* as an internal control. We also used NanoString nCounter assays to measure expression of *BAALC*, *ERG* and *miR-155*, and real-time reverse transcription-PCR to measure *miR-3151* expression, all of which have been previously shown to affect prognosis of older CN-AML patients.<sup>10,15,16</sup>

# Mutational analyses

The presence or absence of *FLT3* internal tandem duplication (*FLT3*-ITD), <sup>35,36</sup> *FLT3* tyrosine kinase domain mutations (*FLT3*-TKD), <sup>37</sup> *MLL* partial tandem duplication (*MLL*-PTD), <sup>38</sup> and mutations in the *NPM1*, <sup>5</sup> *CEBPA*, <sup>39</sup> *WT1*, <sup>40</sup> *IDH1* and *IDH2*, <sup>7</sup> *TET2*, <sup>41</sup> *ASXL1*, <sup>8</sup> *DNMT3A*<sup>9</sup> and *RUNX1*<sup>42</sup> genes were determined centrally as previously described.

# Gene and microRNA expression profiling

The gene and microRNA expression profiling were assessed using the Affymetrix U133 plus 2.0 array (Affymetrix, Santa Clara, CA, USA) and The Ohio State University custom microRNA array (OSU\_CCC Version 4.0, The Ohio State University, Columbus, OH, USA), respectively, as previously reported,<sup>5,43</sup> and detailed in the Supplementary Information. For *DNMT3B*, the Affymetrix U133 plus 2.0 arrays measured global *DNMT3B* expression levels, and did not quantify expression of the individual *DNMT3B* isoforms. For the gene and microRNA expression profiling, summary measures of gene and microRNA expression profiling, summary measures of gene and microRNA expression for details) was derived by comparing gene expression between high and low *DNMT3B* expressers in the CALGB/Alliance cohort and in two additional sets of CN-AML patients with microarray and RNAseq gene expression data publicly available (German AML Cooperative Group (AMLCG)<sup>44</sup> and The Cancer Genome Atlas (TCGA)<sup>25</sup>). For comparison of the high *DNMT3B* 

expression signature with the *FLT3*-ITD signature we used gene set enrichment analysis (for details see Supplementary Information). For the microRNA expression signature, only the CALGB/Alliance patients were used. Univariable significance levels of *P* < 0.001 (false discovery rates < 0.01) were used to select genes and microRNAs that constituted the signatures. To assess enrichment of genes in the *DNMT3B* gene expressionassociated signature in distinct biologic processes, a Gene Ontology analysis was performed using the Database for Annotation, Visualization and Integrated Discovery (DAVID).<sup>45</sup> We identified as statistically significant 'annotation clusters' those clusters of Gene Ontology terms with enrichment scores of > 2.0, *P*-values  $\leq$  0.001 and Benjamini corrected *P*-values  $\leq$  0.05. All molecular analyses were performed centrally at The Ohio State University.

#### DNA methylation

Genome-wide DNA methylation and levels of DNA methylation across the genome's functional regions (that is, genomic features) were measured using the MethylCap-seq assay as previously reported.<sup>18</sup>

#### Statistical analyses

The patients were dichotomized into high and low expressers using the median cut. This cut was supported by significant results of the trend test applied to outcome of patients divided into quartiles by DNMT3B expression ( $P \leq 0.001$ ). We compared pretreatment features and outcome between patients with high and low DNMT3B expression. Definitions of clinical end points (that is, CR rates, disease-free (DFS) and overall (OS) survival) are provided in the Supplementary Information. Baseline characteristics between high and low DNMT3B expressers were compared using the Fisher's exact test for categorical and the Wilcoxon rank-sum test for continuous variables.<sup>46</sup> The categorical variables included the European LeukemiaNet (ELN) Genetic Groups.<sup>47</sup> The ELN guidelines classify CN-AML patients within the Favorable or Intermediate-I Genetic Groups based on CEBPA, NPM1 and FLT3 mutational status. The ELN Favorable Genetic Group consists of CN-AML patients with CEBPA mutation and/or NPM1 mutation without FLT3-ITD, whereas the Intermediate-I Genetic Group is comprised of patients with wild-type CEBPA and FLT3-ITD with or without NPM1 mutation, or wild-type NPM1 without FLT3-ITD.47

For time-to-event analyses, we calculated survival estimates using the Kaplan–Meier method, and compared groups by the log rank test.<sup>46</sup> In order to provide the odds ratios and hazard ratios and associated confidence intervals, logistic regression and Cox proportional hazards models were generated to compare outcomes between high and low *DNMT3B* expressers for CR and survival end points (DFS, OS), respectively, and *P*-values from the Wald test are reported. We constructed multivariable logistic regression models to analyze factors associated with the achievement of CR, and multivariable Cox proportional hazards models for factors associated with survival end points,<sup>46</sup> the details of which are provided in the Supplementary Information. All analyses were performed by the Alliance for Clinical Trials in Oncology Statistics and Data Center.

# RESULTS

Associations of *DNMT3B* expression with pretreatment clinical and molecular characteristics

At diagnosis, high *DNMT3B* expressers had higher white blood counts (WBC; P = 0.004), and percentages of blood (P = 0.004) and marrow (P = 0.02) blasts than low *DNMT3B* expressers. Concerning molecular features, high *DNMT3B* expressers were more often *FLT3*-ITD-positive (P < 0.001) and classified in the ELN Intermediate-I Genetic Group (P = 0.02). *IDH2*-R140 mutations were less frequent in high *DNMT3B* expressers, whereas six of seven *IDH2*-R172 mutations were detected in this patient group. High *DNMT3B* expressers also had higher *ERG* (P < 0.001), *BAALC* (P = 0.002) and *miR-155* (P = 0.006) expression than low expressers (Table 1, Supplementary Figure S1).

Associations of *DNMT3B* expression with clinical outcome in the entire patient cohort

With a median follow-up of 5.1 years (range, 2.3–11.6 years) for patients who are alive, high *DNMT3B* expressers had lower CR

**Table 1.** Comparison of clinical and molecular characteristics ofpatients with cytogenetically normal acute myeloid leukemia withhigh versus low DNMT3B expression

Characteristic	High DNMT3B (n = 105)	Low DNMT3B (n = 105)	P-value
<i>Age, years</i> Median Range	68 60–83	68 60–81	0.82
<i>Sex,</i> n <i>(%)</i> Male Female	58 (55) 47 (45)	53 (50) 52 (50)	0.58
<i>Race,</i> n (%) White Nonwhite	96 (92) 8 (8)	92 (89) 11 (11)	0.48
<i>Hemoglobin, g/dl</i> Median Range	9.4 6.5–12.4	9.3 5.4–15.0	0.71
<i>Platelet count, x 10<sup>9</sup>/l</i> Median Range	68 4–850	71 11–510	0.87
<i>WBC, x 10<sup>9</sup>/I</i> Median Range	43.7 1.0–450.0	21.8 0.8–249.3	0.004
<i>Blood blasts, %</i> Median Range	64 0–99	40 0–97	0.004
<i>Bone marrow blasts, %</i> Median Range	72 21–97	64 4–97	0.02
Extramedullary involvement, n (%)	27 (27)	24 (23)	0.63
<i>NPM1,</i> n (%) Mutated Wild type	67 (66) 34 (34)	60 (59) 42 (41)	0.31
<i>FLT3-ITD</i> , n (%) Present Absent	54 (53) 48 (47)	21 (21) 81 (79)	< 0.001
CEBPA, n (%) Mutated Single mutated Double mutated Wild type	13 (13) 10 3 88 (87)	12 (12) 5 7 90 (88)	0.83 <sup>a</sup>
<i>ELN Genetic Group</i> , n (%) <sup>b</sup> Modified Favorable Intermediate-I	38 (38) 63 (62)	56 (55) 45 (45)	0.02
<i>FLT3-TKD,</i> n (%) Present Absent	12 (12) 89 (88)	11 (11) 91 (89)	0.83
<i>WT1,</i> n (%) Mutated Wild type	8 (8) 93 (92)	5 (5) 97 (95)	0.41
<i>TET2,</i> n <i>(%)</i> Mutated Wild type	32 (32) 68 (68)	31 (32) 67 (68)	1.00
<i>MLL-PTD</i> , n (%) Present Absent	5 (6) 74 (94)	5 (6) 81 (94)	1.00
<i>IDH1,</i> n (%) Mutated Wild type	14 (14) 86 (86)	8 (8) 94 (92)	0.18

Table 1. (Continued)			
Characteristic	High DNMT3B (n = 105)	<i>Low</i> <i>DNMT3B</i> (n = 105)	P-value
<i>IDH2,</i> n <i>(%)</i> Mutated R140	18 (18) 12	31 (30) 30	0.05 0.005 <sup>c</sup>
R172 Wild type	6 82 (82)	1 71 (70)	0.13ª
<i>RUNX1,</i> n (%) Mutated Wild type	17 (18) 79 (82)	11 (12) 81 (88)	0.31
<i>ASXL1,</i> n (%) Mutated Wild type	13 (13) 87 (87)	15 (15) 84 (85)	0.69
DNMT3A, n (%) Mutated R882 Non-R882 Wild type	35 (35) 19 16 64 (65)	31 (32) 20 11 66 (68)	0.65 1.00 <sup>e</sup> 0.40 <sup>f</sup>
<i>ERG expression group</i> , n (%) <sup>9,h</sup> High Low	65 (62) 40 (38)	40 (38) 65 (62)	< 0.001
<i>BAALC expression group,</i> n (%) <sup>g,h</sup> High Low	64 (61) 41 (39)	41 (39) 64 (61)	0.002
<i>miR-155 expression group</i> , n (%) <sup>g,h</sup> High Low	63 (60) 42 (40)	42 (40) 63 (60)	0.006
<i>miR-3151 expression group,</i> n (%) <sup>g,i</sup> High Low	42 (49) 43 (51)	39 (46) 46 (54)	0.76

Abbreviations: ELN, European LeukemiaNet; FLT3-ITD, internal tandem duplication of the FLT3 gene; FLT3-TKD, tyrosine kinase domain mutation in the FLT3 gene; MLL-PTD, partial tandem duplication of the MLL gene; n, number; WBC, white blood count. <sup>a</sup>The P-value pertains to a comparison of frequencies of CEBPA mutations (single and double combined) versus CEBPA wild-type between high and low DNMT3B expressers. <sup>b</sup>The ELN modified Favorable Genetic Group is defined as CN-AML patients with mutated CEBPA and/or mutated NPM1 without FLT3-ITD. All remaining CN-AML patients (that is, those with wild-type CEBPA and wild-type NPM1 with or without FLT3-ITD, or mutated NPM1 with FLT3-ITD) belong to the ELN Intermediate-I Genetic Group.<sup>47</sup> <sup>c</sup>The *P*-value pertains to a comparison of frequencies of IDH2-R140 mutations versus IDH2 wild-type between high and low DNMT3B expressers. <sup>d</sup>The P-value pertains to a comparison of frequencies of IDH2-R172 mutations versus IDH2 wild-type between high and low DNMT3B expressers. <sup>e</sup>The P-value pertains to a comparison of frequencies of DNMT3A-R882 mutations versus DNMT3A wild-type between high and low DNMT3B expressers. <sup>f</sup>The P-value pertains to a comparison of frequencies of DNMT3A non-R882 mutations versus DNMT3A wild-type between high and low DNMT3B expressers. <sup>9</sup>The median expression value was used as a cut point. <sup>h</sup>Data was assessed by the NanoString nCounter assay. <sup>i</sup>Data was assessed by real-time reverse transcription-PCR.

rates (P = 0.002, Wald test; 58% vs 78%), and shorter DFS (P = 0.02, Wald test) and OS (P < 0.001, Wald test) than *DNMT3B* low expressers (Table 2, Figure 1).

In a multivariable model for CR, *DNMT3B* expression remained prognostic (P = 0.04), after adjustment for *BAALC* expression status (P < 0.001), WBC (P = 0.007) and age (P = 0.02) (Table 3). High *DNMT3B* expressers were half as likely to achieve a CR as low expressers. In multivariable analysis for DFS, high *DNMT3B* expression associated with shorter DFS (P = 0.04), once adjusted for *BAALC* expression (P = 0.004), *DNMT3A*-R882 mutation status (P = 0.009) and ELN Genetic Groups (P = 0.03). The risk of experiencing relapse or death was 46% higher for high *DNMT3B* 

End point	High DNMT3B (I)	Low DNMT3B (II)	P-value <sup>a</sup>	OR/HR (95% CI) I vs II
All patients, n	105	105		
Complete remission, n (%)	61 (58)	82 (78)	0.002	0.39 (0.21–0.71)
Disease-free survival			0.02	1.55 (1.09–2.20)
Median, years	0.6	1.1		
% Disease-free at 3 years (95% CI)	13 (6–23)	22 (14–31)		
% Disease-free at 5 years (95% CI)	11 (5–21)	15 (8–23)		
Overall survival			< 0.001	1.85 (1.38–2.47)
Median, years	0.8	1.5		
% Alive at 3 years (95% CI)	11 (6–18)	29 (20-37)		
% Alive at 5 years (95% CI)	8 (4–14)	21 (14–29)		
Patients in the ELN modified Favorable Genetic Group <sup>b</sup> , n	38	56		
Complete remission, n (%)	27 (71)	47 (84)	0.14	0.47 (0.17-1.28)
Disease-free survival			0.10	1.54 (0.92–2.57)
Median, years	0.9	1.3		
% Disease-free at 3 years (95% CI)	15 (5–30)	30 (18–43)		
% Disease-free at 5 years (95% CI)	11 (3–26)	21 (11–34)		
Overall survival			0.002	2.04 (1.29–3.22)
Median, years	1.3	2.1		
% Alive at 3 years (95% CI)	18 (7–31)	39 (27–52)		
% Alive at 5 years (95% CI)	12 (4–24)	30 (19–43)		
Patients in the ELN Intermediate-I Genetic Group <sup>b</sup> , n	63	45		
Complete remission, n (%)	31 (49)	33 (73)	0.01	0.35 (0.15–0.80)
Disease-free survival			0.25	1.36 (0.81–2.27)
Median, years	0.5	0.7		
% Disease-free at 3 years (95% CI)	13 (4–27)	9 (2–22)		
% Disease-free at 5 years (95% CI)	13 (4–27)	3 (0–13)		
Overall survival			0.03	1.57 (1.06–2.33)
Median, years	0.6	1.1		
% Alive at 3 years (95% CI)	8 (3–16)	16 (7–28)		
% Alive at 5 years (95% CI)	6 (2–14)	9 (3–19)		

Abbreviations: CI, confidence interval; ELN, European Leukemia Net; HR, hazard ratio; n, number; OR, odds ratio. <sup>a</sup>*P*-values provided are generated by logistic regression and Cox proportional hazards models to compare outcome of patients for CR and survival end points (DFS, OS), respectively, using the Wald test. <sup>b</sup>The ELN modified Favorable Genetic Group is defined as CN-AML patients with mutated *CEBPA* and/or mutated *NPM1* without *FLT3*-ITD. All remaining CN-AML patients (that is, those with wild-type *CEBPA* and wild-type *NPM1* with or without *FLT3*-ITD, or mutated *NPM1* with *FLT3*-ITD) belong to the ELN Intermediate-I Genetic Group.<sup>47</sup>

expressers than for low expressers. *DNMT3B* expression also remained prognostic for OS (P = 0.001), after adjustment for *BAALC* (P < 0.001), *miR-3151* (P = 0.02) and *miR-155* (P = 0.02) expression. The risk of death was 72% higher for high *DNMT3B* expressers compared with low expressers (Table 3).

# Associations of *DNMT3B* expression with clinical outcome in ELN Genetic Groups

We analyzed the associations of *DNMT3B* expression with outcome separately within the ELN Favorable and Intermediate-I Genetic Groups. Within the Favorable Group (n = 94), there was no significant difference in CR rates (71% vs 84%, P = 0.14, Wald test) or DFS (P = 0.10, Wald test) between high and low *DNMT3B* expressers. However, high expressers had shorter OS (P = 0.002, Wald test) than low expressers (Table 2, Figures 2a and b). In multivariable analyses for the ELN Favorable Genetic Group (Table 3), *DNMT3B* expression remained significant for OS (P = 0.003) after adjustment for *BAALC* expression (P = 0.01). High *DNMT3B* expressers were twice as likely to die as low expressers.

In the Intermediate-I Group (n = 108), high *DNMT3B* expressers had a lower CR rate (49% vs 73%, P = 0.01, Wald test) and shorter OS (P = 0.03, Wald test) than low *DNMT3B* expressers, but there was no significant difference in DFS between the groups (Table 2, Figures 2c and d). In multivariable analyses within the ELN Intermediate-I Genetic Group (Table 3), *DNMT3B* expression was significant for OS (P = 0.02), after adjustment for *BAALC* expression (P = 0.03), miR-3151 expression (P = 0.03) and WBC (P = 0.03). High DNMT3B expressers were 1.7 times more likely to die than low expressers.

Genome-wide gene expression profiles associated with DNMT3B expression

To gain biologic insights into the role of DNMT3B, we derived a DNMT3B-associated gene expression profile using three independent sets of CN-AML patients, that is, CALGB/Alliance (n = 177), AMLCG (n = 75) and The Cancer Genome Atlas (n = 88). We identified 195 upregulated genes and 168 downregulated genes that were significantly associated with higher DNMT3B expression in each of the three cohorts (Supplementary Table S1). As high DNMT3B expression was associated with the presence of FLT3-ITD (Table 1), we performed gene set enrichment analysis to test whether a set of 195 genes that are upregulated in high DNMT3B expressers was associated with a set of genes differentially expressed between patients who harbored FLT3-ITD versus those who did not (Supplementary Information). We found a significant correlation between the high DNMT3B expression and FLT3-ITD signatures (P = 0.006; false discovery rate = 0.006; Supplementary Figure S2). Among the genes upregulated in high DNMT3B expressers, we noted a variety of genes previously involved in AML including CDK6 and WT1 that encode cyclin kinase and transcription factor proteins, respectively. Among the downregulated genes, we noted genes involved with both normal



**Figure 1.** Clinical outcome of CN-AML patients with high and low *DNMT3B* expression. Kaplan–Meier survival curves for (**a**) disease-free survival and (**b**) overall survival. *P*-values presented are from the log rank test.

monocyte/macrophage differentiation and immune function including *CD14*, *TLR4*, *CEBPB* and *TLR8*.

Gene Ontology was used to assess the biologic features of the *DNMT3B* expression profile (Table 4). For *DNMT3B*-associated upregulated genes, there were three Gene Ontology terms comprising genes involved in nucleotide biosynthetic processes and metabolism and included in annotation cluster 1 that had a trend for statistical significance (Benjamini *P*-value < 0.1). For *DNMT3B*-associated downregulated genes, cellular processes included lysosome biology, endocytosis and membrane signaling. These results may be interpreted as consistent with the previously noted dysregulated genes involved in monocyte/macrophage differentiation and activity.

# Genome-wide microRNA profiles associated with DNMT3B expression

The influence of *DNMT3B* expression on microRNA genome-wide profiles could be evaluated in 162 patients. In contrast to coding genes, only four microRNAs were differentially expressed between high and low *DNMT3B* expressers ( $P \le 0.001$ ). High *DNMT3B* expression was associated with *miR-133b* upregulation, and *miR-148a*, *miR-122* and *miR-409-3p* downregulation. *miR-133b* upregulation in high *DNMT3B* expressers was somewhat surprising as this microRNA was reported to have tumor suppressor activity in other cancers.<sup>48,49</sup> However, consistent with the downregulated gene expression profile as discussed above, *miR-133b* has recently been shown to target granulocyte-macrophage colonystimulating factor (GM-CSF), a cytokine involved in granulocytemonocyte/macrophage differentiation.<sup>50</sup> Among the downregulated



microRNAs, *miR-148a* was reported to target *DNMT3B* and to be itself a target of aberrant hypermethylation in cancer.<sup>51,52</sup> Lower expression of *miR-122* has been associated with aggressive hepatocellular carcinoma and *miR-409-3p* with cell invasion and metastasis in gastric cancer<sup>53–56</sup>; however, a role for these microRNAs in AML is currently unknown.

Genome-wide methylation profiling associated with DNMT3B expression

As *DNMT3B* encodes a methyltransferase that mediates *de novo* DNA methylation, we assessed whether high and low *DNMT3B* expressers differed in DNA methylation patterns. Surprisingly, we found no significant differences in genome-wide DNA methylation levels or in the numbers of differentially methylated regions<sup>18</sup> in distinct functional genomic regions (for example, gene promoters) when high versus low *DNMT3B* expressers were compared.

#### DISCUSSION

In this study, we report that high *DNMT3B* expression associates with lower CR rates and shorter DFS and OS in chemotherapy-treated CN-AML patients aged  $\ge 60$  years. High *DNMT3B* expression was associated with such adverse prognostic factors as *FLT3*-ITD, high *ERG*, *BAALC* and *miR-155* expression and the ELN Intermediate-I Genetic Group; nevertheless the association of *DNMT3B* expression with clinical outcome is independent from the aforementioned and other established molecular and clinical prognosticators for all outcome end points studied.

Our findings are consistent to some extent with the only, to our knowledge, previous study that assessed the prognostic value of *DNMT3B* expression.<sup>26</sup> Although, in the subset of 93 CN-AML patients, Hayette *et al.*<sup>26</sup> did not find significant differences in event-free survival or OS between high and low *DNMT3B* expressers, high *DNMT3B* expressers had a shorter event-free survival than low *DNMT3B* expressers in the whole cytogenetically diverse cohort of 191 AML patients analyzed. It is difficult to directly compare their results with ours as approximately one-half of the patients analyzed by Hayette *et al.*<sup>26</sup> had various abnormal karyotypes, more than two-thirds of the patients were younger than 60 years and a quarter underwent allogeneic hematopoietic stem cell transplantation in first CR. Thus, although the two studies are not comparable, they both conclude that higher *DNMT3B* expression is associated with worse outcome in AML.

Recently, the ELN reporting system,<sup>47</sup> which for CN-AML is based on only three molecular markers (that is, FLT3-ITD, CEBPA and *NPM1* mutations), was shown to provide important prognostic information in AML.<sup>57</sup> However, we and others have shown that additional molecular markers, such as TET2,<sup>41</sup> ASXL1,<sup>8</sup> RUNX1<sup>42</sup> and DNMT3A<sup>58</sup> mutations and expression of MN1,<sup>12</sup> miR-155<sup>15</sup> and miR-3151,16 may refine outcome prediction of CN-AML patients within the ELN Genetic Groups. Hence, in the current study, we investigated whether considering DNMT3B expression as a novel prognosticator could alter patient classification within the ELN Genetic Groups. In the Favorable Group, we found that low DNMT3B expression identified a subset of CN-AML patients with a significantly longer OS, thus making DNMT3B expression the third molecular marker, in addition to ASXL1 mutations<sup>8</sup> and miR-155 expression,<sup>15</sup> capable of refining prognostication of older patients in this ELN Genetic Groups. We also observed a significant difference in OS between high and low DNMT3B expressers classified in the ELN Intermediate-I Genetic Group. Previously, RUNX1 mutations<sup>42</sup> and expression levels of MN1,<sup>12</sup> miR-155<sup>15</sup> and miR-3151<sup>16</sup> were demonstrated to add prognostic information in this ELN Genetic Group.

We report the first, to our knowledge, *DNMT3B*-associated gene and microRNA expression and DNA methylation profiles in CN-AML. We were able to derive a strong gene expression profile ----

Variable	Complete remission		Disease-free survival		Overall survival	
	OR (95% CI)	P-value	HR (95% CI)	P-value	HR (95% CI)	P-value
All patients						
DNMT3B expression, high vs low	0.49 (0.25–0.97)	0.04	1.47 (1.01–2.13)	0.04	1.72 (1.24–2.38)	0.001
BAALC expression, high vs low	0.21 (0.10–0.43)	< 0.001	1.79 (1.21–2.65)	0.004	1.98 (1.39–2.80)	< 0.001
WBC, per 50-unit increase	0.69 (0.52–0.90)	0.007				
Age, per 10-year increase DNMT3A <sup>a</sup>	0.49 (0.27–0.89)	0.02				
R882 mutated vs wild type			1.85 (1.17–2.95)	0.009		
non-R882 mutated vs wild type			0.92 (0.52-1.61)	0.76		
ELN Genetic Group, Favorable vs Intermediate-I <sup>b</sup>			0.65 (0.45-0.96)	0.03		
miR-3151 expression, high vs low					1.51 (1.07–2.13)	0.02
miR-155 expression, high vs low					1.47 (1.06–2.05)	0.02
Patients in the ELN modified Favorable Genetic Group	0					
DNMT3B expression, high vs low	No models includin	g a significa	nt term for DNMT3B	expression	1.99 (1.26–3.15)	0.003
BAALC expression, high vs low	were found	5 5			1.81 (1.13–2.92)	0.01
Patients in the ELN Intermediate L Canatis Groun <sup>b</sup>						
DNMT38 expression high vs low	No models includin	a a significa	at term for DNMT3B	evpression	1 73 (1 10_2 72)	0.02
BAALC expression, high vs low	were found	g a signinca		expression	1.88 (1.05_3.35)	0.02
miR-3151 expression high vs low	were lound				1.00(1.05-3.05) 1 79 (1 05-3 07)	0.03
WBC per 50-unit increase					$1.75(1.05 \ 5.07)$ 1.16(1.01 - 1.33)	0.03
wbc, per 50 unit increase					1.10 (1.01-1.55)	0.05
CR rate for the higher values of the continuous variable nigher (lower) risk of an event for higher values of continuous variable niclusion in the multivariable models if they had a univariable analyses. As <i>NPM1</i> , <i>FLT3</i> -ITD and <i>CEBPA</i> mindividual variables. In the entire patient cohort, variable <i>miR-3151</i> expression, ELN Genetic Groups, <i>WT1</i> and <i>ASX DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, ELN Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, <i>BLN</i> Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>miR-3151</i> expression, <i>BLN</i> Genetic or <i>DNMT3B</i> , <i>ERG</i> , <i>BAALC</i> and <i>DNMT3B</i> , <i>BAALC</i> and <i>DNMT3B</i> , <i></i>	Accordinated; FIK, hazard se and the first categor nuous variables and the variable <i>P</i> -value of $\leq 0$ utations are integrated se considered for inclus <i>L1</i> mutation status, WI tic Groups, <i>FLT3</i> -TKD, <i>A</i> <i>IALC</i> , mi <i>R</i> -155 and mi <i>R</i> - illary involvement. For	y listed for the first catego 20. See the S d in the ELN sion in the mo 3C, age and e <i>SXL1, DNMT3/</i> 3151 expressi patients in the	as ratio; WBC, White Bi he categorical variable ory listed of a dichotor Supplementary Inform genetic classification odel for achievement of extramedullary involve A-R882 and DNMT3A no on, ELN Genetic Grou he ELN modified Favor	s. A hazard ra nous variable lation for a fu , they were of CR were <i>DI</i> ement. In the on-R882 muta ps, <i>MLL</i> -PTD, able Genetic	atio > 1 (< 1) corres e. Variables were con ull list of variables er not additionally cor VMT3B, ERG, BAALC, n model for DFS, we ation status and extr. WT1, ASXL1, DNMT3, Group, variables con	ponds to sidered fo valuated i niR-155 an considere amedullar A-R882 an sidered fo

RUNX1, IDH1, DNMT3A-R882 and DNMT3A non-R882 mutation status, and WBC and hemoglobin. <sup>a</sup>The types of DNMT3A mutations detected in our cohort are provided in Supplementary Table S2. <sup>b</sup>The ELN modified Favorable Genetic Group is defined as CN-AML patients with mutated CEBPA and/or mutated NPM1 without FLT3-ITD. All remaining CN-AML patients (that is, those with wild-type CEBPA and wild-type NPM1 with or without FLT3-ITD, or mutated NPM1 with FLT3-ITD) belong to the ELN Intermediate-I Genetic Group.44

comprising 363 genes by overlapping the microarray results from three independent sets of patients. The profile was quite heterogeneous, comprising genes encoding for proteins involved in multiple biologic processes that play a role in leukemia cell differentiation, proliferation and survival. Among the downregulated genes, we noted enrichment of genes involved in monocyte/macrophage differentiation and activity, suggesting a role of *DNMT3B* in impairing differentiation of the leukemic blasts into cells with normal innate immunity activity. Using gene set enrichment analysis, we found a significant correlation between the high DNMT3B expression and FLT3-ITD signatures (Supplementary Figure S2). This, along with the increased frequency of FLT3-ITD in high DNMT3B expressers (Table 1), suggests the existence of a functional association between high expression of the DNMT3B gene and FLT3-ITD.

In contrast, the DNMT3B-associated microRNA profile was relatively weak, comprising only four microRNAs that were differentially expressed in high versus low DNMT3B expressers. Nevertheless, the unique upregulation of expression of miR-133b, recently reported to target GM-CSF<sup>50</sup> in DNMT3B high expressers was somewhat consistent with the enrichment of the gene expression profile in multiple downregulated genes involved in the differentiation and activity of hematopoietic cells participating in innate immunity.

Surprisingly, despite the fact that DNMT3B encodes a DNA methyltransferase, we observed no significant association of high

DNMT3B levels and DNA methylation changes. No difference in global DNA methylation levels and number of differentially methylated regions could be identified between DNMT3B high and low expressers. Our results are reminiscent of a recent report showing that changes in DNMT3B expression did not affect methylation levels of putative DNMT3B target genes.<sup>59</sup> Moreover, Russler-Germain et al.60 have recently demonstrated that DNA methylation levels in leukemic blasts from CN-AML patients are not influenced by DNMT3B expression as mainly inactive splice variants of DNMT3B are expressed in these cells. Overall, therefore, these data may suggest that although overexpressed DNMT3B is a potentially valuable predictive marker for response to conventional chemotherapy, it does not necessarily identify subsets of older AML patients characterized by aberrant DNA methylation who might be responsive to hypomethylating azanucleosides.

In summary, we have demonstrated that DNMT3B expression constitutes an independent prognostic factor in older CN-AML patients treated intensively, and could also refine the ELN classification. Furthermore, we have provided some insights into the biologic activity of DNMT3B in CN-AML, which is seemingly independent from mechanisms of DNA hypermethylation and/or microRNA-dependent gene repression. Further studies focused on gaining more clinical and mechanistic insights into the leukemogenic role of DNMT3B expression are

а С 10 1.0 P=0.10 by log rank test P=0.24 by log rank test **Disease-Free Survival** 0.8 Disease-Free Survival 0.8 0.6 0.6 0.4 0.4 \_ow DNMT3B (n=47) Low DNMT3B (n=33) 0.2 0.2 High DNMT3B (n High DNMT3B (n=27 0.0 0.0 0 2 3 5 0 2 4 1 3 Years Years b d 1.0 1.0 P=0.002 by log rank test P=0.02 by log rank test 0.8 0.8 **Overall Survival Overall Survival** 0.6 0.6 DNMT3B (n=56) 0.4 0.4 0.2 0.2 ow DNMT3B (n=45) High DNMT3B (n=38 High DNMT3B (n 0.0 0.0 0 2 0 2 3 5 3 1 4 1 4 5 Years Years

**Figure 2.** Clinical outcome of CN-AML patients with high and low *DNMT3B* expression classified into ELN Genetic Groups. Kaplan–Meier survival curves for (**a**) disease-free survival and (**b**) overall survival of patients in the ELN modified Favorable Genetic Group; (**c**) disease-free survival and (**d**) overall survival of patients in the ELN Intermediate-I Genetic Group. *P*-values presented are from the log rank test.

Biologic process or cellular component	Number of genes	Benjamini P-value
Associations with genes upregulated in the high DNMT3B expression group		
GOTERM RP: nucleotide biosynthetic process	10	0.098
GOTERM BP: nucleobase, nucleoside and nucleotide biosynthetic process	10	0.066
GOTERM_BP: nucleobase, nucleoside, nucleotide and nucleic acid biosynthetic process	10	0.066
Associations with genes downregulated in the high DNMT3B expression group Annotation Cluster 1		
GOTERM CC: vacuole	12	0.0052
GOTERM_CC: lytic vacuole	10	0.012
GOTERM_CC: lysosome	10	0.012
Annotation Cluster 2		
GOTERM_BP: endocytosis	11	0.0091
GOTERM_BP: membrane invagination	11	0.0091
GOTERM_BP: membrane organization	12	0.089
Annotation Cluster 3		
GOTERM_CC: intrinsic to membrane	74	0.028

warranted to design active therapeutic strategies for high *DNMT3B* expressers in CN-AML.

#### **CONFLICT OF INTEREST**

The authors declare no conflict of interest.

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# **AUTHOR CONTRIBUTIONS**

CN, JK, SV, KMr, GM and CDB designed the study, analyzed the data and wrote the manuscript, and all authors agreed on the final version; SPW, KHM, A-KE, PY, DF, HB, SS, JHM, JPC, Y-ZW, and RB carried out laboratory-based research; JK, KMa, SV and DN performed statistical analyses; and AJC, MRB, BLP, JEK, JOM, THC, RAL, RMS, KMr, GM and CDB were involved directly or indirectly in the care of patients and/or sample procurement.

#### REFERENCES

- 1 Mrózek K, Heerema NA, Bloomfield CD. Cytogenetics in acute leukemia. Blood Rev 2004; 18: 115–136.
- 2 Mrózek K, Marcucci G, Paschka P, Whitman SP, Bloomfield CD. Clinical relevance of mutations and gene-expression changes in adult acute myeloid leukemia with normal cytogenetics: are we ready for a prognostically prioritized molecular classification?. *Blood* 2007; **109**: 431–448.
- 3 Walker A, Marcucci G. Molecular prognostic factors in cytogenetically normal acute myeloid leukemia. *Expert Rev Hematol* 2012; **5**: 547–558.
- 4 Whitman SP, Maharry K, Radmacher MD, Becker H, Mrózek K, Margeson D *et al. FLT3* internal tandem duplication associates with adverse outcome and gene- and microRNA-expression signatures in patients 60 years of age or older with primary cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. *Blood* 2010; **116**: 3622–3626.
- 5 Becker H, Marcucci G, Maharry K, Radmacher MD, Mrózek K, Margeson D et al. Favorable prognostic impact of *NPM1* mutations in older patients with cytogenetically normal *de novo* acute myeloid leukemia and associated gene- and microRNA-expression signatures: a Cancer and Leukemia Group B study. *J Clin Oncol* 2010; **28**: 596–604.
- 6 Becker H, Marcucci G, Maharry K, Radmacher MD, Mrózek K, Margeson D et al. Mutations of the Wilms tumor 1 gene (WT1) in older patients with primary cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. Blood 2010; **116**: 788–792.
- 7 Marcucci G, Maharry K, Wu Y-Z, Radmacher MD, Mrózek K, Margeson D et al. IDH1 and IDH2 gene mutations identify novel molecular subsets within de novo cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. J Clin Oncol 2010; 28: 2348–2355.
- 8 Metzeler KH, Becker H, Maharry K, Radmacher MD, Kohlschmidt J, Mrózek K *et al. ASXL1* mutations identify a high-risk subgroup of older patients with primary cytogenetically normal AML within the ELN Favorable genetic category. *Blood* 2011; **118**: 6920–6929.
- 9 Marcucci G, Metzeler KH, Schwind S, Becker H, Maharry K, Mrózek K et al. Age-related prognostic impact of different types of DNMT3A mutations in adults with primary cytogenetically normal acute myeloid leukemia. J Clin Oncol 2012; 30: 742–750.
- 10 Schwind S, Marcucci G, Maharry K, Radmacher MD, Mrózek K, Holland KB et al. BAALC and ERG expression levels are associated with outcome and distinct gene and microRNA expression profiles in older patients with *de novo* cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. Blood 2010; **116**: 5660–5669.
- 11 Heuser M, Beutel G, Krauter J, Döhner K, von Neuhoff N, Schlegelberger B et al. High meningioma 1 (MN1) expression as a predictor for poor outcome in acute myeloid leukemia with normal cytogenetics. Blood 2006; **108**: 3898–3905.
- 12 Schwind S, Marcucci G, Kohlschmidt J, Radmacher MD, Mrózek K, Maharry K et al. Low expression of MN1 associates with better treatment response in older patients with de novo cytogenetically normal acute myeloid leukemia. Blood 2011; 118: 4188–4198.
- 13 Marcucci G, Mrózek K, Radmacher MD, Garzon R, Bloomfield CD. The prognostic and functional role of microRNAs in acute myeloid leukemia. *Blood* 2011; **117**: 1121–1129.
- 14 Schwind S, Maharry K, Radmacher MD, Mrózek K, Holland KB, Margeson D et al. Prognostic significance of expression of a single microRNA, miR-181a, in cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. J Clin Oncol 2010; 28: 5257–5264.
- 15 Marcucci G, Maharry KS, Metzeler KH, Volinia S, Wu Y-Z, Mrózek K et al. Clinical role of microRNAs in cytogenetically normal acute myeloid leukemia: miR-155 upregulation independently identifies high-risk patients. J Clin Oncol 2013; 31: 2086–2093.
- 16 Eisfeld AK, Marcucci G, Maharry K, Schwind S, Radmacher MD, Nicolet D *et al. miR-3151* interplays with its host gene *BAALC* and independently affects outcome of patients with cytogenetically normal acute myeloid leukemia. *Blood* 2012; **120**: 249–258.
- 17 Marcucci G, Yan P, Maharry K, Frankhouser D, Nicolet D, Metzeler KH et al. Epigenetics meets genetics in acute myeloid leukemia: clinical impact of a novel seven-gene score. J Clin Oncol 2014; 32: 548–556.

- 18 Yan P, Frankhouser D, Murphy M, Tam HH, Rodriguez B, Curfman J et al. Genome-wide methylation profiling in decitabine-treated patients with acute myeloid leukemia. *Blood* 2012; **120**: 2466–2474.
- 19 Renneville A, Boissel N, Nibourel O, Berthon C, Helevaut N, Gardin C et al. Prognostic significance of DNA methyltransferase 3A mutations in cytogenetically normal acute myeloid leukemia: a study by the Acute Leukemia French Association. Leukemia 2012; 26: 1247–1254.
- 20 Hervouet E, Vallette FM, Cartron PF. Dnmt3/transcription factor interactions as crucial players in targeted DNA methylation. *Epigenetics* 2009; **4**: 487–499.
- 21 Trowbridge JJ, Sinha AU, Zhu N, Li M, Armstrong SA, Orkin SH. Haploinsufficiency of *Dnmt1* impairs leukemia stem cell function through derepression of bivalent chromatin domains. *Genes Dev* 2012; **26**: 344–349.
- 22 Li KK, Luo LF, Shen Y, Xu J, Chen Z, Chen SJ. DNA methyltransferases in hematologic malignancies. *Semin Hematol* 2013; **50**: 48–60.
- 23 Wang J, Walsh G, Liu DD, Lee JJ, Mao L. Expression of ΔDNMT3B variants and its association with promoter methylation of *p16* and *RASSF1A* in primary non-small cell lung cancer. *Cancer Res* 2006; **66**: 8361–8366.
- 24 Hlady RA, Novakova S, Opavska J, Klinkebiel D, Peters SL, Bies J *et al.* Loss of Dnmt3b function upregulates the tumor modifier Ment and accelerates mouse lymphomagenesis. *J Clin Invest* 2012; **122**: 163–177.
- 25 Cancer Genome Atlas Research Network Genomic and epigenomic landscapes of adult *de novo* acute myeloid leukemia. *New Engl J Med* 2013; 368: 2059–2074.
- 26 Hayette S, Thomas X, Jallades L, Chabane K, Charlot C, Tigaud I et al. High DNA methyltransferase DNMT3B levels: a poor prognostic marker in acute myeloid leukemia. PLoS One 2012; 7: e51527.
- 27 Blum W, Garzon R, Klisovic RB, Schwind S, Walker A, Geyer S et al. Clinical response and miR-29b predictive significance in older AML patients treated with a 10-day schedule of decitabine. Proc Natl Acad Sci USA 2010; 107: 7473–7478.
- 28 Mayer RJ, Davis RB, Schiffer CA, Berg DT, Powell BL, Schulman P *et al.* Intensive postremission chemotherapy in adults with acute myeloid leukemia. *N Engl J Med* 1994; **331**: 896–903.
- 29 Stone RM, Berg DT, George SL, Dodge RK, Paciucci PA, Schulman P et al. Granulocyte-macrophage colony-stimulating factor after initial chemotherapy for elderly patients with primary acute myelogenous leukemia. N Engl J Med 1995; 332: 1671–1677.
- 30 Lee EJ, George SL, Caligiuri M, Szatrowski TP, Powell BL, Lemke S et al. Parallel phase I studies of daunorubicin given with cytarabine and etoposide with or without the multidrug resistance modulator PSC-833 in previously untreated patients 60 years of age or older with acute myeloid leukemia: results of Cancer and Leukemia Group B study 9420. J Clin Oncol 1999; 17: 2831–2839.
- 31 Baer MR, George SL, Sanford BL, Mrózek K, Kolitz JE, Moore JO et al. Escalation of daunorubicin and addition of etoposide in the ADE regimen in acute myeloid leukemia patients aged 60 years and older: Cancer and Leukemia Group B study 9720. Leukemia 2011; 25: 800–807.
- 32 Marcucci G, Moser B, Blum W, Stock W, Wetzler M, Kolitz JE *et al.* A phase III randomized trial of intensive induction and consolidation chemotherapy ± oblimersen, a pro-apoptotic Bcl-2 antisense oligonucleotide in untreated acute myeloid leukemia patients >60 years old. *J Clin Oncol* 2007; **25** (suppl): 360s (abstract 7012).
- 33 Mrózek K, Carroll AJ, Maharry K, Rao KW, Patil SR, Pettenati MJ *et al*. Central review of cytogenetics is necessary for cooperative group correlative and clinical studies of adult acute leukemia: the Cancer and Leukemia Group B experience. *Int J Oncol* 2008; **33**: 239–244.
- 34 Payton JE, Grieselhuber NR, Chang LW, Murakami M, Geiss GK, Link DC et al. High throughput digital quantification of mRNA abundance in primary human acute myeloid leukemia samples. J Clin Invest 2009; 119: 1714–1726.
- 35 Whitman SP, Archer KJ, Feng L, Baldus C, Becknell B, Carlson BD et al. Absence of the wild-type allele predicts poor prognosis in adult *de novo* acute myeloid leukemia with normal cytogenetics and the internal tandem duplication of *FLT3*: a Cancer and Leukemia Group B study. *Cancer Res* 2001; **61**: 7233–7239.
- 36 Thiede C, Steudel C, Mohr B, Schaich M, Schäkel U, Platzbecker U et al. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. Blood 2002; 99: 4326–4335.
- 37 Yamamoto Y, Kiyoi H, Nakano Y, Suzuki R, Kodera Y, Miyawaki S et al. Activating mutation of D835 within the activation loop of FLT3 in human hematologic malignancies. Blood 2001; 97: 2434–2439.
- 38 Whitman SP, Ruppert AS, Marcucci G, Mrózek K, Paschka P, Langer C et al. Long-term disease-free survivors with cytogenetically normal acute myeloid leukemia and *MLL* partial tandem duplication: a Cancer and Leukemia Group B study. *Blood* 2007; **109**: 5164–5167.
- 39 Marcucci G, Maharry K, Radmacher MD, Mrózek K, Vukosavljevic T, Paschka P et al. Prognostic significance of, and gene and microRNA expression signatures associated with, CEBPA mutations in cytogenetically normal acute myeloid leukemia



with high-risk molecular features: a Cancer and Leukemia Group B study. J Clin Oncol 2008; **26**: 5078–5087.

- 40 Paschka P, Marcucci G, Ruppert AS, Whitman SP, Mrózek K, Maharry K *et al.* Wilms' tumor 1 gene mutations independently predict poor outcome in adults with cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. *J Clin Oncol* 2008; **26**: 4595–4602.
- 41 Metzeler KH, Maharry K, Radmacher MD, Mrózek K, Margeson D, Becker H et al. TET2 mutations improve the new European LeukemiaNet risk classification of acute myeloid leukemia: a Cancer and Leukemia Group B study. J Clin Oncol 2011; 29: 1373–1381.
- 42 Mendler JH, Maharry K, Radmacher MD, Mrózek K, Becker H, Metzeler KH et al. RUNX1 mutations are associated with poor outcome in younger and older patients with cytogenetically normal acute myeloid leukemia and with distinct gene and microRNA expression signatures. J Clin Oncol 2012; 30: 3109–3118.
- 43 Marcucci G, Radmacher MD, Maharry K, Mrózek K, Ruppert AS, Paschka P et al. MicroRNA expression in cytogenetically normal acute myeloid leukemia. N Engl J Med 2008; 358: 1919–1928.
- 44 Metzeler KH, Hummel M, Bloomfield CD, Spiekermann K, Braess J, Sauerland MC *et al.* An 86-probe-set gene-expression signature predicts survival in cytogenetically normal acute myeloid leukemia. *Blood* 2008; **112**: 4193–4201.
- 45 Huang da W, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 2009; 4: 44–57.
- 46 Vittinghoff E, Glidden DV, Shiboski SC, McCulloch CE. Regression Methods in Biostatistics: Linear, Logistic, Survival and Repeated Measures Models. Springer: New York, NY, USA, 2005.
- 47 Döhner H, Estey EH, Amadori S, Appelbaum FR, Büchner T, Burnett AK *et al.* Diagnosis and management of acute myeloid leukemia in adults: recommendations from an international expert panel, on behalf of the European LeukemiaNet. *Blood* 2010; **115**: 453–474.
- 48 Zhao H, Li M, Li L, Yang X, Lan G, Zhang Y. MiR-133b is down-regulated in human osteosarcoma and inhibits osteosarcoma cells proliferation, migration and invasion, and promotes apoptosis. *PLoS One* 2013; 8: e83571.
- 49 Duan F-T, Qian F, Fang K, Lin K-Y, Wang W-T, Chen Y-Q. miR-133b, a muscle-specific microRNA, is a novel prognostic marker that participates in the progression of human colorectal cancer via regulation of CXCR4 expression. *Mol Cancer* 2013; **12**: 164.

- 50 Sturrock A, Mir-Kasimov M, Baker J, Rowley J, Paine R 3rd. Key role of microRNA in the regulation of granulocyte macrophage colony-stimulating factor expression in murine alveolar epithelial cells during oxidative stress. J Biol Chem 2014; 289: 4095–4105.
- 51 Duursma AM, Kedde M, Schrier M, le Sage C, Agami R. miR-148 targets human DNMT3b protein coding region. *RNA* 2008; **14**: 872–877.
- 52 Zhu A, Xia J, Zuo J, Jin S, Zhou H, Yao L *et al.* MicroRNA-148a is silenced by hypermethylation and interacts with DNA methyltransferase 1 in gastric cancer. *Med Oncol* 2012; **29**: 2701–2709.
- 53 Hsu SH, Wang B, Kota J, Yu J, Costinean S, Kutay H *et al.* Essential metabolic, anti-inflammatory, and anti-tumorigenic functions of miR-122 in liver. *J Clin Invest* 2012; **122**: 2871–2883.
- 54 Köberle V, Kronenberger B, Pleli T, Trojan J, Imelmann E, Peveling-Oberhag J *et al.* Serum microRNA-1 and microRNA-122 are prognostic markers in patients with hepatocellular carcinoma. *Eur J Cancer* 2013; **49**: 3442–3449.
- 55 Li A, Song W, Qian J, Li Y, He J, Zhang Q et al. MiR-122 modulates type I interferon expression through blocking suppressor of cytokine signaling 1. Int J Biochem Cell Biol 2013; 45: 858–865.
- 56 Zheng B, Liang L, Huang S, Zha R, Liu L, Jia D *et al.* MicroRNA-409 suppresses tumour cell invasion and metastasis by directly targeting radixin in gastric cancers. *Oncogene* 2012; **31**: 4509–4516.
- 57 Mrózek K, Marcucci G, Nicolet D, Maharry KS, Becker H, Whitman SP *et al.* Prognostic significance of the European LeukemiaNet standardized system for reporting cytogenetic and molecular alterations in adults with acute myeloid leukemia. *J Clin Oncol* 2012; **30**: 4515–4523.
- 58 Gaidzik VI, Schlenk RF, Paschka P, Stölzle A, Späth D, Kuendgen A et al. Clinical impact of DNMT3A mutations in younger adult patients with acute myeloid leukemia: results of the AML Study Group (AMLSG). Blood 2013; 121: 4769–4777.
- 59 Hagemann S, Kuck D, Stresemann C, Prinz F, Brueckner B, Mund C *et al.* Antiproliferative effects of DNA methyltransferase 3B depletion are not associated with DNA demethylation. *PLoS One* 2012; **7**: e36125.
- 60 Russler-Germain DA, Spencer DH, Young MA, Lamprecht TL, Miller CA, Fulton R *et al.* The R882H DNMT3A mutation associated with AML dominantly inhibits wild-type DNMT3A by blocking its ability to form active tetramers. *Cancer Cell* 2014; **25**: 442–454.

Supplementary Information accompanies this paper on the Leukemia website (http://www.nature.com/leu)