



Current Medical Research and Opinion

ISSN: 0300-7995 (Print) 1473-4877 (Online) Journal homepage: https://www.tandfonline.com/loi/icmo20

Environmental toxins exposure in PCOS women and possible ovarian neoplastic repercussion

Ilaria Soave, Tommaso Occhiali, Chiara Assorgi, Roberto Marci & Donatella Caserta

To cite this article: Ilaria Soave, Tommaso Occhiali, Chiara Assorgi, Roberto Marci & Donatella Caserta (2020): Environmental toxins exposure in PCOS women and possible ovarian neoplastic repercussion, Current Medical Research and Opinion, DOI: <u>10.1080/03007995.2020.1729108</u>

To link to this article: https://doi.org/10.1080/03007995.2020.1729108



Accepted author version posted online: 11 Feb 2020.



🖉 Submit your article to this journal 🗹



View related articles $oldsymbol{C}$



View Crossmark data 🗹

Environmental toxins exposure in PCOS women and possible ovarian neoplastic repercussion

Soave Ilaria¹, Occhiali Tommaso², Assorgi Chiara¹, Marci Roberto², Caserta Donatella¹

- ¹ Department of Surgical and Clinical Sciences and Translational Medicine, S. Andrea Hospital, "Sapienza" University of Rome, Italy
- ² Department of Morphology, Surgery and Experimental Medicine, University of Ferrara, Italy

Correspondence: Soave Ilaria, Department of Surgical and Medical Sciences and Translational Medicine, "Sapienza" University of Rome, S. Andrea Hospital, via di Grottarossa 1035 Rome, Italy; email: ilaria.soave@uniroma1.it

Transparency Declaration of funding This manuscript was not funded.

Declaration of financial/other relationships

cè

IS, TO, CA, RM, and DC have disclosed that they have no significant relationships with, or financial interests in, any commercial companies related to this article. Peer reviewers on this manuscript have no relevant financial or other relationships to disclose.

Acknowledgements

None reported.

Abstract

Purpose: Over the last two decades, increasing attention has been paid to environmental toxins and their effects on the female reproductive system. Endocrine disrupting chemicals (EDCs) are exogenous substances or mixtures that can mimic the action of steroid hormones and interfere with their metabolism. Advanced glycation end products (AGEs) are proinflammatory molecules that can interact with cell surface receptors and mediate the triggering of proinflammatory pathways and oxidative stress. The purpose of this review is to explore the effects of environmental toxins exposure in the pathogenesis of both polycystic ovary syndrome (PCOS) and OC (ovarian cancer), considered separately, and also to evaluate possible neoplastic ovarian repercussion after exposure in patients diagnosed with PCOS.

Materials and methods: We searched PubMed for articles published in the English language with the use of the following MeSH search terms: "Polycystic Ovary Syndrome" and "Ovarian Cancer" combined with "endocrine disruptors". Titles and abstracts were examined and full articles that met the selection criteria were retrieved. A manual search of review articles and cross-references completed the search.

Results: Extensive data from different studies collected in recent years concerning the effects of EDCs/AGEs exposure have confirmed their role in the pathophysiology of both PCOS and OC. They favour PCOS/OC development through different mechanisms that finally lead to hormonal and metabolic disruption and epigenetic modifications.

Conclusions: Environmental toxin exposure in PCOS women could favour neoplastic transformation by exacerbating and potentiating some PCOS features. Further research, although difficult, is needed in order to prevent further diffusion of these substances in the environment, or at least to provide adequate information to the population considered at risk.

Keywords: polycystic ovary syndrome; ovarian cancer; endocrine disrupting chemicals; advanced glycation end products

ce

BACKGROUND

Endocrine Disrupting Chemicals and Advanced Glycation End products

Endocrine disrupting chemicals (EDCs) are defined as "exogenous substances or mixtures that alter function(s) of the endocrine system and consequently cause adverse effects in an intact organisms or its progeny or (sub)population"¹. Also known as endocrine disruptors, they can mimic the action of steroid hormones and "interfere with the synthesis, secretion, transport, metabolism, binding action or elimination of natural blood-born hormones that are present in the body and are responsible for homeostasis, reproduction and developmental process"². They usually possess a phenol group and express affinity for hormone receptors (e.g. oestrogen, progestin, androgen), although lower than the natural ligand. They may also interfere with the synthesis of carrier proteins, such as sex-hormone binding globulin (SHGB), and alter the transport of steroid hormones to target cells^{3,4}. Over the last two decades, increasing attention has been paid to EDCs and their effects on the female reproductive system. More than 800 chemicals have been classified as EDCs, including drugs (diethylstilbestrol (DES)), pesticides

(dichlorodiphenyltrichloroethane (DTT), atrazine, vinclozin), polychlorinated biphenyls (PBCs), dioxin and dioxin-like compounds, plasticizers (bisphenol A (BPA), bisphenol F, bisphenol S, phthalates), heavy metals (lead, arsenic, aluminum, cadmium) and the list is growing rapidly⁵. These chemicals are present in common industrial and home products and possess a high potential for bioaccumulation in the food web⁶. Therefore, humans are continually exposed to a mixture of EDCs and the contact with them has become inevitable today^{1,7}. In addition, globalisation and consumerism have amplified the phenomenon. Heat-processed food and highly processed products represent an important source of advanced glycation end products (AGEs)⁸. Also called glycotoxins, AGEs are proinflammatory molecules that can interact with cell surface receptors (receptors for AGEs, RAGE) and mediate the triggering of proinflammatory pathways and oxidative stress^{9,10}. They are believed to take part in atherosclerosis and diabetes pathogenesis, female fertility disruption and carcinogenesis^{8,11-13}.

The exact mechanism of action by which EDCs interfere with the female reproductive system is not yet fully understood. However, it is well established that through binding to nuclear hormone receptors, with agonistic or antagonistic effect, they may activate a rapid downstream intracellular signalling, resulting transcription errors (*genomic pathway*)¹⁴. They primarily alter the oestrogen (E) signalling pathway, probably because it is evolutionarily conserved among animals and regulates many functions of the female reproductive system¹⁵. They may also act through binding to membrane steroids hormone receptors or G protein-coupled receptor 30 (GPR30), leading to protein kinase activation and recruitment of second messengers, which interfere with synthesis, secretion, transport and metabolism of endogenous hormones (*non-genomic pathway*)¹⁶. Recently, other mechanisms have been taken into account, including oxidative stress¹⁷ and epigenetic effects^{18,19}.

Polycystic Ovarian Syndrome

Polycystic Ovarian Syndrome (PCOS) is a common and complex endocrinopathy that affect women of reproductive age²⁰. PCOS affects approximately 5 to 10% of the general female population²¹. Depending on the diagnostic criteria and the geographic location this percentage may rise up to 21%²². PCOS was redefined in 2003 in a consensus meeting between the European Society of Human Reproduction and Embryology and the American Society of Reproductive Medicine (ESHRE/ASRM)²³. Affected individuals must meet at least two of the following criteria (Rotterdam criteria):

- (i) clinical and/or biochemical hyperandrogenism
- (ii) oligo-/anovulation
- (iii) polycystic ovaries.

The pathophysiology of this endocrine disorder is still unclear, however a genetic basis (multifactorial and polygenic) is suspected²⁴. Some authors have suggested a possible autosomal dominant inheritance, based on the higher prevalence of increased androgens (A) levels and insulin-resistance in relatives of PCOS women²⁵. In addition a possible role of environmental factors and lifestyle has also been taken into account²⁴. Key features of PCOS are hyperandrogenism and insulin resistance, that exhibit in various degree. Hyperandrogenism contributes to clinical phenotypes and fertility dysregulation. The most

common clinical signs include hirsutism, acne, menstrual cycle dysfunction (oligo-/amenorrhea, unpredictable bleeding) and subfertility/infertility^{20,26}. Amenorrhea and oligomenorrhea result from anovulation. The lack of endogenous progesterone (P), generally produced by the corpus luteum after ovulation, impedes the triggering of a normal menstrual cycle. In addition, elevated A levels counteract the proliferative effect of E on the endometrium, resulting in a thin endometrial stripe. Insulin resistance (IR) and compensatory hyperinsulinemia are present in the majority of PCOS women²⁷. Elevated insulin levels stimulates A production (further promoting anovulation) and may also favour the development of obesity²⁸. However, given the fact that obesity itself, in particular central obesity, exacerbates IR, its prevalence increases with the rate of obesity²⁸. The metabolic profile of PCOS women is similar to that seen in the metabolic syndrome and includes atherogenic dyslipidemia (increased LDL-cholesterol and triglycerides levels, depressed HDL-cholesterol levels and elevated total cholesterol:HDL-cholesterol ratios) and impaired glucose tolerance. Consequently, patients with PCOS are more likely to develop type 2 diabetes (T2DM)²⁹ and cardiovascular disease (CVD)³⁰ when compared to the general population.

Ovarian Cancer

Worldwide, ovarian cancer (OC) is the 7th leading cause of cancer in females³¹ and the 1st cause of death from gynaecological malignancies³² and its incidence is continuously rising both in Western countries and Asian nations^{31,33}. Postmenopausal women are more likely to be diagnosed with OC. Although the pathophysiology is poorly understood, several factors including genetic, reproductive, hormonal and behavioural factors, may contribute to the tumorigenic process³⁴. Evidence suggests that hormonal imbalance plays a key role in OC development. OC cells express higher levels of oestrogen receptor alpha (ERa) when compared to normal ovarian cells, suggesting that E may provide a favourable hormonal milieu for tumour progression by directly regulating cellular growth and proliferation³⁵. Recent advances in molecular techniques have led to consider OC as a complex group of distinct diseases sharing a common anatomical location³⁶. In 90% of the cases, OCs arise from the neoplastic transformation of epithelial cells on the ovarian surface³⁷. Epithelial ovarian tumours are further divided into type 1 and type 2 subgroups³⁸. Type 1 OCs are generally low-grade and well differentiated and include low-grade serous and mucinous carcinoma, endometrioid carcinoma, clear-cell carcinoma and transitional carcinoma. They are usually ovarian derived or may arise from pre-invasive endometriotic lesions. On the other hand, type 2 OCs are high-grade, undifferentiated and metastasized tumours and account for the 75% of all OC diagnoses. This subgroup includes high-grade serous adenocarcinoma, high-grade endometrioid-carcinoma, carcinosarcoma and undifferentiated carcinoma^{39,40}. OC is usually considered a sporadic disease, but in 10 to 15% of the cases it shows hereditary etiology⁴¹. The genetic profiling of OC varies among the two subgroups. Type 1 OCs are usually characterised by genomic stability and frequently exhibit point mutations⁴². Type 2 OCs are usually the result of multiple mutational events that mainly involve p53 genes. The reduced activity of the tumour suppressor protein 53 (TP53) leads to uncontrolled cellular growth and division, favouring the development of cancer⁴³. In addition, constitutive mutations in breast cancer antigen (BRCA) 1 and 2 genes are also often associated with type 2 OCs. Indeed, women with a BRCA1 gene mutation have 40% to 60% lifetime risk of developing OC, whereas the risk for women with BRCA 2 gene mutation is 20% to 35%⁴⁴.

MATERIAL AND METHODS

We search PubMed for articles published in the English language with the use of the following MeSH search terms: "Polycystic Ovary Syndrome" and "Ovarian Cancer" combined with "endocrine disruptors". Titles and abstracts were examined by two reviewers (I.S. and T.O.) and full articles that met the selection criteria were retrieved. A manual search of review articles and cross-references completed the search. All selected articles were assessed for study design, patient characteristics, diagnosis of PCOS/OC, type of EDC studied and completeness of information of the data sets.

ENVIRONMENTAL TOXINS EXPOSURE AND PCOS

Increased levels of BPA in biological fluids have been reported in both women and adolescents with PCOS when compared to healthy, non hyperandrogenic and ovulating controls⁴⁵⁻⁴⁷. Higher serum levels of other EDCs (PBCs, organochlorine pesticides, perfluorooctanoate, perfluorooctane sulfonate, polycyclic aromatic

hydrocarbons)^{48,49} and AGEs have also been observed in PCOS patients⁵⁰. In one study elevated circulating serum levels of AGEs were positively associated with higher serum concentrations of antimüllerian hormone (AMH), testosterone (T) and insulin⁵¹. The link between environmental toxins exposure and PCOS may lay in the interaction between EDCs/AGEs and sex hormones, but human data are still limited⁶. The molecular similarity of environmental toxins to endogenous steroids hormones may favour the development of PCOS through different mechanisms that could finally lead to hormonal and metabolic disruption (Figure 1).

Hormonal disruption

EDCs may contribute to the destabilisation of hormonal homeostasis both by acting directly at gonadal level (*direct effect*) or via alteration of the hypothalamic-pituitary-ovarian axis (*indirect effect*)⁵². At ovarian level, possible effects on both granulosa and theca cells have been hypothesized. Elevated levels of BPA in follicular fluid may be responsible for decreased expression of aromatase and reduced production of E in granulosa cells, finally disrupting the intrafollicular environment and affecting oocytes growth and maturation⁵³. Similar effects have also been shown in case of exposure to other popular plasticizerphthalates⁵⁴. The impact of BPA on oogenesis may be modulated by the interaction with a variety of receptors, including ER α and β^{55} , nonclassical membrane ER and GPR30⁵⁶. In theca cells, BPA may stimulate A secretion⁵³ and may inhibit T clearance and catabolism⁵⁷, consequently increasing A circulating levels. In addition, BPA is a powerful SHBG ligand and it is able to displace T from SHBG, increasing free T serum levels⁵⁸. Moreover, the increased levels of A reduce the hepatic clearance of BPA by decreasing the activity of UDP glucuronyl-transferase liver enzyme, consequently increasing free BPA levels and therefore contributing to the perpetuation of BPA/A reciprocal relationship⁵⁹. The result of all these actions is the disruption of A-E balance, thus impairing oocytes maturation. EDCs may also be involved in the pathogenesis of anovulation by suppressing E production through a receptor-mediated signalling pathway (peroxisome proliferator-activated receptors, PPARy)⁶⁰. AGEs have also been reported to be able to impact oocytes development and maturation through different mechanisms of action: chronic inflammation and oxidative stress are though to play a pivotal role in AGE-mediated disruption of reproductive function in preovulatory follicles in PCOS women^{61,62}.

Metabolic disruption

As stated before, the metabolic profile of PCOS women is characterized by IR with compensatory hyperinsulinemia and obesity and environmental toxins may be involved in the pathophysiology of both of them. Indeed, BPA may promote the development of IR by reducing adiponectine secretion, resulting in increased oxidative stress, inflammation and disruption of adipogenesis, which favour the onset of both T2DM and CVD^{63,64}. In addition, it has also been observed that BPA has a direct effect on pancreatic cells, which contributes to the disruption of glucose homeostasis, leading to hyperinsulinemia, thus further increasing the risk of T2DM^{65,66}. In vitro studies have also suggested a possible involvement of AGEs in the pathogenesis of IR. Indeed, glycotoxins may reduce glucose uptake by adipocytes⁶⁷ and alter glucose transport in human cells⁶⁸. AGE-induced inflammation may also decrease insulin sensitivity⁶⁹. Moreover, it is well known that obesity itself worsens IR and central and visceral obesity are present in about 30-75% of PCOS women⁷⁰. A positive correlation between glycotoxins serum levels and waist-hip ratio has been reported⁶⁸ and in vitro studies support the role of AGEs in stimulating adipogenesis⁷¹. Moreover, it has also been observed that higher body mass index correlates with lower levels of soluble RAGE, thus reducing AGEs clearance and favouring their deposition in reproductive tissues (e.g. ovaries), possibly affecting steroidogenesis^{72,73}. In addition, increased AGEs serum levels potentiate the inflammatory process already present in obese patients and worsen the metabolic aberrations characteristic of PCOS women, thus aggravating the metabolic syndrome components of the disease⁷⁴. Furthermore, EDCs are often classified as "environmental obesogens". Indeed, they promote fat accumulation, by altering lipid homeostasis and fat storage⁷⁵. The molecular pathways by which these compounds participate in obesity development may include the interaction with both PPARy and ER^{55,76}, which may promote the expression of adipogenic genes⁷⁷ and may enhance adipocytes differentiation⁷⁸.

ENVIRONMENTAL TOXINS EXPOSURE AND OC

Several studies have given substantial evidence concerning the link between EDCs/AGEs and OC development (Figure 2). Human exposure to different EDCs (e.g. pesticides, triazine herbicides, organophosphate diazinon, triclosan, methoxychlor (MXC)) has been proven to increase the risk of OC⁷⁹⁻⁸³ and many studies conducted on animal models support this hypothesis. Mice exposed to BPA and 2,2-bis(p-hydroxyphenyl)-1,1,1-trichloroethane (HPTE) have shown an increased incidence of ovarian cysts and ovarian adenomas, along with an enhanced OC cells proliferation^{84,85}. Additionally, BPA has also been reported to interfere with sex steroid hormones synthesis, leading to hormonal imbalance⁸⁶.

Hormonal effects

The role of E in the development and progression of OC is still debated, but a possible involvement in the early stages of malignancy seems plausible. Several studies conducted on postmenopausal women have linked hormone replacement therapy to an increased risk of OC⁸⁷⁻⁸⁹ and recent studies have reported that both E and E+P preparations are equally associated with OC⁹⁰. Given the fact that some EDCs exhibit estrogenic properties, the environmental exposure may thus activate ER signalling, favouring cancer development⁹¹. Some in vitro studies support this hypothesis: plasticizers and pesticides have been shown to promote OC cells proliferation via E pathways activation^{83,92} and DDT via nuclear receptor binding⁹³. Also EDCs with antiestrogenic properties might disrupt the hormonal milieu. The perimenopausal phase is characterized by increased levels of gonadotropins (hypergonadotropic condition) that are able to activate different molecular pathways involved in cellular growth and invasion and EDCs exposure may enhance this process, further increasing gonadotropins levels and consequently the risk of OC⁹⁴. It has been reported that chlorpyrifos and MXC might be able to alter gonadotropin-releasing hormones production^{95,96} and that DDT might disturb the normal functioning of gonadotropins via the expression of FSH and LH⁹⁷.

The overexpression of androgen receptor in OC cells⁹⁸ suggests that A may also play a role in the tumorigenic process. Androgens are involved in the regulation of cells proliferation and, in particular, are able to decrease cell death⁹⁸. Some authors have depicted a possible link between the use of exogenous A and an increased risk of OC^{99,100} and some studies conducted on animal models have shown that the exposure to EDCs with androgenic properties (e.g. 4-Nitro-3-phenylphenol) is associated with hormonal disruption¹⁰¹.

Epigenetic effects

Long-term exposure to environmental toxins may also be associated with changes in gene expression (epigenetic modifications)¹⁰². Several epigenetic markers have been taken into account, including histone modifications, non coding RNAs transcription and changes in DNA methylation¹⁰³. The latter is the most studied and consists in the transfer of a methyl group(s) to a DNA molecule by DNA methyl transferases (DNMTs) activation¹⁰⁴. This process is essential for normal cellular development, but it is also believed to play a key role in carcinogenesis¹⁰⁵. Chemicals and environmental toxins may interfere with the methylation of CpG islands (DNA regions where a cytosine nucleotide is followed by guanine nucleotide, separated only by a phospahte group) which can cause permanent silencing or activation of particular genes¹⁰⁵. The epigenetic effect of EDCs in case of reproductive cancer has been observed^{106,107}, but only few studies have examined the EDC-related epigenetic changes in the ovary. It has been reported that EDCs may alter the expression of E responsive genes by promoting hypermethylation in the promoter region of hormone receptor genes, leading to gene silencing^{108,109}. In addition, studies on animal models have shown that high doses of MXC are associated with hypermethylation of ER **β**, DNA methyltransferase 3 **β** and pregnancy associated plasma protein A genes in rats¹¹⁰. Reduced expression of ER **β** gene and increased activity of DNMT gene under the influence of EDCs have also been observed¹⁰⁸.

POSSIBLE OVARIAN NEOPLASTIC REPERCUSSION IN PCOS WOMEN EXPOSED TO EDCs

The previous sections explored the effects of environmental toxins exposure in the pathogenesis of both PCOS and OC, considered separately. The current section pertains to the possible neoplastic ovarian repercussion after environmental toxins exposure in patients with PCOS.

The possible association between PCOS and increased cancer risk was first hypothesized in 1940¹¹¹ and, more recently, concerns that women with PCOS might be at increased risk of OC have been raised¹¹². PCOS has been postulated to promote OC development mainly through excessive A stimulation^{98,113}. Normal ovarian cells as well as borderline ovarian tumours cells express androgen receptors on their surface, and increased A levels positively correlate with an increased risk of borderline serous and invasive mucinous tumours^{114,115}. Only few studies examined the possible link between PCOS and OC with inconsistent results^{98,99,116-123}. Many of these studies had small cohort sizes and different diagnostic criteria for PCOS were used, limiting the power of the study and introducing selection bias. A recent review published by Harris et al. failed to identify a clear association between PCOS and OC and several potential confounders have been pointed out¹²⁶. Firstly, obesity is a common finding in PCOS women who are also, as stated before, at higher risk of T2DM, IR and metabolic syndrome^{27,29,125}. All these conditions have been associated with ovarian cancer^{126,127} and, in case of PCOS women, could act as potential intermediate variables. Few studies adjusted their results for these confounders, resulting in an attenuation of the true association¹²⁰. In addition, oral contraceptives are often a first-line treatment in PCOS women not seeking for pregnancy, which use seems to play a protective role in ovarian neoplasms development by decreasing the number of potentially damaging ovulations^{128,129}. Furthermore, metformin is frequently prescribed in PCOS patients in order to improve IR and reduce the risk of T2DM. Laboratory studies have advised a possible anti-cancer activity of metformin¹³⁰ and several studies have confirmed its influence on ovarian cells growth and proliferation^{131,132}. One study reported a decreased risk of OC in patients treated with metformin¹¹⁸. Age represents another possible confounding factor. Given the fact that postmenopausal women are more likely to be diagnosed with OC, in a recent meta-analysis published by Barry and colleagues only PCOS patients <54 years of age (premenopausal women) were considered and even in this case, no robust association between PCOS and OC was found¹¹⁹.

Although there may be several confounding factors, environmental toxins exposure in patients with PCOS may exacerbate some PCOS features (hormonal, metabolic, genetic) that in the end may favour the neoplastic process.

Hormonal pathways

Cells within the ovaries and on their surface are responsive to A, which have antiapoptotic effects and induce cells proliferation, thus promoting cell survivability, mutations' accumulation and, eventually, neoplastic transformation. Moreover, once ovarian cells turn malignant, they possess the enzymatic capabilities to synthesized A with greater biological effects and tend to overexpress androgen receptors¹³³⁻¹³⁵. Both PCOS and BPA synergically increase not only A levels, but also free A levels, since BPA manages to unbind T from SHBG⁵⁸. BPA plasmatic concentrations positively correlate with T, free T, androstenedione and DHEA levels in PCOS women^{45,46}. Considering the above-mentioned premises, this hormonal disruption can furthermore increase ovarian cancer growth both directly, acting on A receptors, and indirectly, providing weaker A to the tumour enzymatic machinery, which in turn will convert them into more biologically active ones^{52,136}. Additionally, elevated A levels impede the hepatic clearance capacity of BPA, creating a vicious circle that promotes A production and in the end tumour mass growth¹³⁷.

Oestrogens effects on ovarian cancer cells are somewhat less clearly defined, considering that OC is typically found in postmenopausal women. Still, both ER α and β are found in more than half of all ovarian tumours¹³⁸. Alfa receptors support cellular growth and are counterbalanced by β receptors. In time, ovarian malignant cells that only express α receptors are selected^{139,140}. PCOS patients are usually overweight/obese¹⁴¹. The hypertrophic fatty tissue is the production site of estrone, an E with low biological effects, but still a proliferative stimulus to ovarian cells. Additionally, the hyperandrogenic condition typical of PCOS provides other substrates to the aromatase located in adipocytes, favouring A conversion to E. EDCs in general can act either with estrogenic effect or with antiestrogenic effect, depending on the substance(s), on its dose and on possible synergic effects with other EDCs¹⁴². When the net effect tips toward the estrogenic effect, PCOS and EDCs could act directly toward premalignant and malignant cells proliferation by binding to ER, while when the net effect is rather antiestrogenic, we may occur in a negative feedback at hypothalamic level, with subsequent gonadotropins release, which, in turn,

will stimulate gonadal cells proliferation and hormones synthesis, still concurring in tumour proliferation^{92,93,142-144}.

Metabolic disruption

The low-grade systemic inflammation typical of the metabolic syndrome of PCOS women is bolstered by EDCs, as stated above. This meta-inflammation promotes tumour growth through, at least, two major mechanisms: oxidative stress and IR.

Oxidative stress, a known carcinogenetic stimulus, is a by-product of chronic metainflammation mainly sustained by TNF- α and IL-6, powerful cytokines produced by adipose tissue and its residing macrophages^{145,146}. TNF- α leads to lipolysis and subsequent IL-6 secretion. In the liver, IL-6 is responsible both for acute phase proteins synthesis (further promoting systemic inflammatory response) and, also, for IR, not only in hepatic cells but also in peripheral tissues (skeletal muscles cells). EDCs (namely organochlorines) increase adipogenesis and maturation of mesenchymal stem cells into adipocytes through growth factors expression (BMP, EGF, IGF-1), leading to deposition of fatty tissue, both visceral and peripheral^{147,148}. IR comes from EDCs-derived TNF- α activity, which impedes glucose uptake at cellular level by downregulating GLUT-4 transporters function¹⁴⁹. EDCs exposure (BPA and diethylhexyl phthalate) during gestation and during the immediate post-natal period, could cause reduced insulin secretion, hyperglicemia and pancreatic insulitis in adult life¹⁵⁰⁻¹⁵². In addition, EDCs alter the surrounding environment of pancreatic islets, adipose muscle and hepatic cells by affecting the extracellular matrix¹⁵³. Inflammation, deposition of connective tissue and angiogenesis are finally promoted, altering the blood flow in metabolically active tissues and causing chronic low grade ischemic damages, further stimulating the inflammatory response and decreasing hormonal secretion capability via a multitude of pathways (hormonal receptor, intracellular second messenger, growth factors)¹⁵⁴⁻¹⁵⁶. The result is an important disruption in the ability of insulin to exert its effects favouring hyperglycaemia and compensatory hyperinsulinemia. Hyperglycaemia leads to AGEs formation. Considering AGEs pro-inflammatory effects, we circle back to reactive oxygen species (ROS) and their oxidative stress, amplified not only as a local ischemic-derived damage to adipose tissue and skeletal muscle, but as systemic insults, especially in ovarian pre-malignant and malignant cells, where DNA strands get damaged, steroidogenesis is impaired and overall ovarian disfunction is found⁷⁴. AGEs alter insulin sensitivity and glucose uptake directly in granulosa cells⁶⁸ and in adipocytes⁶⁷. Finally, PCOS derived hyperandrogenism further impairs the body capacity of AGEs clearance, amplifying their effects. Hyperinsulinemia (derived from PCOS, EDCs and AGEs) promotes A production in the ovaries, thus providing a proliferative stimulus, also sustained by IGF-1, a molecule capable of sustaining cellular mitosis and of dampening cisplatin pro-apoptotic effects when neoplastic transformation has already taken place¹⁵⁷.

Epigenetics

EDCs can cause epigenetic modifications, especially during pre- and peri-natal exposition¹⁵⁸. These "subtle" epigenetic alterations accompany the individual throughout its life and can concur to carcinogenesis. The data available on epigenetic changes and ovarian cancer mainly show that ER are involved, particularly downregulating suppressor receptors (ER β), through gene promoter hypermethylation, resulting in silencing gene activity¹⁰⁹. In the context of PCOS and EDCs exposure, the shift toward ER α -only expression may be included in a pro-estrogenic milieu granted by abundant adipose tissue (PCOS), xenoestrogens (EDCs) and a spontaneous receptor selection already undertaken by the ovarian neoplasm striving to grow. Many xenoestrogen can only exert a modest estrogenic effect, but they have greater bioavailability (almost complete) when compared to endogenous oestrogens. Moreover, xenoestrogens of different kind can work synergically with one another and with endogenous oestrogens¹⁵⁹⁻¹⁶².

CONCLUSION

In conclusion, the molecular similarity of environmental toxins to endogenous steroids hormones may lead to hormonal and metabolic disruption, favouring both PCOS and OC development. In addition, patients diagnosed with PCOS who are exposed to environmental toxins may be at higher risk of OC. Several mechanisms seem to play a role in this process:

- hyperandrogenism, a key feature of PCOS patients, seems to be further boosted by EDCs exposure, finally promoting cells proliferation and in the end tumour mass growth;
- some EDCs show estrogenic properties and especially in overweight/obese patients with PCOS (where the hypertrophic fatty tissue represents the production site of estrone and the conversion site of A to E) may promote tumour proliferation by binding to ERα present in more than half of all ovarian tumours;
- the low-grade systemic inflammation typical of the metabolic syndrome of PCOS women is bolstered by EDCs by two major mechanisms, oxidative stress and insulin-resistance, which may lead to DNA strands damage and impaired steroidogenesis, finally providing a proliferative stimulus;
- EDCs may be responsible for epigenetic modifications, in particular for the shift toward ERαonly expression, which may intensify the pro-estrogenic milieu granted by abundant adipose tissue, typical of PCOS women, favouring carcinogenesis.

From a public health perspective a possible association between PCOS, EDCs exposure and OC would be highly important. Indeed, it would allow to recognise women at risk, to monitor them properly and to offer preventive treatment(s). This lack of recognition further delays the diagnosis of pre-malignant/malignant ovarian diseases (which is already delayed in OC because of the nature of the cancer itself). However important, the association between PCOS, EDCs and OC remains challenging for several reasons:

- PCOS still has a poorly understood aetiology and it is often encased in patient with other metabolic abnormalities;
- the list of EDCs is growing, and a single EDC can exert different effects according to:
 - timing of exposure (pre-natal life/post natal/pre-pubertal...),
 - length of exposure,
 - total dose,

, ccei

- latency between exposure and effects,
- possible synergic effects with other molecules.

Further research, although difficult, is needed in this field. The identification of possible cancerogenic molecules would allow to prevent further diffusion of these substances in the environment, or at least to provide adequate information to the population considered at risk.

References

- 1. Koch CA, Diamanti-Kandarakis E. Introduction to endocrine disrupting chemicals: is it time to act? Rev Endocrine Metab Disord 2015;16:269–70
- 2. Kavlock RJ, Daston GP, DeRosa C, et al. Research needs for the risk assessment of health and environmental effects of endocrine disruptors: a report of the U.S. EPA-sponsored workshop. Environ Health Perspect 1996;104:715–40
- 3. Hong H, Branham WS, Ng HW, et al. Human sex hormone–binding globulin binding affinities of 125 structurally diverse chemicals and comparison with their binding to androgen receptor, estrogen receptor, and alpha-fetoprotein. Toxicol Sci 2015;143:333–48
- 4. Sheikh IA, Turki RF, Abuzenadah AM, et al. Endocrine disruption: computational perspectives on human sex hormone–binding globulin and phthalate plasticizers. PloS One 2016;11:e0151444
- 5. Bergman A, Heindel JJ, Kasten T, et al. The impact of endocrine disruption: a consensus statement on the state of the science. Environ Health Perspect 2013;121:A104–6.
- 6. Rutkowska AZ, Diamanti-Kandarakis E. Polycystic ovary syndrome and environmental toxins. Fertil Steril. 2016;106:948-58
- 7. Rehan M, Ahmad E, Sheikh IA, et al. Androgen and progesterone receptors are targets for bisphenol A (BPA), 4-Methyl-2,4-bis-(P-Hydroxyphenyl)Pent-1-Ene—a potent metabolite of BPA, and 4-Tert-Octylphenol: a computational insight. PloS One 2015;10:e0138438
- Koschinsky T, He CJ, Mitsuhashi T, et al. Orally absorbed reactive glycation products (glycotoxins): an environmental risk factor in diabetic nephropathy. Proc Nat Acad Sci U S A 1997;94:6474–9
- 9. Bierhaus A, Stern DM, Nawroth PP. RAGE in inflammation: a new therapeutic target? Curr Opin Investig Drugs 2006;7:985–91
- Schiekofer S, Andrassy M, Chen J, et al. Acute hyperglycemia causes intracellular formation of CML and activation of ras, p42/44MAPK, and nuclear factor kappaB in PBMCs. Diabetes 2003;52:621–33
- 11. Palimeri S, Palioura E, Diamanti-Kandarakis E. Current perspectives on the health risks associated with the consumption of advanced glycation end products: recommendations for dietary management. Diabetes Metab Syndr Obes 2015;8:415–26
- 12. Soave I, Caserta D, Wenger JM, et al. Environment and Endometriosis: a toxic relationship. Eur Rev Med Pharmacol Sci 2015;19:1964-72
- 13. Mallozzi M, Leone C, Manurita F, et al. Endocrine Disrupting Chemicals and Endometrial Cancer: An Overview of Recent Laboratory Evidence and Epidemiological Studies. Int J Environ Res Public Health 2017 Mar 22;14(3).
- 14. Lee HR, Jeung EB, Cho MH, et al. Molecular mechanism (s) of endocrine-disrupting chemicals and their potent oestrogenicity in diverse cells and tissues that express oestrogen receptors. J Cell Mol Med 2013;17:1–11
- 15. Crews D, McLachlan JA. Epigenetics, evolution, endocrine disruption, health, and disease. Endocrinology 2006;147:S4–10
- 16. Scsukova S, Rollerova E, Bujnakova Mlynarcikova A. Impact of endocrine disrupting chemicals on onset and development of female reproductive disorders and hormone-related cancer. Reprod Biol 2016;16:243-54
- Neier K, Marchlewicz EH, Dolinoy DC, et al. Assessing human health risk to endocrine disrupting chemicals: a focus on prenatal exposures and oxidative stress. Endocrine Disruptors 2015;3:e1069916
- 18. Vaiserman A. Early-life exposure to endocrine disrupting chemicals and later- life health outcomes: an epigenetic bridge? Aging Dis 2014;5:419–29
- 19. Casati L, Sendra R, Sibilia V, et al. Endocrine disrupters: the new players able to affect the epigenome. Front Cell Dev Biol 2015;3:37
- 20. Diamanti-Kandarakis E. Polycystic ovarian syndrome: pathophysiology, molecular aspects and clinical implications. Exp Rev Mol Med 2008;10:e3
- 21. Azziz R, Woods KS, Reyna R, et al. The prevalence and features of the polycystic ovary

syndrome in an unselected population. J Clin Endocrinol Metab 2004;89:2745-9

- 22. Boyle J, Teede HJ. Polycystic ovary syndrome:an update. Aust Fam Physician 2012;41:752–6
- 23. Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). Hum Reprod 2004;19:41-7
- 24. Norman RJ, Dewailly D, Legro RS, et al. Polycystic ovary syndrome. Lancet 2007;370:685–97
- 25. Legro RS, Kunselman AR, Demers L, et al. Elevated dehydroepiandrosterone sulfate levels as the reproductive phenotype in the brothers of women with polycystic ovary syndrome. J Clin Endocrinol Metab 2002;87:2134
- 26. Livadas S, Pappas C, Karachalios A, et al. Prevalence and impact of hyperandrogenemia in 1,218 women with polycystic ovary syndrome. Endocrine 2014;47:631–8
- 27. Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. Endocrine Rev 2012;33:981–1030
- 28. Rachon D, Teede H. Ovarian function and obesity—interrelationship, impact on women's reproductive lifespan and treatment options. Mol Cell Endocrinol 2010;316:172–9
- 29. Lo JC, Feigenbaum SL, Yang J, et al. Epidemiology and adverse cardiovascular risk profile of diagnosed polycystic ovary syndrome. J Clin Endocrinol Metab 2006;91:1357
- 30. Wild RA, Carmina E, Diamanti-Kandarakis E, et al. Assessment of cardiovascular risk and prevention of cardiovascular disease in women with the polycystic ovary syndrome: a consensus statement by the Androgen Excess and Polycystic Ovary Syndrome (AE-PCOS) Society. J Clin Endocrinol Metab 201;95:2038
- 31. Coburn SB, Bray F, Sherman ME, et al. International patterns and trends in ovarian cancer incidence, overall and by histologic subtype. Int J Cancer 2017;140:2451-60
- 32. Torre LA, Bray F, Siegel RL, et al. Global cancer statistics, 2012. CA Cancer J Clin 2015;65:87-108
- 33. Teng Z, Han R, Huang X, et al. Increase of incidence and mortality of ovarian cancer during 2003-2012 in Jiangsu Province, China. Front Public Health 2016;4:146
- 34. Stewart SL. Ovarian cancer incidence: current and comprehensive statistics. In: Samir F, ed. Ovarian Cancer-Clinical and Therapeutic Perspectives. Europe: InTech; 2012: 1-15
- 35. Gao H, Yang BJ, Li N, Feng LM, Shi XY, Zhao WH, Liu SJ. Bisphenol A and hormone-associated cancers: current progress and perspectives. Medicine (Baltimore) 2015;94:e211
- 36. Vaughan S, Coward JI, Bast RC Jr, et al. Rethinking ovarian cancer: recommendations for improving outcomes. Nat Rev Cancer 2011;11:719-25
- 37. Matulonis UA, Sood AK, Fallowfield L, et al. Ovarian cancer. Nat Rev Dis Primers 2016;2:16061
- 38. Karnezis AN, Cho KR, Gilks CB, et al. The disparate origins of ovarian cancers: pathogenesis and prevention strategies. Nat Rev Cancer 2017;17:65-74
- 39. Kurman RJ, Shih IM. The dualistic model of ovarian carcinogenesis: revisited, revised, and expanded. Am J Pathol 2016;186:733-47
- 40. Nezhat FR, Apostol R, Nezhat C, et al. New insights in the pathophysiology of ovarian cancer and implications for screening and prevention. Am J Obstet Gynecol 2015;213:262-7
- 41. Samtani R, Sharma N, Garg D. Effects of Endocrine-Disrupting Chemicals and Epigenetic Modifications in Ovarian Cancer: A Review. Reprod Sci 2018;25:7-18
- 42. Chakravarthi BV, Nepal S, Varambally S. Genomic and epigenomic alterations in cancer. Am J Pathol 2016;186:1724-35
- 43. Caron O. Oncogenetics in the management of ovarian cancer: state of the art. Gynecol Obstet Fertil 2015:43:335-7
- 44. Streff H, Profato J, Ye Y, et al. Cancer incidence in first and second degree relatives of BRCA1 and BRCA2 mutation carriers. Oncologist 2016;21:869-74
- 45. Takeuchi T, Tsutsumi O, Ikezuki Y, et al. Positive relationship between androgen and the endocrine disruptor, bisphenol A, in normal women and women with ovarian dysfunction. Endocr J 2004;51:165–9
- 46. Kandaraki E, Chatzigeorgiou A, Livadas S, et al. Endocrine disruptors and polycystic ovary syndrome (PCOS): elevated serum levels of bisphenol A in women with PCOS. J Clin Endocrinol Metab 2011;96:E480–4

- 47. Akin L, Kendirci M, Narin F, et al. The endocrine disruptor bisphenol A may play a role in the aetiopathogenesis of polycystic ovary syndrome in adolescent girls. Acta Paediatr 2015;104:e171–7
- 48. Vagi SJ, Azziz-Baumgartner E, Sjodin A, et al. Exploring the potential association between brominated diphenyl ethers, polychlorinated biphenyls, organochlorine pesticides, perfluorinated compounds, phthalates, and bisphenol A in polycystic ovary syndrome: a casecontrol study. BMC Endocr Disord 2014;14:86
- 49. Yang Q, Zhao Y, Qiu X, et al. Association of serum levels of typical organic pollutants with polycystic ovary syndrome (PCOS): a case control study. Hum Reprod 2015;30:1964–73
- 50. Diamanti-Kandarakis E, Katsikis I, Piperi C, et al. Increased serum advanced glycation endproducts is a distinct finding in lean women with polycystic ovary syndrome (PCOS). Clin Endocrinol (Oxf) 2008;69:634–41
- 51. Diamanti-Kandarakis E, Piperi C, Patsouris E, et al. Immunohistochemical localization of advanced glycation end-products (AGEs) and their receptor (RAGE) in polycystic and normal ovaries. Histochem Cell Biol 2007;127:581–9
- 52. Patel S, Zhou C, Rattan S, et al. Effects of endocrine-disrupting chemicalson the ovary. Biol Reprod 2015;93:20
- 53. Zhou W, Liu J, Liao L, et al. Effect of bisphenol A on steroid hormone production in rat ovarian theca-interstitial and granulosa cells. Mol Cell Endocrinol 2008;283:12–8
- 54. Gunnarsson D, Leffler P, Ekwurtzel E, et al. Mono-(2-ethylhexyl) phthalate stimulates basal steroidogenesis by a cAMP-independent mechanism in mouse gonadal cells of both sexes. Reproduction 2008;135:693–703
- 55. Kuiper GG, Lemmen JG, Carlsson B, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. Endocrinology 1998;139:4252–63
- 56. Thomas P, Dong J. Binding and activation of the seven-transmembrane estrogen receptor GPR30 by environmental estrogens: a potential novel mechanism of endocrine disruption. J Steroid Biochem Mol Biol 2006;102:175–9
- 57. Hanioka N, Jinno H, Nishimura T, et al. Suppression of male-specific cytochrome P450 isoforms by bisphenol A in rat liver. Arch Toxicol 1998;72:387–94
- 58. Dechaud H, Ravard C, Claustrat F, et al. Xenoestrogen interaction with human sex hormonebinding globulin (hSHBG). Steroids 1999;64:328–34
- 59. Takeuchi T, Tsutsumi O, Ikezuki Y, et al. Elevated serum bisphenol A levels under hyperandrogenic conditions may be caused by decreased UDP-glucuronosyltransferase activity. Endocr J 2006;53:485–91
- 60. Lovekamp-Swan T, Davis BJ. Mechanisms of phthalate ester toxicity in the female reproductive system. Environ Health Perspect 2003;111:139–45
- 61. Papalou O, Victor VM, Diamanti-Kandarakis E. Oxidative stress in polycystic ovary syndrome. Curr Pharmaceut Des 2016;22:2709–22
- 62. Tatone C, Amicarelli F, Carbone MC, et al. Cellular and molecular aspects of ovarian follicle ageing. HumReprod Update 2008;14:131–42
- 63. Ben-Jonathan N, Hugo ER, Brandebourg TD. Effects of bisphenol A on adipokine release from human adipose tissue: Implications for the metabolic syndrome. Mol Cell Endocrinol 2009;304:49–54
- 64. Menale C, Grandone A, Nicolucci C, et al. Bisphenol A is associated with insulin resistance and modulates adiponectin and resistin gene expression in obese children. Pediatr Obes 2017;12:380-7
- 65. Alonso-Magdalena P, Morimoto S, Ripoll C, et al. The estrogenic effect of bisphenol A disrupts pancreatic beta-cell function in vivo and induces insulin resistance. Environ Health Perspect 2006;114:106–12
- 66. Lang IA, Galloway TS, Scarlett A, et al. Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. JAMA 2008;300:1303–10
- 67. Jia X, Wu L. Accumulation of endogenous methylglyoxal impaired insulin signaling in adipose tissue of fructose-fed rats. Mol Cell Biochem 2007;306:133–9

- 68. Diamanti-Kandarakis E, Chatzigeorgiou A, Papageorgiou E, et al. Advanced glycation endproducts and insulin signaling in granulosa cells. Exp Biol Med (Maywood) 2016;241:1438-45
- 69. Naitoh T, Kitahara M, Tsuruzoe N. Tumor necrosis factor-alpha is induced through phorbol ester—and glycated human albumin-dependent pathway in THP-1 cells. Cell Signal 2001;13:331–4
- 70. Pasquali R, Gambineri A, Pagotto U. The impact of obesity on reproduction in women with polycystic ovary syndrome. Br J Obstet Gynecol 2006;113: 148–59
- 71. Jia X, Chang T, Wilson TW, et al. Methylglyoxal mediates adipocyte proliferation by increasing phosphorylation of Akt1. PLoS One 2012;7:e36610
- 72. Vazzana N, Guagnano MT, Cuccurullo C, et al. Endogenous secretory RAGE in obese women: association with platelet activation and oxidative stress. J Clin Endocrinol Metab 2012;97:E1726–30
- 73. Koyama H, Shoji T, Yokoyama H, et al. Plasma level of endogenous secretory RAGE is associated with components of the metabolic syndrome and atherosclerosis. Arterioscler Thromb Vasc Biol 2005;25:2587–93
- 74. Merhi Z, McGee EA, Buyuk E. Role of advanced glycation end-products in obesity-related ovarian dysfunction. Minerva Endocrinol 2014;39:167–74
- 75. Holtcamp W. Obesogens: an environmental link to obesity. Environ Health Perspect 2012;120:a62–8
- 76. Maradonna F, Evangelisti M, Gioacchini G, et al. Assay of vtg, ERs and PPARs as endpoint for the rapid in vitro screening of the harmful effect of Di-(2-ethylhexyl)-phthalate (DEHP) and phthalic acid (PA) in zebrafish primary hepatocyte cultures. Toxicol In Vitro 2013;27:84–91
- 77. Somm E, Schwitzgebel VM, Toulotte A, et al. Perinatal exposure to bisphenol a alters early adipogenesis in the rat. Environ Health Perspect 2009;117:1549–55
- 78. Masuno H, Iwanami J, Kidani T, et al. Bisphenol A accelerates terminal differentiation of 3T3-L1 cells into adipocytes through the phosphatidylinositol 3-kinase pathway. Toxicol Sci 2005;84:319–27
- 79. Koutros S, Alavanja MC, Lubin JH, et al. An update of cancer incidence in the agricultural health study. J Occup Environ Med 2010;52:1098-105
- 80. Young HA, Mills PK, Riordan DG, et al. Triazine herbicides and epithelial ovarian cancer risk in central California. J Occup Environ Med 2005;47:1148-56
- 81. Lerro CC, Koutros S, Andreotti G, et al. Organophosphate insecticide use and cancer incidence among spouses of pesticide applicators in the agricultural health study. Occup Environ Med 2015;72:736-44
- 82. Kim JY, Yi BR, Go RE, et al. Methoxychlor and triclosan stimulates ovarian cancer growth by regulating cell cycle- and apoptosis-related genes via an estrogen receptor-dependent pathway. Environ Toxicol Pharmacol 2014;37:1264-74
- 83. Park SH, Kim KY, An BS, et al. Cell growth of ovarian cancer cells is stimulated by xenoestrogens through an estrogendependent pathway, but their stimulation of cell growth appears not to be involved in the activation of the mitogenactivated protein kinases ERK-1 and p38. J Reprod Dev 2009;55:23-9
- 84. Adewale HB, Jefferson WN, Newbold RR, et al. Neonatal bisphenol-A exposure alters rat reproductive development and ovarian morphology without impairing activation of gonadotropin-releasing hormone neurons. Biol Reprod 2009;81:690-9
- 85. Ptak A, Wrobel A, Gregoraszczuk EL. Effect of bisphenol-A on the expression of selected genes involved in cell cycle and apoptosis in the OVCAR-3 cell line. Toxicol Lett 2011;202:30-5
- 86. Patel S, Zhou C, Rattan S, et al. The effects of endocrine disrupting chemicals on the ovary. Bio Reprod 2015;93:20
- 87. Shi LF, Wu Y, Li CY. Hormone therapy and risk of ovarian cancer in postmenopausal women: a systematic review and meta-analysis. Menopause 2016;23:417-24
- Tsilidis KK, Allen NE, Key TJ, et al. Menopausal hormone therapy and risk of ovarian cancer in the European prospective investigation into cancer and nutrition. Cancer Causes Control 2011;22:1075-84

- 89. Jordan SJ, Wilson LF, Nagle CM, et al. Cancers in Australia in 2010 attributable to and prevented by the use of menopausal hormone therapy. Aust NZ J Public Health 2015;39:434-40
- 90. Collaborative group on epidemiological studies of ovarian cancer, Beral V, Gaitskell K, et al. Menopausal hormone use and ovarian cancer risk: individual participant meta-analysis of 52 epidemiological studies. Lancet 2015;385:1835-42
- 91. Costa EMF, Spritzer PM, Hohl A, et al. Effects of endocrine disruptors in the development of the female reproductive tract. Arq Bras Endocrinol Metab 2014;58:153-61
- 92. Park MA, Hwang KA, Lee HR, et al. Benzophenone-1 stimulated the growth of BG-1 ovarian cancer cells by cell cycle regulation via an estrogen receptor alphamediated signaling pathway in cellular and xenograft mouse models. Toxicology 2013;305:41-8
- 93. Kojima H, Katsura E, Takeuchi S, et al. Screening for estrogen and androgen receptor activities in 200 pesticides by in vitro reporter gene assays using Chinese hamster ovary cells. Environ Health Perspect 2004;112:524-31
- 94. Cardenas C, Alvero AB, Yun BS, et al. Redefining the origin and evolution of ovarian cancer: a hormonal connection. Endocr Relat Cancer 2016;23:R411-22
- 95. Gore AC. Organochlorine pesticides directly regulate gonadotropin-releasing hormone gene expression and biosynthesis in the GT1-7 hypothalamic cell line. Mol Cell Endocrinol 2002;192:157-70
- 96. Ventura C, Nieto MR, Bourguignon N, et al. Pesticide chlorpyrifos acts as an endocrine disruptor in adult rats causing changes in mammary gland and hormonal balance. J Steroid Biochem Mol Bio 2016;156:1-9
- 97. Munier M, Grouleff J, Gourdin L, et al. In vitro effects of the endocrine disruptor p, p0- DDT on human follitropin receptor. Environ Health Perspect 2016;124(7):991-999.
- 98. Gharwan H, Buch KP, Annunziata CM. The role of reproductive hormones in epithelial ovarian carcinogenesis. Endocr Relat Cancer 2015;22:R339-63
- 99. Olsen CM, Green AC, Nagle CM, et al. Epithelial ovarian cancer: testing the 'androgens hypothesis.' Endocr Relat Cancer 2008;15:1061-68
- 100. Danforth KN, Eliassen AH, Tworoger SS, et al. The association of plasma androgen levels with breast, ovarian and endometrial cancer risk factors among postmenopausal women. Int J Cancer 2010;126:199-207
- 101. Trisomboon J, Li C, Suzuki A, et al. Nitro-3- phenylphenol has both androgenic and antiandrogenic-like effects in rats. J Reprod Dev 2015;61:134-7
- 102. Yang Q, Diamond MP, Al-Hendy A. Early life adverse environmental exposures increase the risk of uterine fibroid development: role of epigenetic regulation. Front Pharmacol 2016;7:40
- 103. Jirtle RL, Skinner MK. Environmental epigenomics and disease susceptibility. Nat Rev Genet 2007;8:253-62
- 104. Hamidi T, Singh AK, Chen T. Genetic alterations of DNA methylation machinery in human diseases. Epigenomics 2015;7:247-65
- 105. Taberlay PC, Jones PA. DNA methylation and cancer. In: Gasser SM, Li E, eds. Epigenetics and Disease, Pharmaceutical Opportunities. Switzerland: Springer Basel AG; 2011:1-5
- 106. Knower KC, To SQ, Leung Y, et al. Endocrine disruption of the epigenome: a breast cancer link. Endocr Relat Cancer 2014;21:T33-55
- 107. Romagnolo DF, Daniels KD, Grunwald JT, et al. Epigenetics of breast cancer: modifying role of environmental and bioactive food compounds. Mol Nutr Food Res 2016;60:1310-29
- 108. Uzumcu M, Zama AM, Oruc E. Epigenetic mechanisms in the actions of endocrine-disrupting chemicals: gonadal effects and role in female reproduction. Reprod Domest Anim 2012;47:338-47
- 109. Orphanides G, Reinberg D. A unified theory of gene expression. Cell 2002;108:439-51
- 110. Zama AM, Uzumcu M. Fetal and neonatal exposure to the endocrine disruptor methoxychlor causes epigenetic alterations in adult ovarian genes. Endocrinology 2009;150:4681-91
- 111. LegroR. In Diamanti-Kandarakis E,Nestler J,PanidisD,Pasquali R, (eds). Insulin resistance and polycystic ovarian syndrome 2007. Humana Press, Totowa, NJ, 335-48
- 112. Schildkraut JM, Schwingl PJ, Bastos E, et al. Epithelial ovarian cancer risk among women with

polycystic ovary syndrome. Obstet Gynecol 1996;88:554-9

- 113. Risch HA. Hormonal Etiology of Epithelial Ovarian Cancer, With a Hypothesis Concerning the Role of Androgens and Progesterone. J Natl Cancer Inst 1998;90:1774–86
- 114. Butler M, Ricciardelli C, Tilley W, et al. Androgen Receptor Protein Levels Are Significantly Reduced in Serous Ovarian Carcinomas Compared with Benign or Borderline Disease but Are Not altered by Cancer Stage or Metastatic Progression. Horm Cancer 2013;4:154-64
- 115. Schock H, Surcel H-M, Zeleniuch-Jacquotte A, et al. Early pregnancy sex steroids and maternal risk of epithelial ovarian cancer. Cancer Res 2014;74:6958-67
- 116. Rossing MA, Daling JR, Weiss NS, et al. Ovarian Tumors in a Cohort of Infertile Women. N Engl J Med 1994;331:771–6
- Brinton L, Moghissi K, Westhoff C, et al. Cancer risk among infertile women with androgen excess or menstrual disorders (including polycystic ovary syndrome). Fertil Steril 2010;94:1787–92
- 118. Bodmer M, Becker C, Meier C, et al. Use of metformin and the risk of ovarian cancer: A case– control analysis. Gynecol Oncol 2011; 23:200–4
- 119. Barry JA, Azizia MM, Hardiman PJ. Risk of endometrial, ovarian and breast cancer in women with polycystic ovary syndrome: a systematic review and meta-analysis. Hum Reprod Update 2014;20:748–58
- 120. Shen C-C, Yang AC, Hung J-H, et al. A Nationwide Population-Based Retrospective Cohort Study of the Risk of Uterine, Ovarian and Breast Cancer in Women With Polycystic Ovary Syndrome. Oncologist 2015;20:45–9
- 121. Gottschau M, Kjaer S, Jensen A, et al. Risk of cancer among women with polycystic ovary syndrome: a Danish cohort study. Gynecol Oncol 2015;136:99–103
- 122. Harris H, Titus L, Cramer D, et al. Long and irregular menstrual cycles, polycystic ovary syndrome, and ovarian cancer risk in a population-based case–control study. Int J Cancer 2017;140:285-91
- 123. Ding DC, Chen W, Wang JH, et al. Association between polycystic ovarian syndrome and endometrial, ovarian, and breast cancer: A population-based cohort study in Taiwan. Medicine (Baltimore) 2018;97:e12608
- 124. Harris HR, Terry KL. Polycystic ovary syndrome and risk of endometrial, ovarian, and breast cancer: a systematic review. Fertil Res Pract 2016;2:14.
- 125. Lim SS, Davies MJ, Norman RJ, et al. Overweight, obesity and central obesity in women with polycystic ovary syndrome: a systematic review and meta-analysis. Hum Reprod Update 2012;18:618–37
- 126. Shikata K, Ninomiya T, Kiyohara Y. Diabetes mellitus and cancer risk: Review of the epidemiological evidence. Cancer Sci 2013;104:9–14
- 127. Giovannucci E. Metabolic syndrome, hyperinsulinemia, and colon cancer: a review. Am J Clin Nutr 2007;86:s836–42
- 128. Fathalla M. Incessant ovulation a factor in ovarian neoplasia? Lancet 1971;2:163
- 129. Rocca M, Venturella R, Mocciaro R, et al. Polycystic ovary syndrome: chemical pharmacotherapy. Expert Opin Pharmocother 2015;16:1369–93
- 130. Pollak MN. Investigating Metformin for Cancer Prevention and Treatment: The End of the Beginning. Cancer Discov 2012;2:778–90
- 131. Rattan R, Giri S, Hartmann LC, et al. Metformin attenuates ovarian cancer cell growth in an AMP-kinase dispensable manner. J Cell Mol Med 2011;15:166–78
- 132. Rattan R, Graham R, Maquire J, et al. Metformin suppresses ovarian cancer growth and metastasis with enhancement of cisplatin cytotoxicity in vivo. Neoplasia 2011;13:483–91
- Blomquist CH, Bonenfant M, McGinley DM, et al. Androgenic and estrogenic 17bhydroxysteroid dehydrogenase/17-ketosteroid reductase in human ovarian epithelial tumors: evidence for the type 1, 2 and 5 isoforms. J Steroid Biochem Mol Biol 2002;81:343-51
- 134. Edmondson RJ, Monaghan JM, Davies. The human ovarian surface epithelium is an androgen responsive tissue. Br J Cancer 2002;86:879-85

- 135. Ilekis JV, Connor JP, Prins GS, et al. Expression of epidermal growth factor and androgen receptors in ovarian cancer. Gynecol Oncol 1997;66:250-4
- 136. Helzlsouer KJ, Alberg AJ, Gordon GB, et al. Serum gonadotropins and steroid hormones and the development of ovarian cancer. JAMA. 1995;274:1926-30
- 137. Yokota H, Iwano H, Endo M, et al. Glucuronidation of the environmental oestrogen bisphenol A by an isoform of UDP-glucuronosyltransferase, UGT2B1, in the rat liver. Biochem J 1999;340:405–9
- 138. Lindgren PR, Cajander S, Backstrom T, et al. Estrogen and progesterone receptors in ovarian epithelial tumors. Mol Cell Endocrinol 2004;221:97-104
- 139. Lau KM, Mok SC, Ho SM. Expression of human estrogen receptor-a and -b, progesterone receptor, and androgen receptor mRNA in normal and malignant ovarian epithelial cells. Proc Natl Acad Sci U S A 1999;96:5722-7
- 140. Bardin A, Boulle N, Lazennec G, et al. Loss of ERb expression as a common step in estrogendependent tumor progression. Endocr Relat Cancer 2004;11:537-51
- 141. Mandrelle K, Kamath MS, Bondu DJ, Chandy A, Aleyamma T, George K. Prevalence of metabolic syndrome in women with polycystic ovary syndrome attending an inferritity clinic in a tertiary care hospital in south India. J Hum Reprod Sci 2012;5:26-31
- 142. Zama AM, Uzumcu M. Epigenetic effects of endocrine-disrupting chemicals on female reproduction: an ovarian perspective. Front Neuroendocrinol 2010;31:420-39
- 143. Tomao F, Russo GL, Spinelli GP, et al. Fertility drugs, reproductive strategies and ovarian cancer risk. J Ovarian Res 2014;7:51
- 144. Choi JH, Wong AS, Huang HF, et al. Gonadotropins and ovarian cancer. Endocr Rev 2007;28:440-61
- Mangum LH, Crow JA, Stokes JV, et al. Exposure to p,p0-DDE alters macrophage reactivity and increases macrophage numbers in adipose stromal vascular fraction. Toxicol Sci 2016;150:169–77
- 146. Weisberg SP, McCann D, Desai M et al. Obesity is associated with macrophage accumulation in adipose tissue. J Clin Investig 2003;112:1796–1808
- 147. Howell G, Mangum L. Exposure to bioaccumulative organochlorine compounds alters adipogenesis, fatty acid uptake, and adipokine production in NIH3T3-L1 cells. Toxicol In Vitro 2011;25:394–402
- 148. Qian SW, Tang Y, Li X, et al.. BMP4-mediated brown fat-like changes in white adipose tissue alter glucose and energy homeostasis. Proc Natl Acad Sci USA 2013;110:E798–807
- 149. Hauner H, Petruschke T, Russ M, et al. Effects of tumour necrosis factor alpha (TNF alpha) on glucose transport and lipid metabolism of newly-differentiated human fat cells in cell culture. Diabetologia 1995;38:764–71
- 150. Lin Y, Wei J, Li Y, et al. Developmental exposure to di(2-ethylhexyl) phthalate impairs endocrine pancreas and leads to long-term adverse effects on glucose homeostasis in the rat. Am J Physiol Endocrinol Metab 2011;301:E527–38
- 151. Bodin J, Bolling AK, Becher R, et al. Transmaternal bisphenol A exposure accelerates diabetes type 1 development in NOD mice. Toxicol Sci 2014;137:311–23
- 152. Lee DH, Steffes MW, Sjodin A, et al. Low dose organochlorine pesticides and polychlorinated biphenyls predict obesity, dyslipidemia, and insulin resistance among people free of diabetes. PLoS ONE 2011;6:e15977
- 153. Kim SH, Turnbull J, Guimond S. Extracellular matrix and cell signalling: The dynamic cooperation of integrin, proteoglycan and growth factor receptor. J Endocrinol 2011;209:139–51
- 154. Shao W, Brown M. Advances in estrogen receptor biology: Prospects for improvements in targeted breast cancer therapy. Breast Cancer Res 2004;6:39–52
- 155. Brahimi-Horn MC, Chiche J, Pouyssegur J. Hypoxia and cancer. J Mol Med 2007;85:1301–7
- 156. He X, Dong X, Zou D, et al. Enantioselective Effects of o,p0-DDT on Cell Invasion and Adhesion of Breast Cancer Cells: Chirality in Cancer Development. Environ Sci Technol 2015;49:10028–37
- 157. Kuroda H, Mandai M, Konishi I, et al. Human ovarian surface epithelial (OSE) cells express LH/hCG receptors, and hCG inhibits apoptosis of OSE cells via upregulation of insulin-like

growth factor-1. Int J Cancer 2001;91:309-15

- 158. Diamanti-Kandarakis E, Bourguignon JP, Giudice LC, et al. Endocrine-disrupting chemicals: an endocrine society scientific statement. Endocr Rev 2009;30:293-342
- 159. Vandenberg LN, Colborn T, Hayes TB, et al. Hormones and endocrine-disrupting chemicals: low dose effects and non monotonic dose responses. Endocr Rev 2012; 33:378-455
- 160. Silva E, Rajapakse N, Kortenkamp A. Something from "nothing"-eight weak estrogenic chemicals combined at concentrations below NOECs produce significant mixture effects. Environ Sci Technol 2002;36:1751–6
- 161. Soto AM, Fernandez MF, Luizzi MF, et al. Developing a marker of exposure to xenoestrogen mixtures in human serum. Environ Health Perspect 1997;105:647
- 162. Rajapakse N, Silva E, Kortenkamp A. Combining xenoestrogens at levels below individual noobserved-effect concentrations dramatically enhances steroid hormone action. Environ Health Perspect 2002;110:917

AUSC k certe 4

Figure Legend

Figure 1: Mechanisms by which EDCs/AGEs exposure may favor PCOS (PPARy: peroxisome proliferatoractivated receptors; IR: insulin resistance; CVD: cardiovascular disease; T2DM: diabetes mellitus type 2; HPO axis: hypothalamic-pituitary-ovarian axis; SHGB: sex hormone-binding globulin)



Figure 2: Mechanisms by which EDCs/AGEs exposure may favor OC

